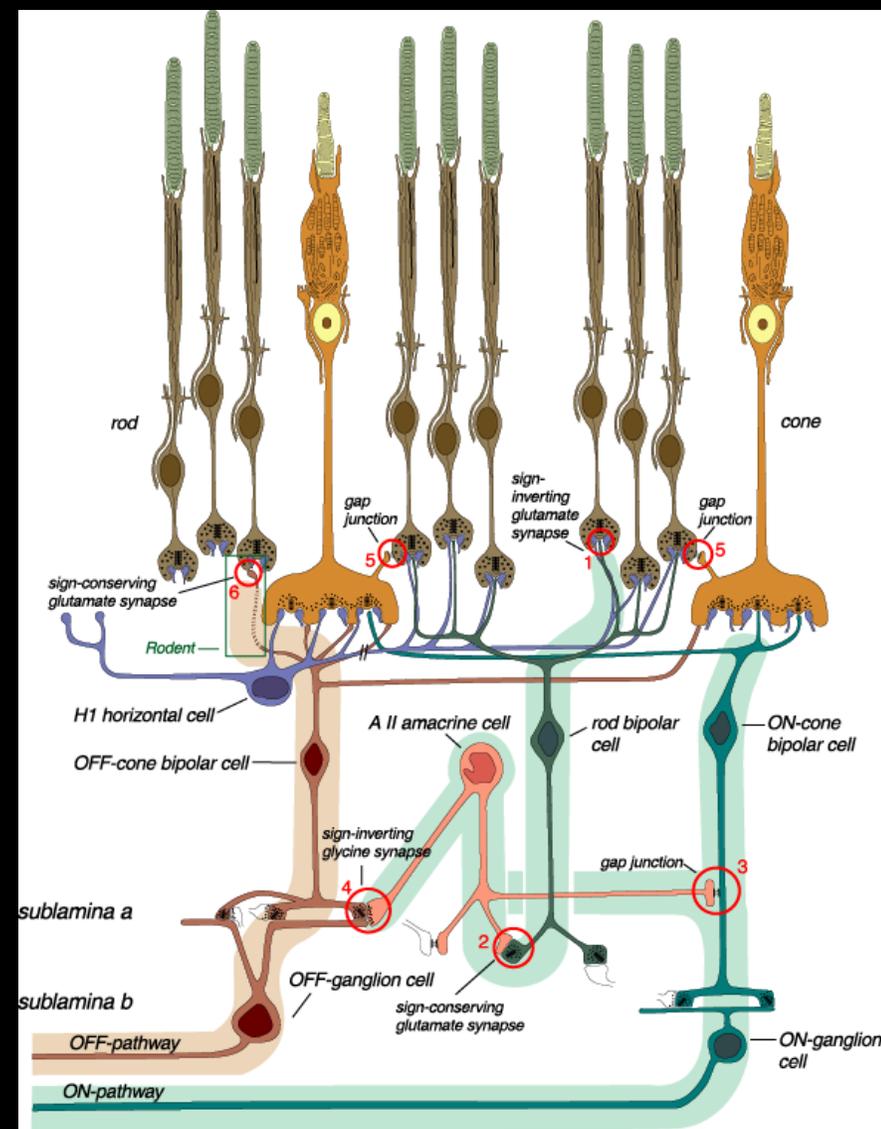


Achromatic and chromatic vision, rods and cones.

Andrew Stockman

NEUR 0017
Visual Neuroscience

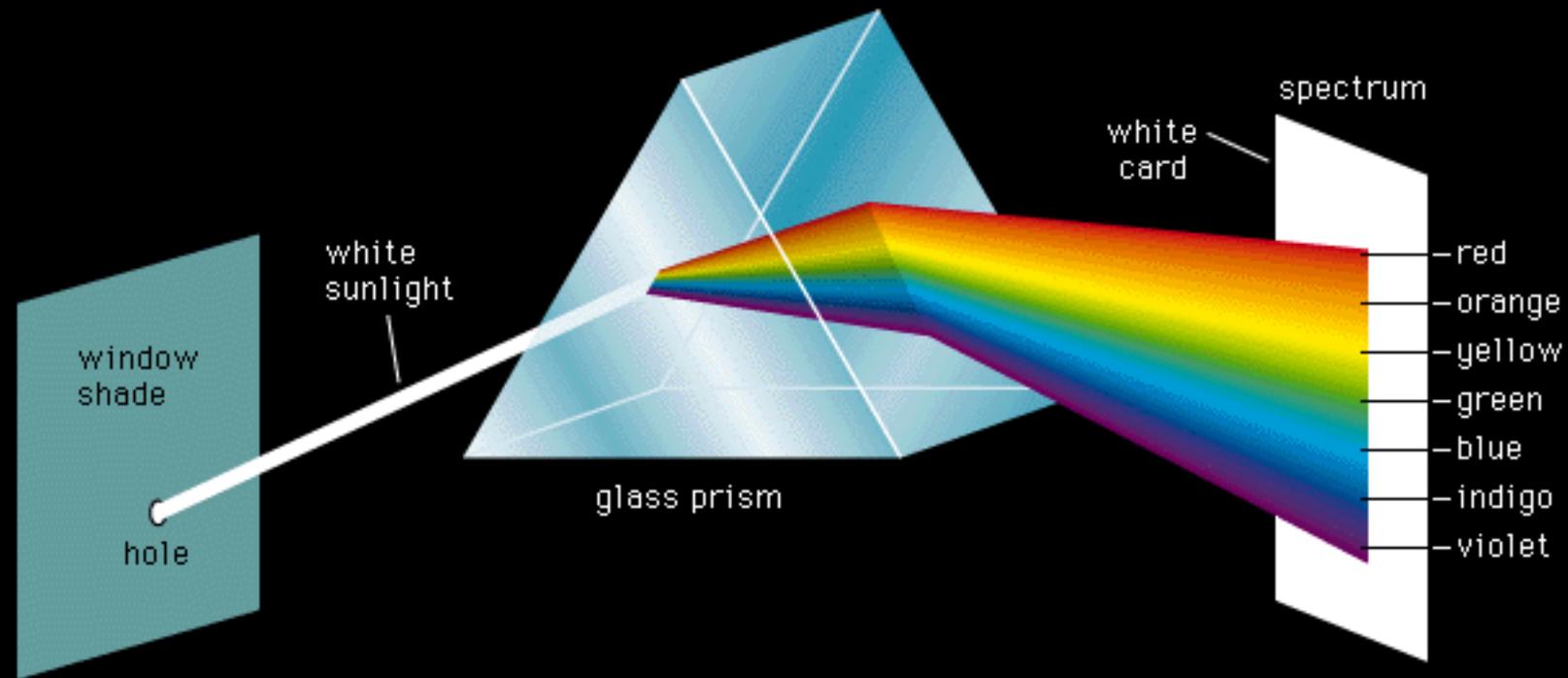


Outline

- ▶ Introduction
- ▶ Rod and cone vision
- ▶ Rod vision is achromatic
- ▶ How do we see colour with cone vision?
- ▶ Vision and visual pathways
- ▶ Achromatic and chromatic cone vision
(colour and luminance)

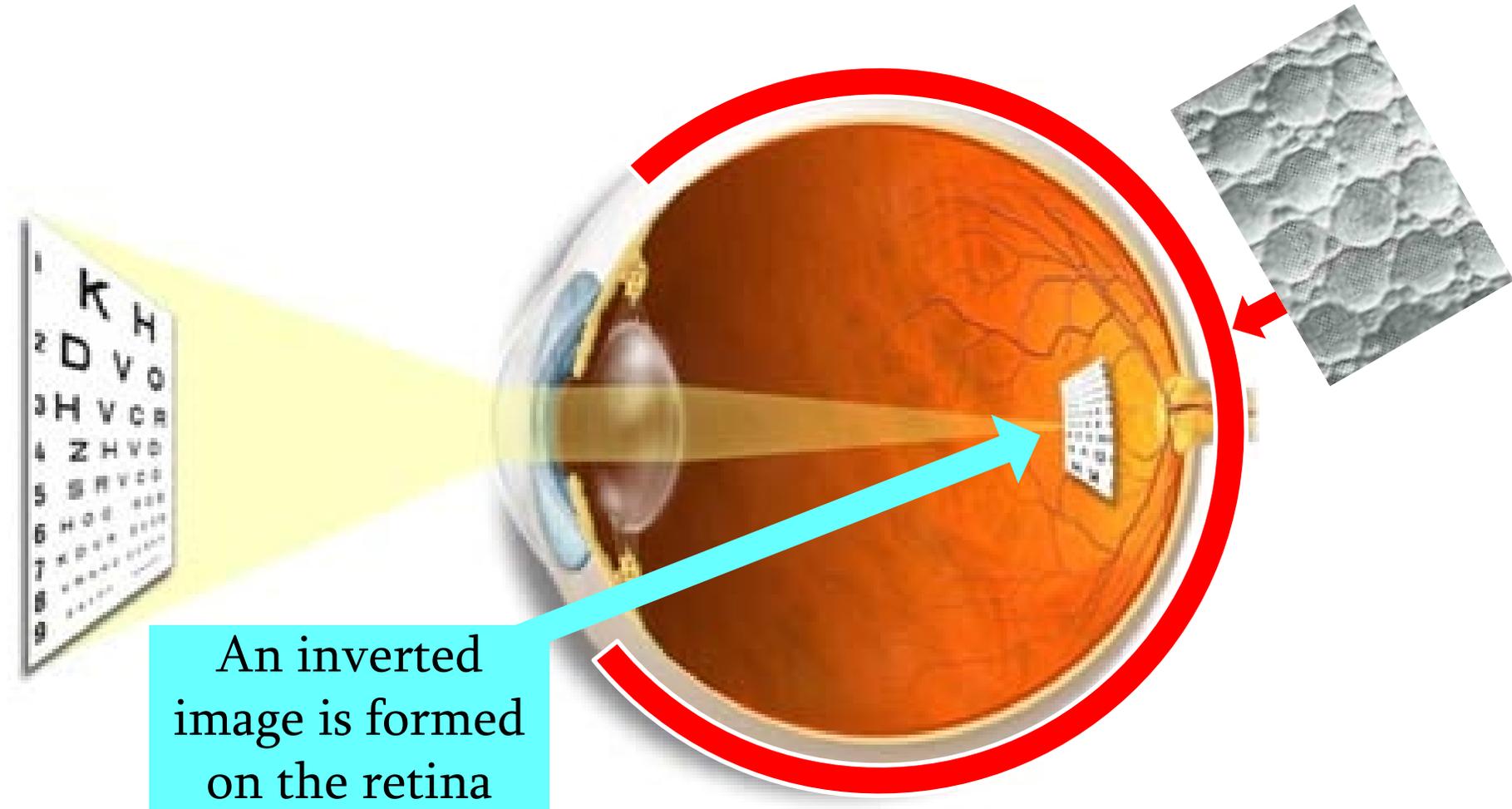
Light

400 - 700 nm is important for vision



ROD AND CONE VISION

The retina is carpeted with light-sensitive rods and cones



Rods and cones

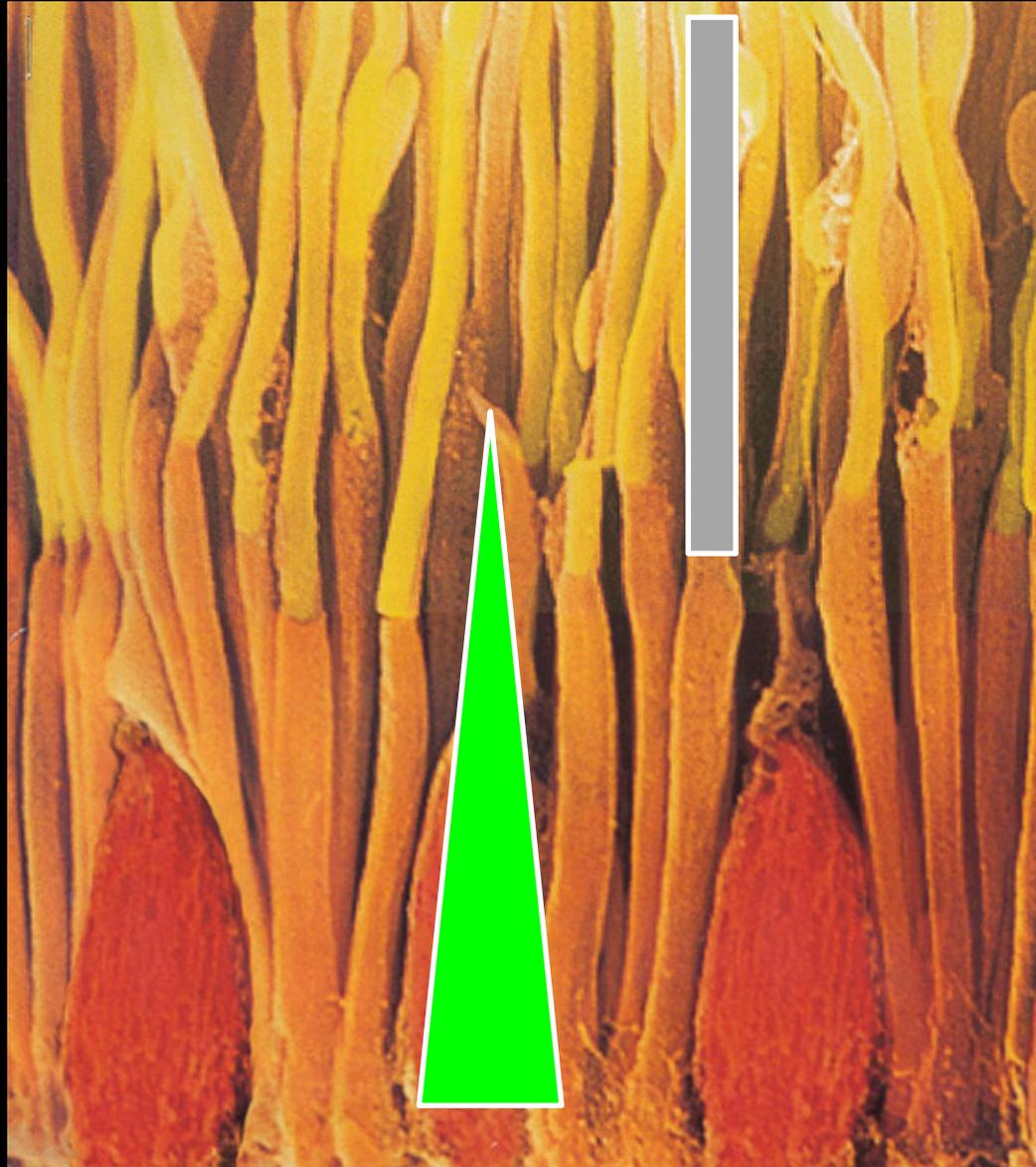
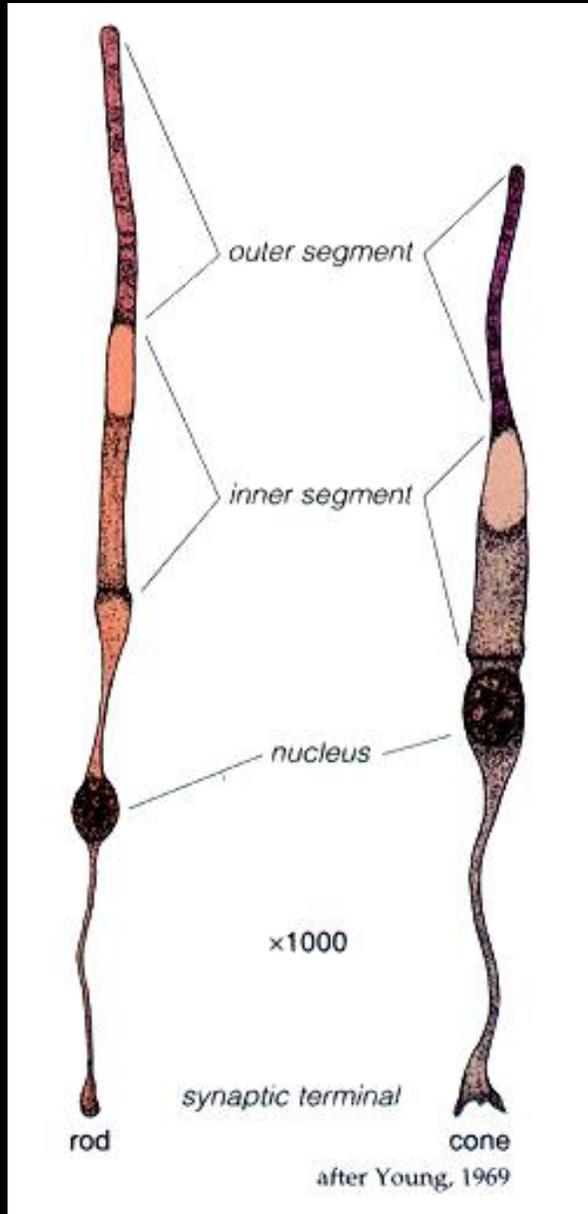


Fig1b. Scanning electron micrograph of the rods and cones of the primate retina. Image adapted from one by Ralph C. Eagle/Photo Researchers, Inc.

Human photoreceptors



Cones

- Daytime, achromatic *and* chromatic vision
- 3 types



Long-wavelength-sensitive (L) or "red" cone

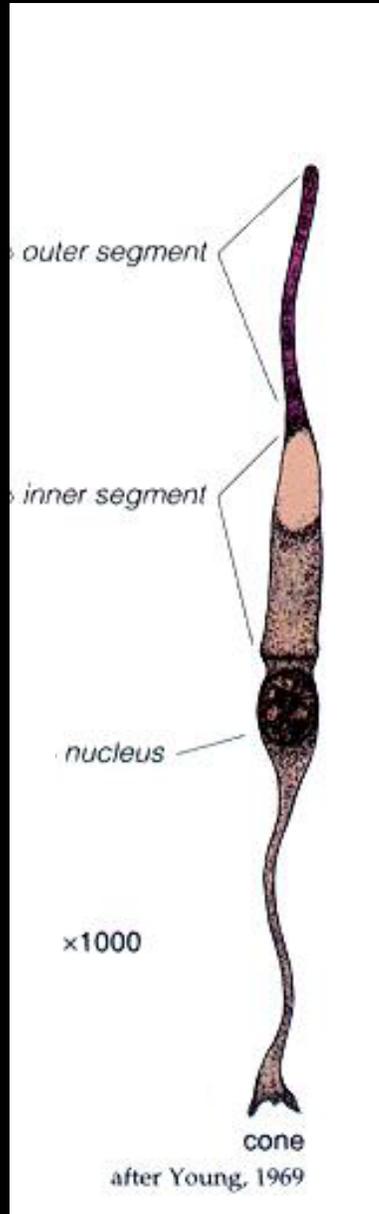


Middle-wavelength-sensitive (M) or "green" cone



Short-wavelength-sensitive (S) or "blue" cone

Human photoreceptors



Rods

- Achromatic night vision
- 1 type



Rod

Cones

- Daytime, achromatic *and* chromatic vision
- 3 types



Long-wavelength-sensitive (L) or "red" cone



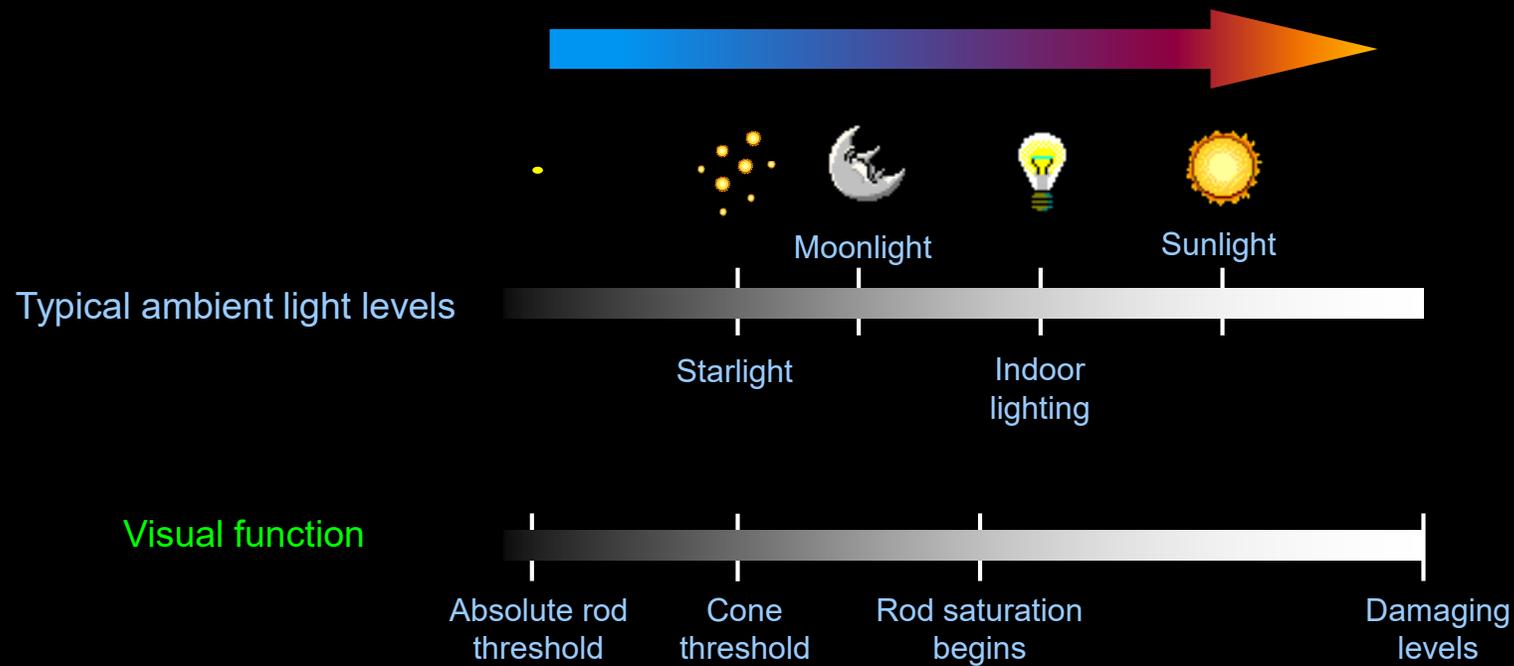
Middle-wavelength-sensitive (M) or "green" cone



Short-wavelength-sensitive (S) or "blue" cone

Why do we have rods and cones?

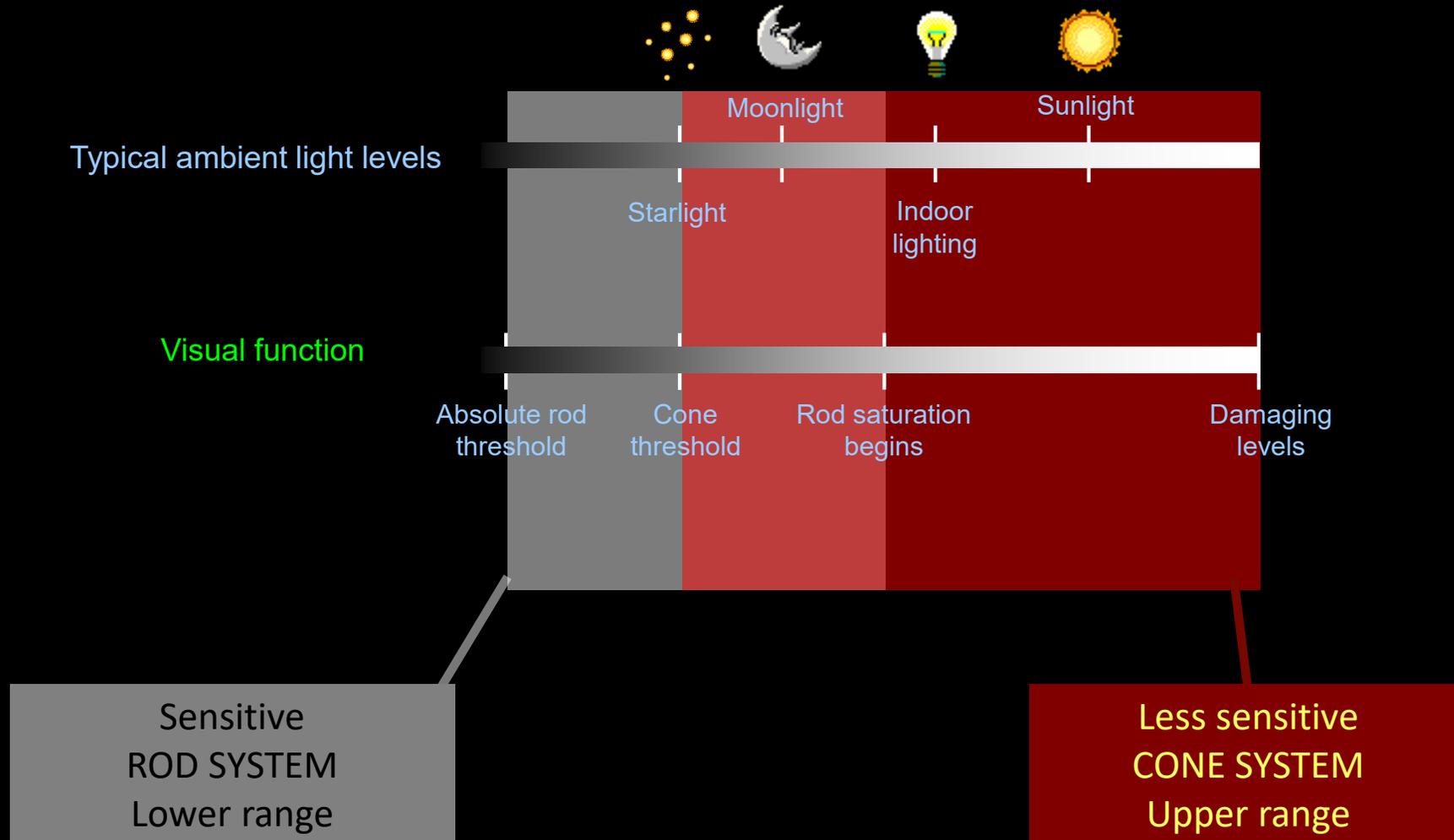
Our vision has to operate over an enormous range of 10^{12} (1,000,000,000,000) levels



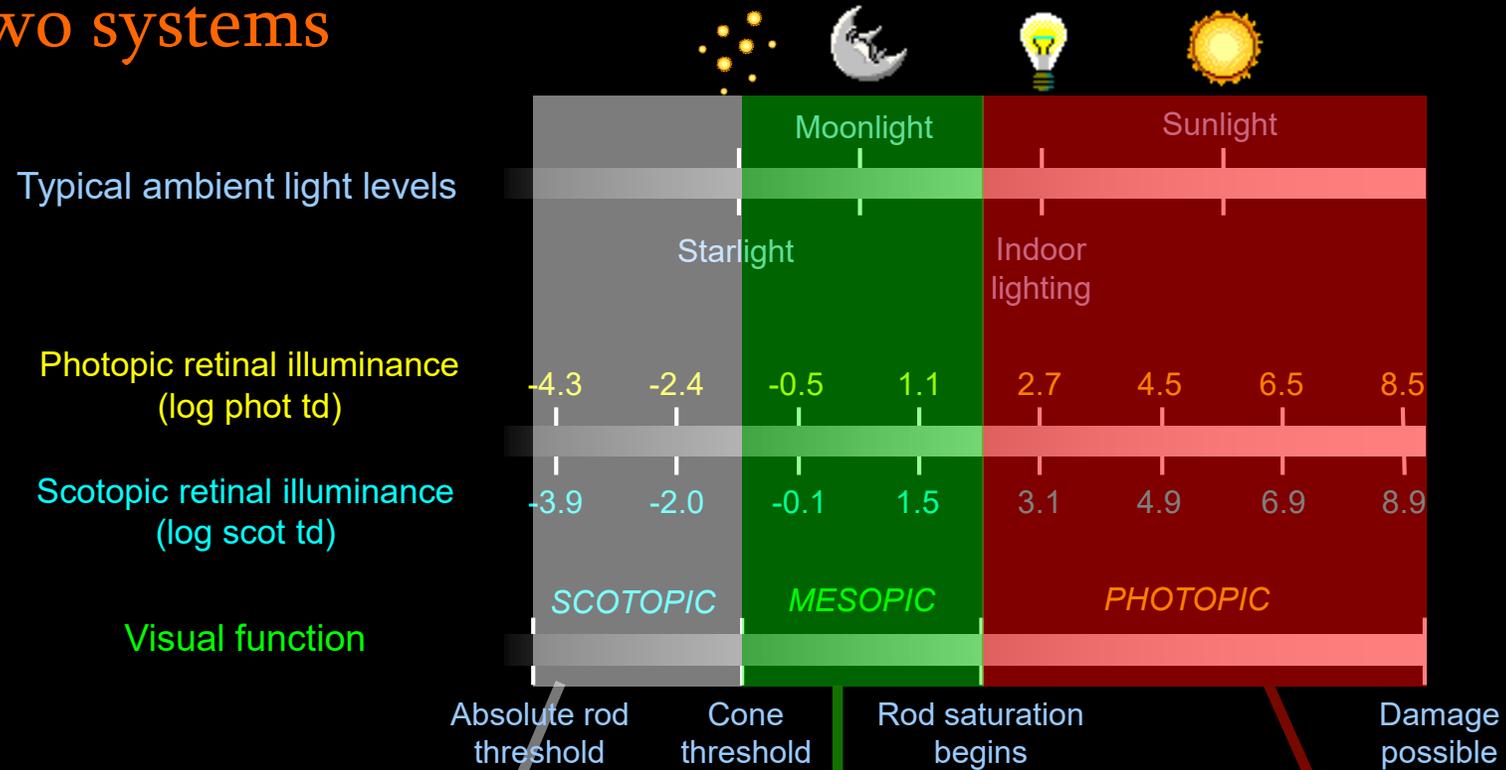
To cover that range we have two different types of photoreceptor...

Rods that are optimized for low light levels

Cones that are optimized for higher light levels



Two systems



Scotopic levels
(below cone threshold)
where rod vision
functions alone.
A range of c. $10^{3.5}$

Mesopic levels
where rod and cone
vision function
together.
A range of c. 10^3

Photopic levels
(above rod saturation)
where cone vision
functions alone.
A range of $> 10^6$

Rod vision

- Achromatic
- High sensitivity
- Poor detail and no colour



Cone vision

- Achromatic and chromatic
- Lower sensitivity
- Detail and good colour

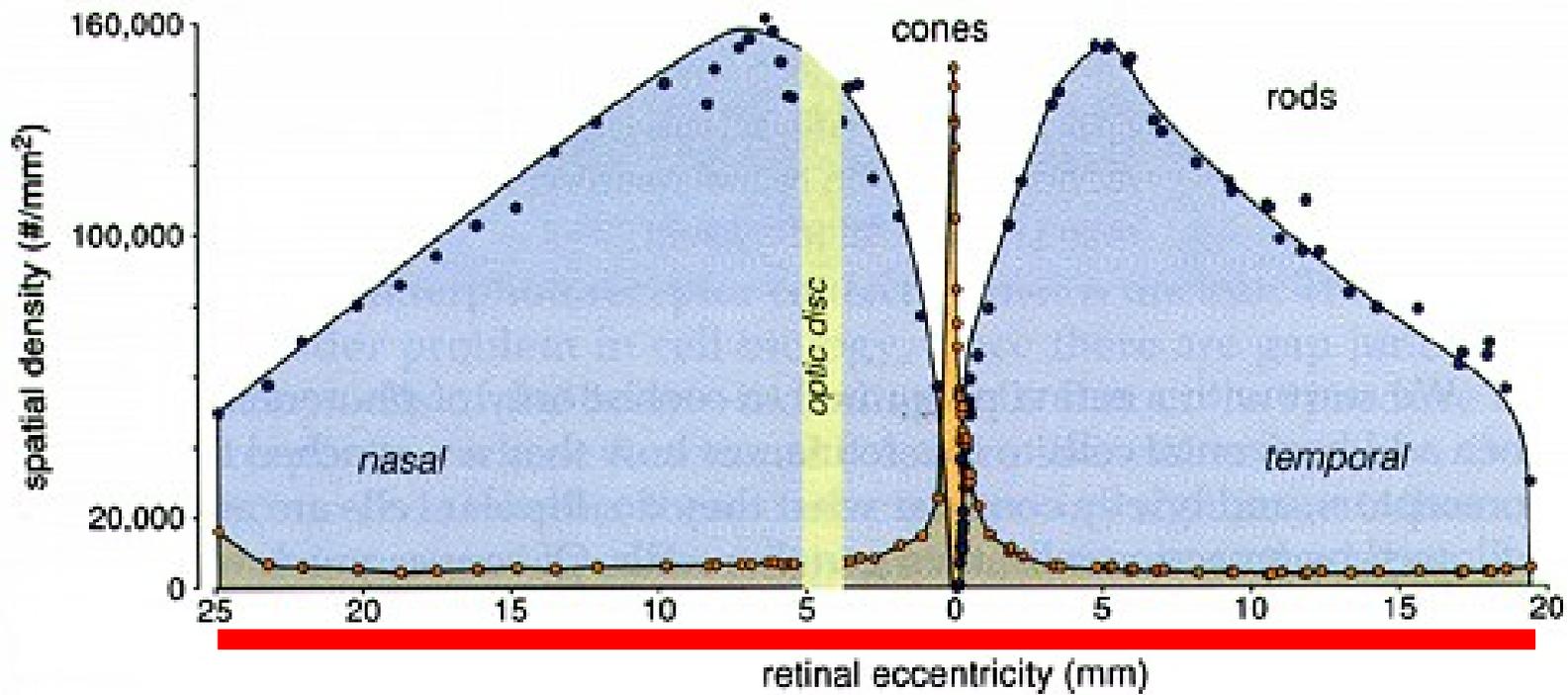
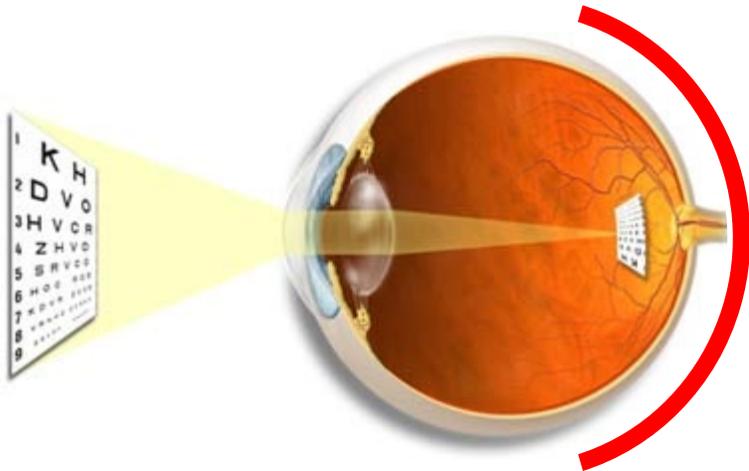


Facts and figures

There are about 120 million rods. They are absent in the central 0.3 mm diameter area of the fovea, known as the *fovea centralis*.

There are only about 6 to 7 million cones. They are much more concentrated in the fovea.

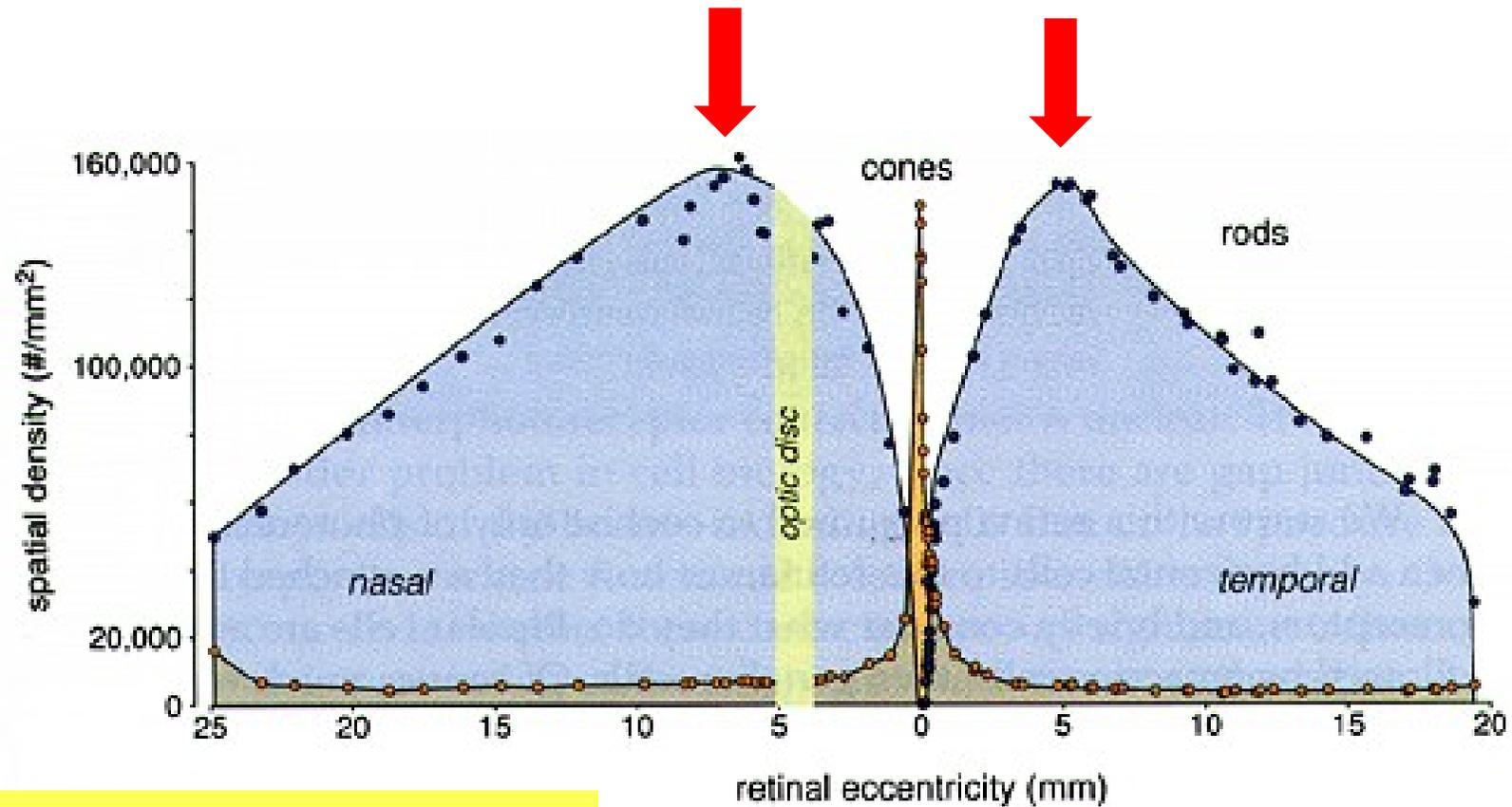
Rod and cone distribution



0.3 mm of eccentricity is about 1 deg of visual angle

after Österberg, 1935; as modified by Rodieck, 1988

Rod density peaks at about
20 deg eccentricity

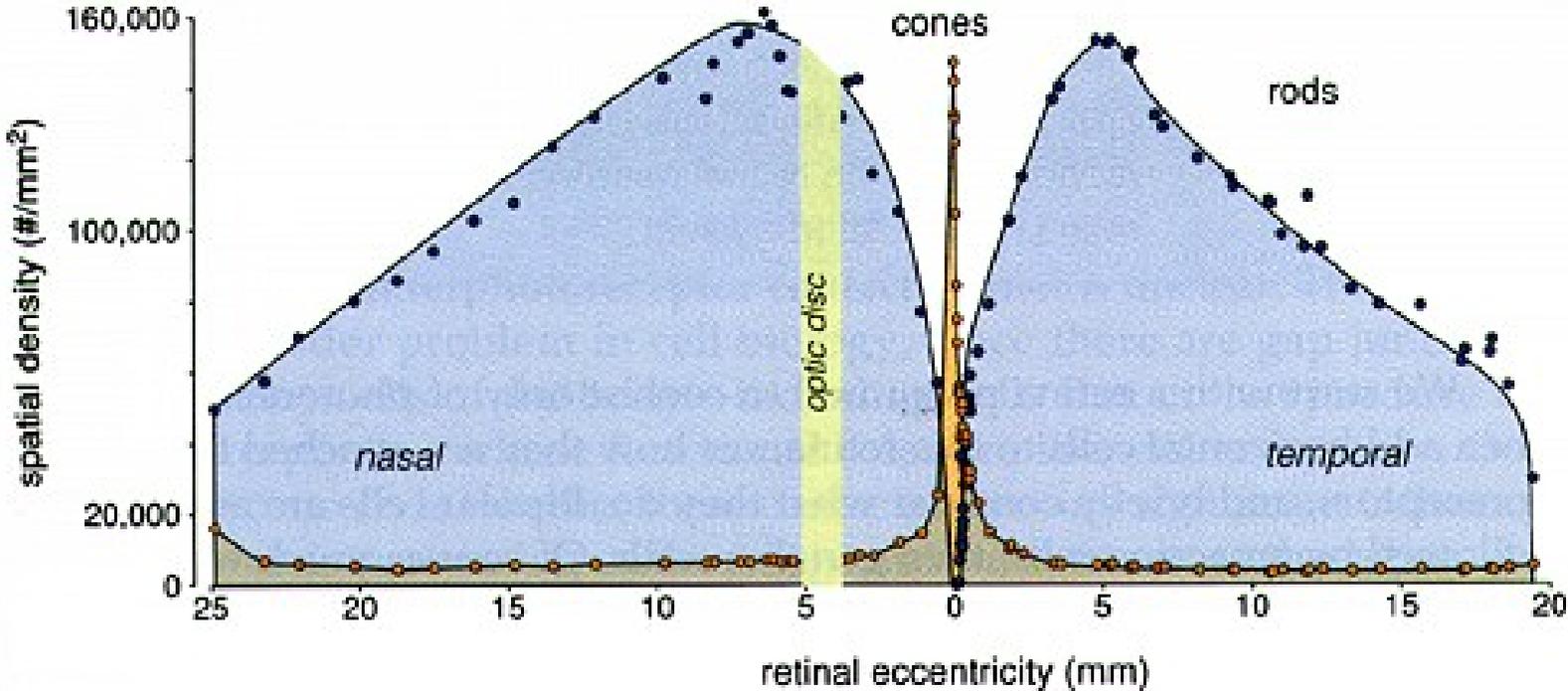


after Österberg, 1935; as modified by Rodieck, 1968

At night, you have to look away from
things to see them in more detail

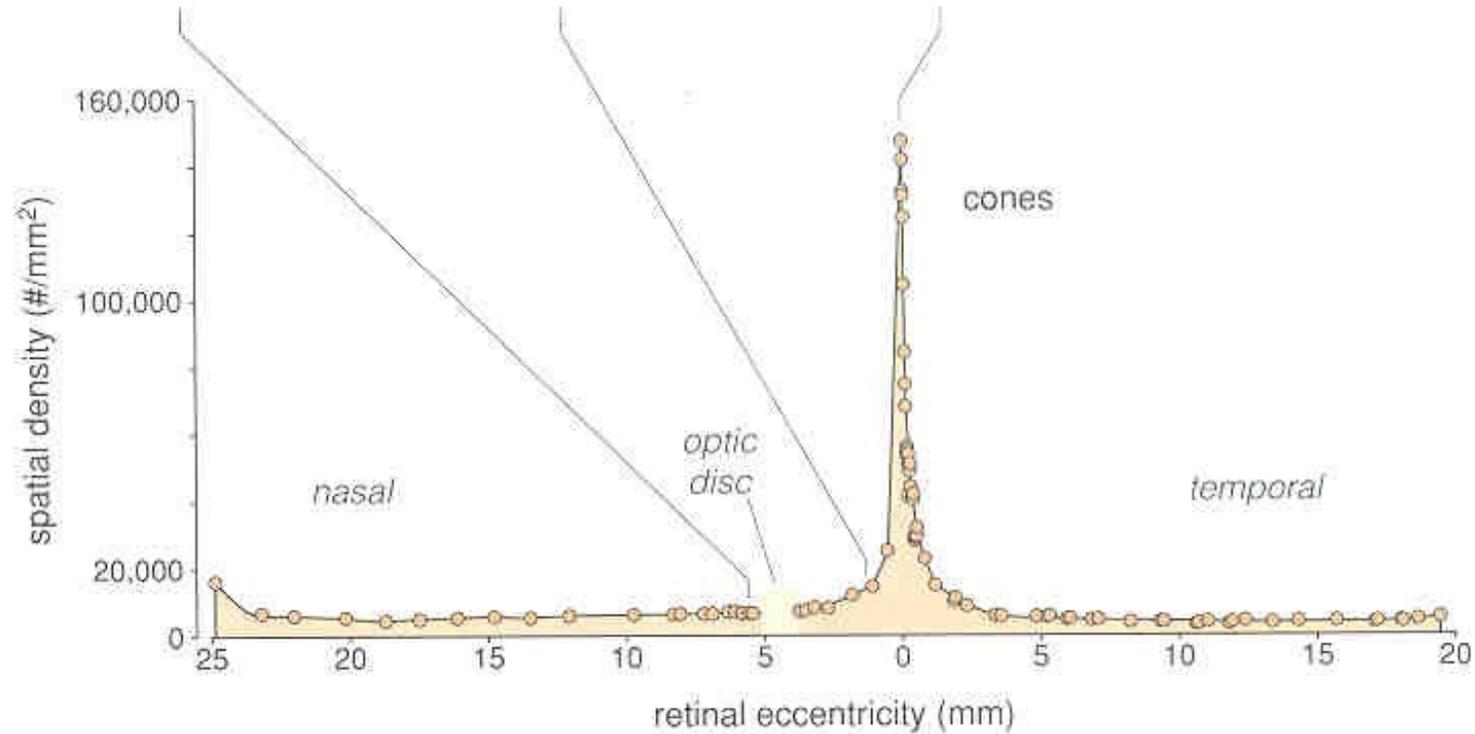
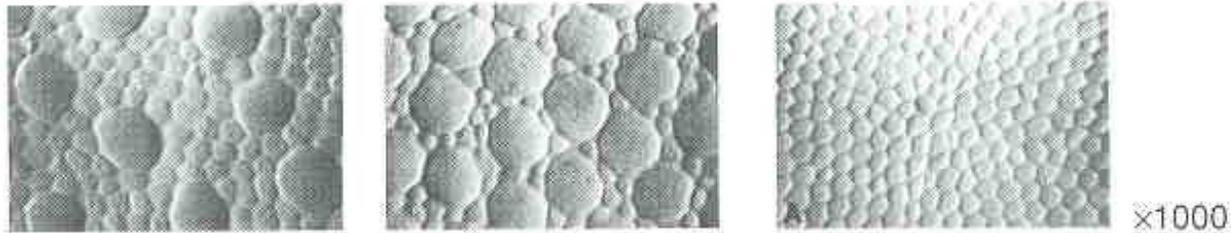
During the day, you have to look at things directly to see them in detail

Cones peak at the centre of vision at 0 deg



after Østerberg, 1935; as modified by Rodieck, 1988

Cone distribution and photoreceptor mosaics



after Østerberg, 1935; as modified by Rodieck 1988;
micrographs from Curcio et al., 1990

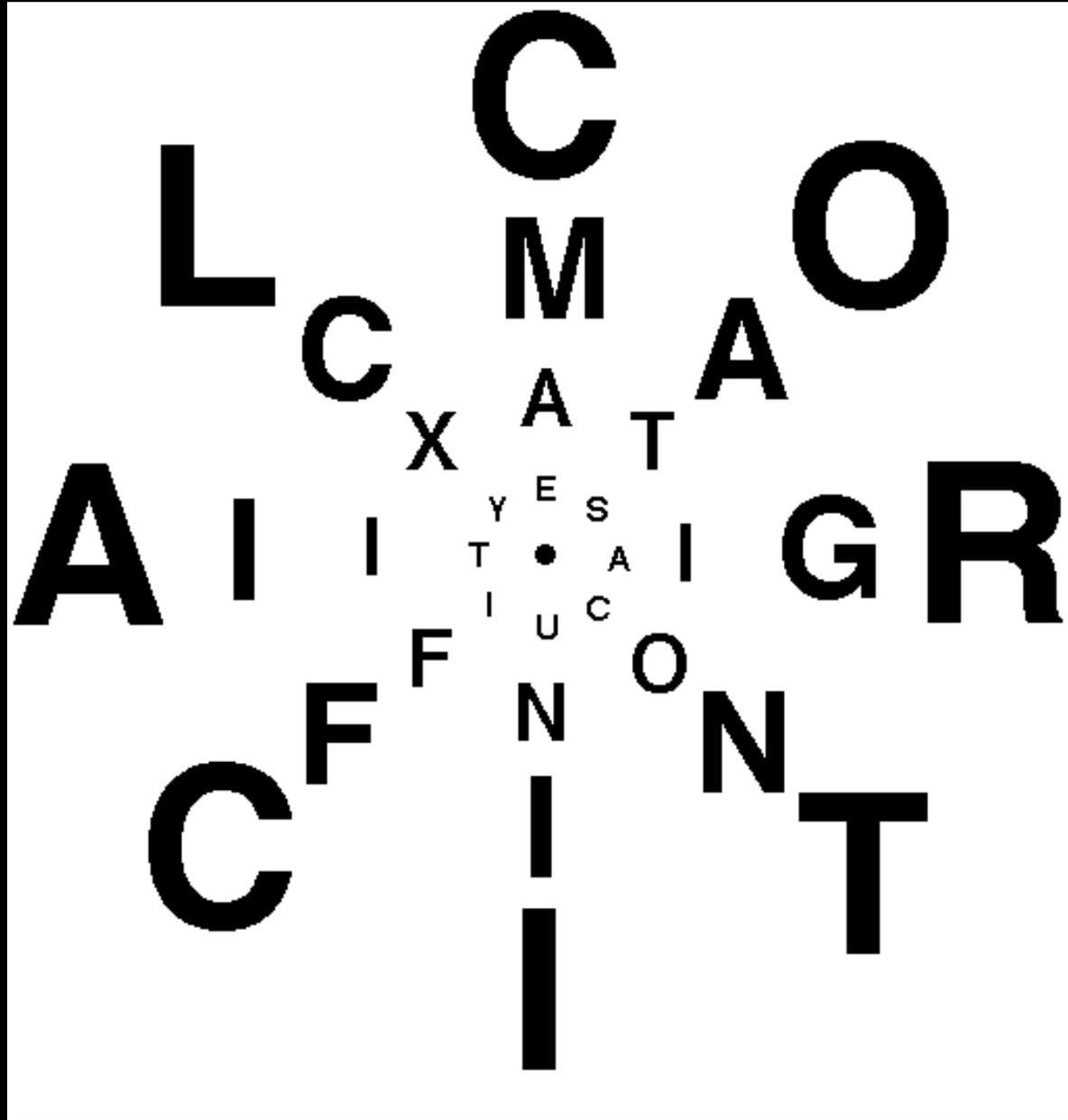
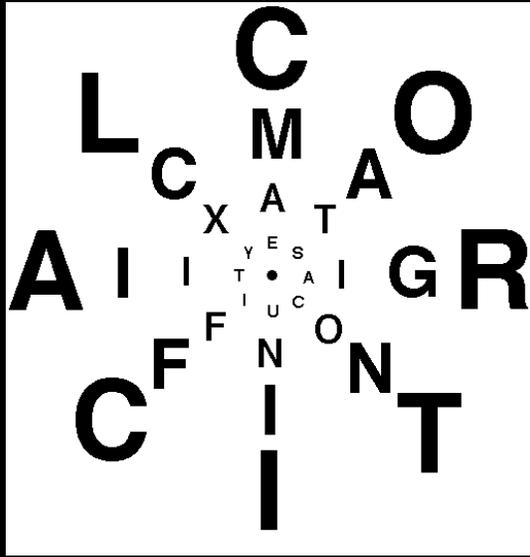
Original photograph



The human visual system is a “foveating” system

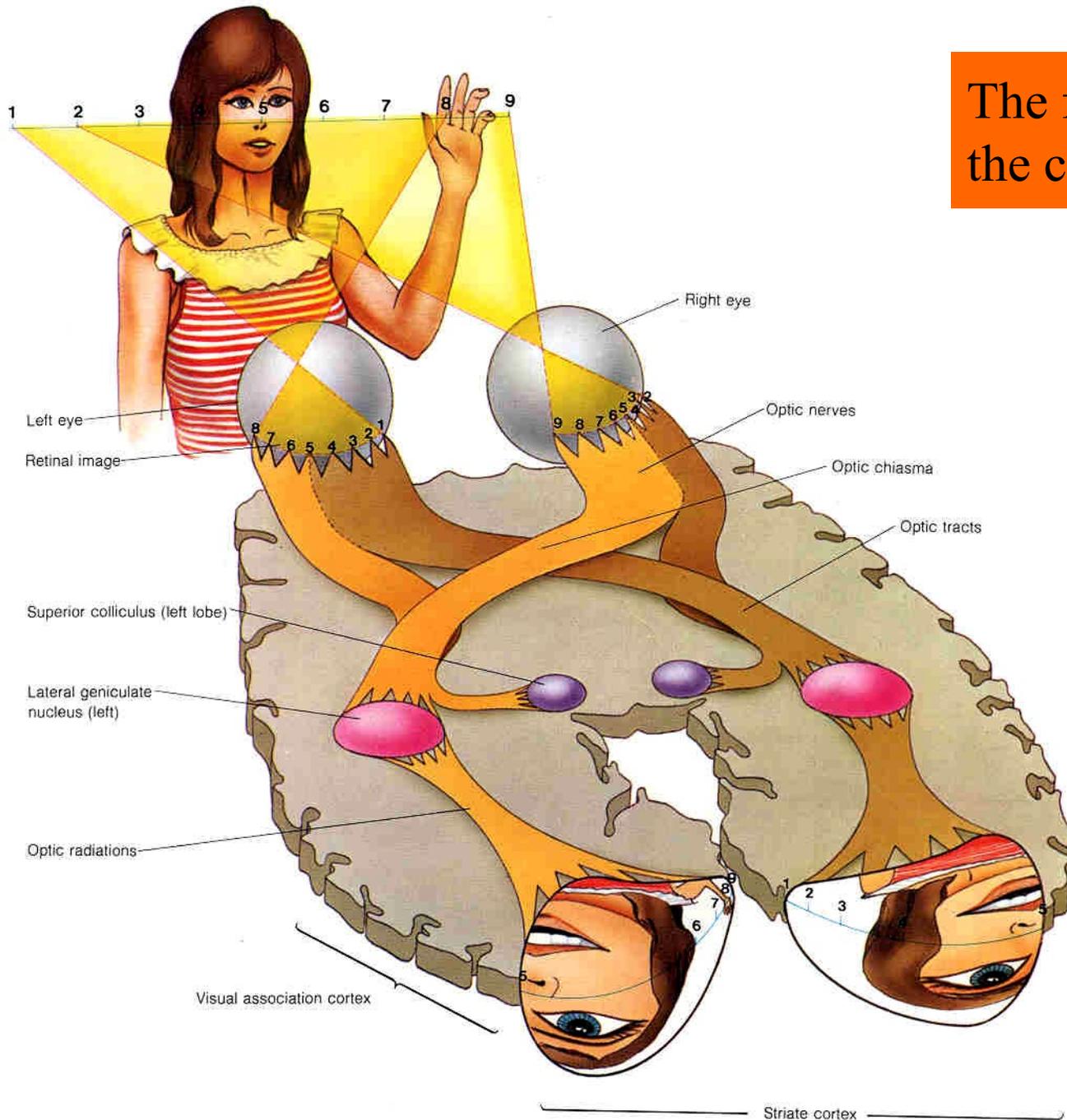
Simulation of what we see when
we fixate with cone vision...





Visual acuity gets much poorer with eccentricity

The foveal region is magnified in the cortical (brain) representation

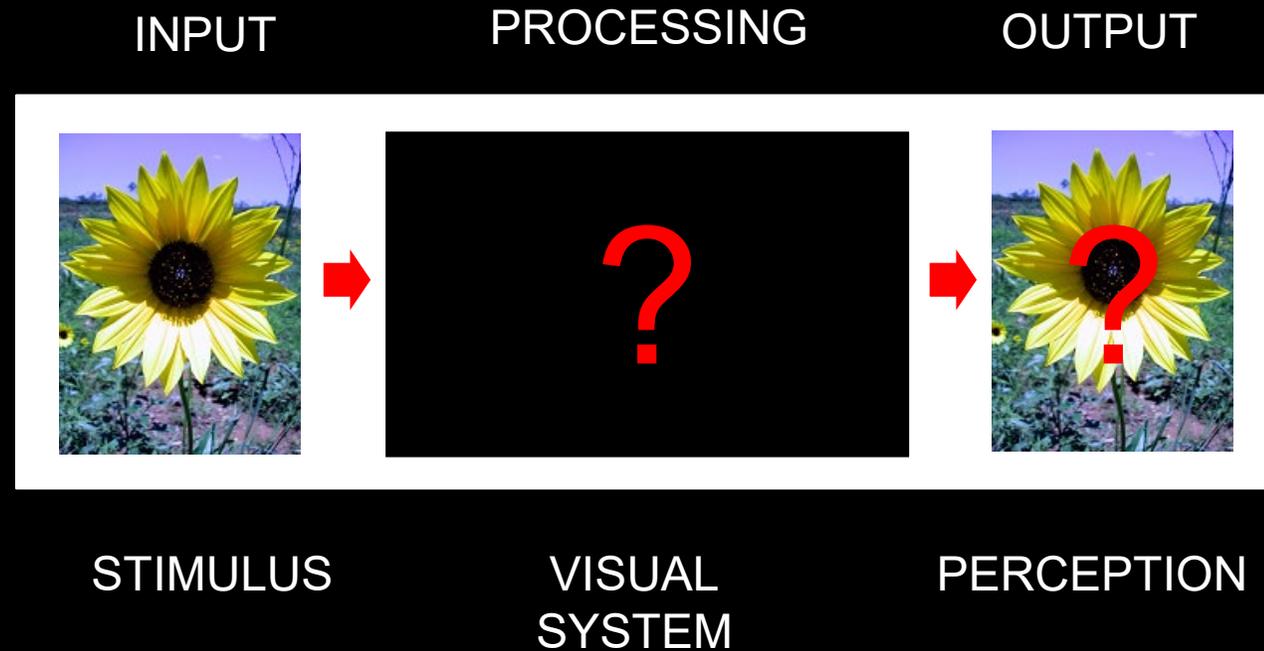


ROD AND CONE DIFFERENCES

Rod and cone differences can be demonstrated using several techniques, including visual psychophysics.

What is visual psychophysics?

Psychophysicists study human vision by measuring an observer's performance on carefully chosen perceptual tasks.



The idea is to work out what is going on inside the visual system from the relationship between the stimulus at the input and the response of the observer.

Rod-cone threshold sensitivity differences

How might we measure them?

Rod and cone
threshold versus
intensity curves

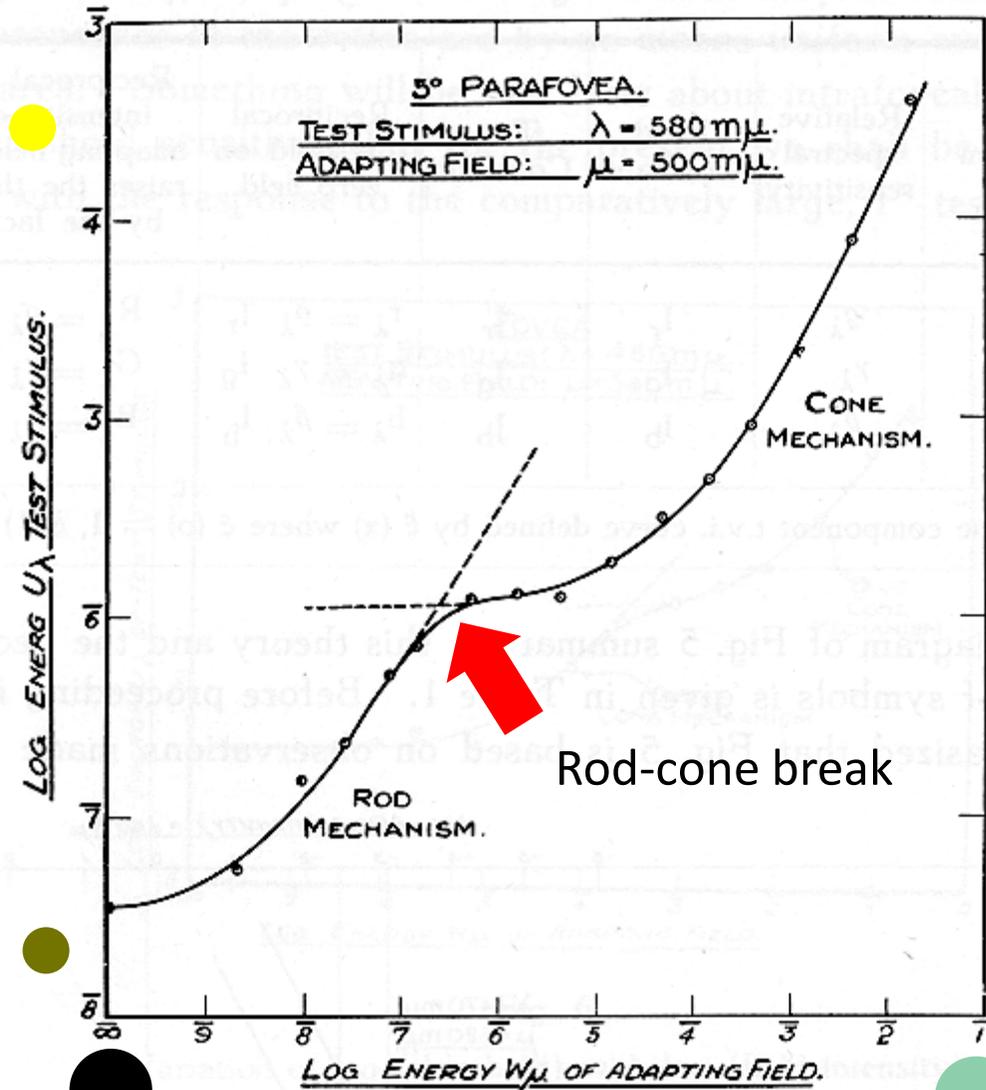
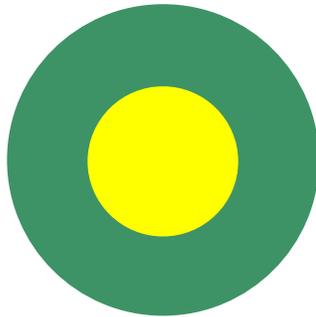


Fig. 4.

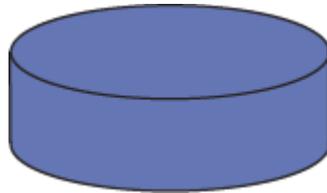
Variation of log (threshold) with log (field intensity) for a 1° flashing test stimulus of yellow light (exposure time 0.063 sec.) on a blue-green field: 5°- parafoveal vision. (Stiles, 1939)

Rods are about one thousand times more sensitive than cones. They can be triggered by individual photons.

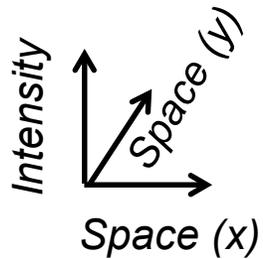
Spectral sensitivity differences

Threshold versus target wavelength measurements

Incremental flash

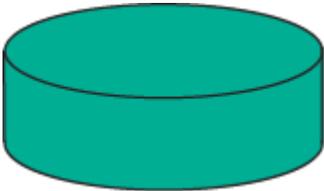


10-deg eccentric fixation

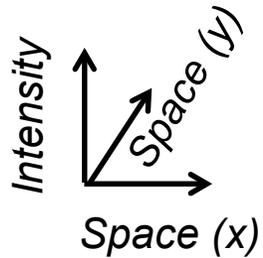


Threshold versus target wavelength measurements

Incremental flash

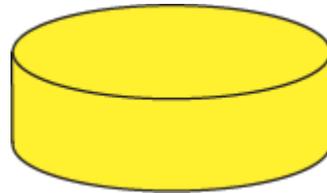


10-deg eccentric fixation

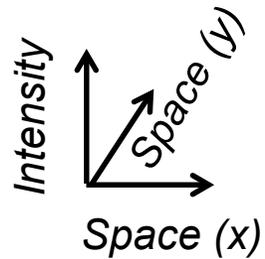


Threshold versus target wavelength measurements

Incremental flash

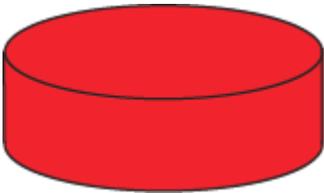


10-deg eccentric fixation

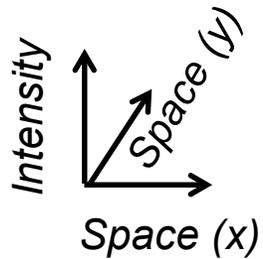


Threshold versus target wavelength measurements

Incremental flash



10-deg eccentric fixation



Rod and cone spectral sensitivity curves

Plotted as “thresholds”
versus wavelength
curves

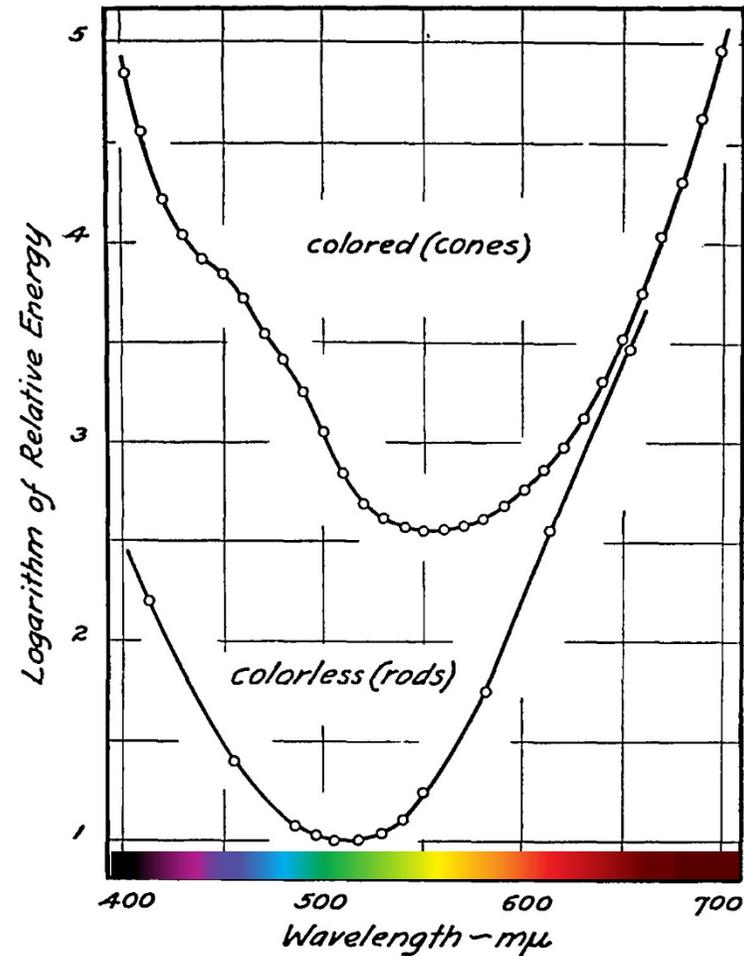
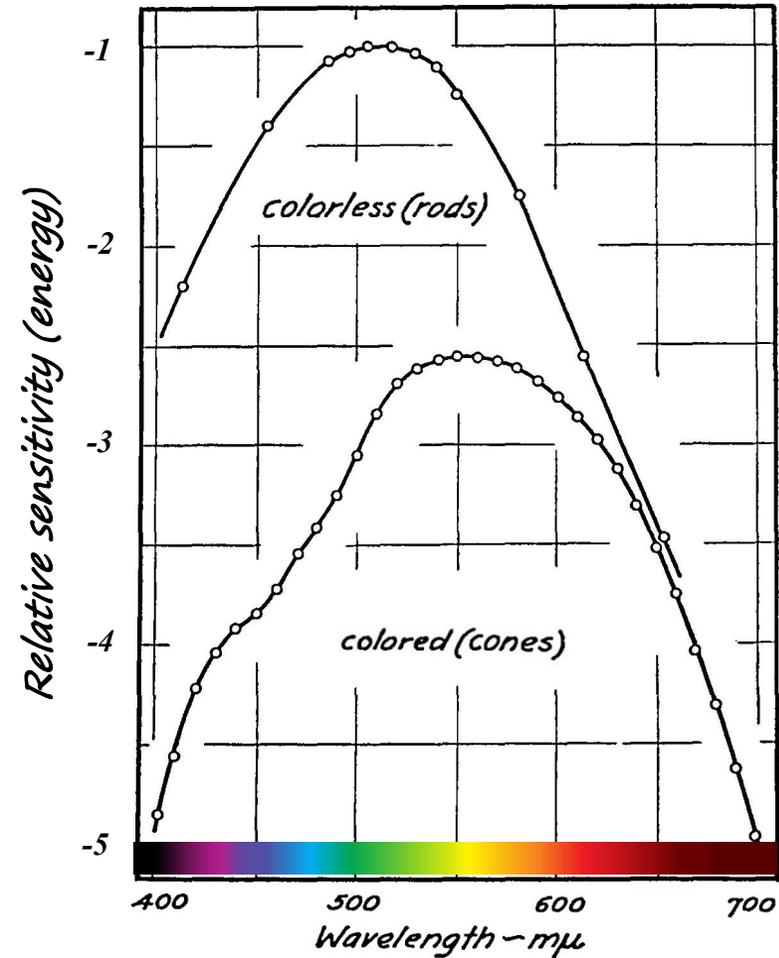


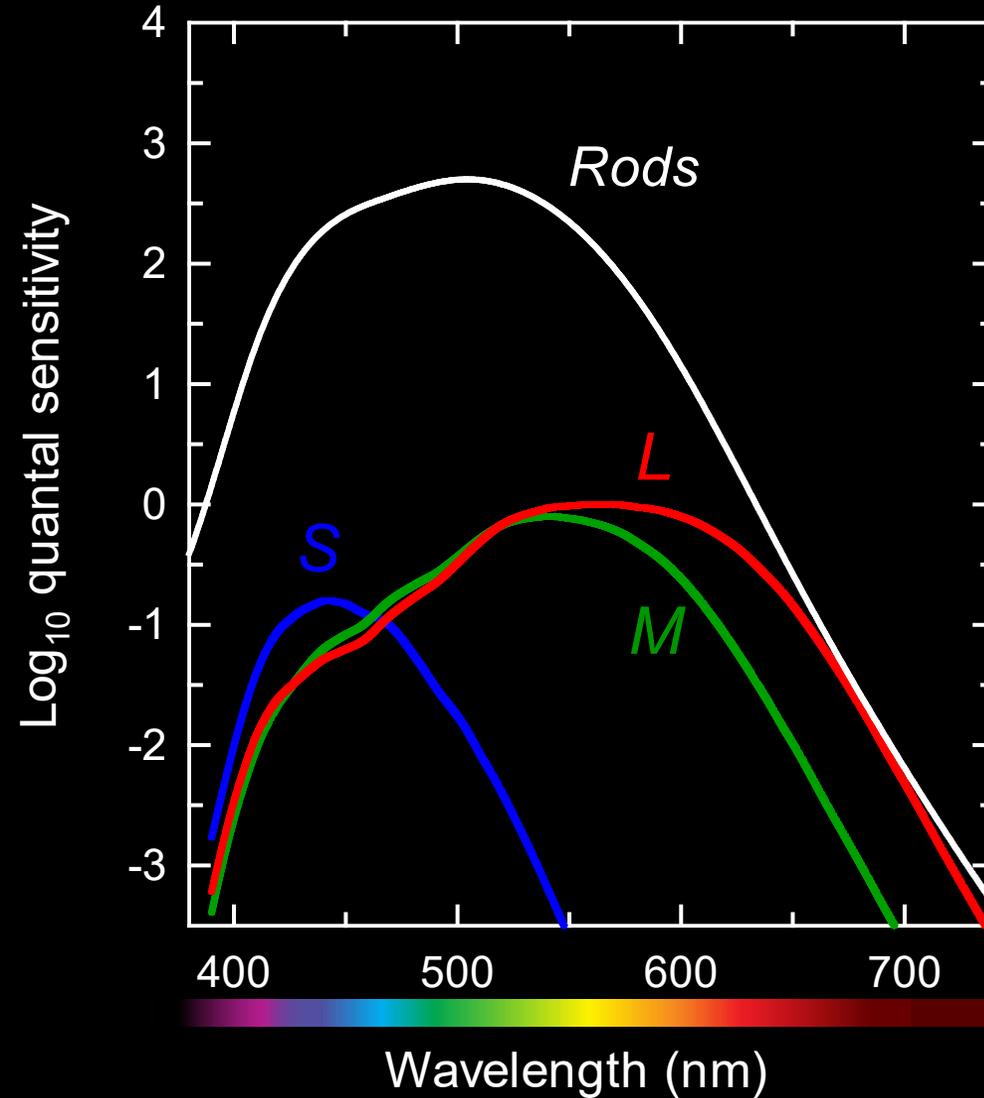
FIG. 2. Spectrum sensibility curves for rod and cone vision on a real energy basis. The data for the separate curves are from the same sources as in Fig. 1. The position of the two curves on the ordinates corresponds to the fact that after complete dark adaptation, any region of the retina outside the fovea sees red light of 650 mμ as colorless at the threshold, and as colored only above the threshold. The precise energy increment above the threshold for the appearance of color (cone function) varies for different parts of the retina; in the parafovea it lies between 0.1 and 1.0 log unit.

Plotted as the more conventional spectral “sensitivity” curve



Sensitivity = 1/threshold
or
 $\log(\text{sensitivity}) = -\log(\text{threshold})$

Approximate dark-adapted photoreceptor sensitivities.



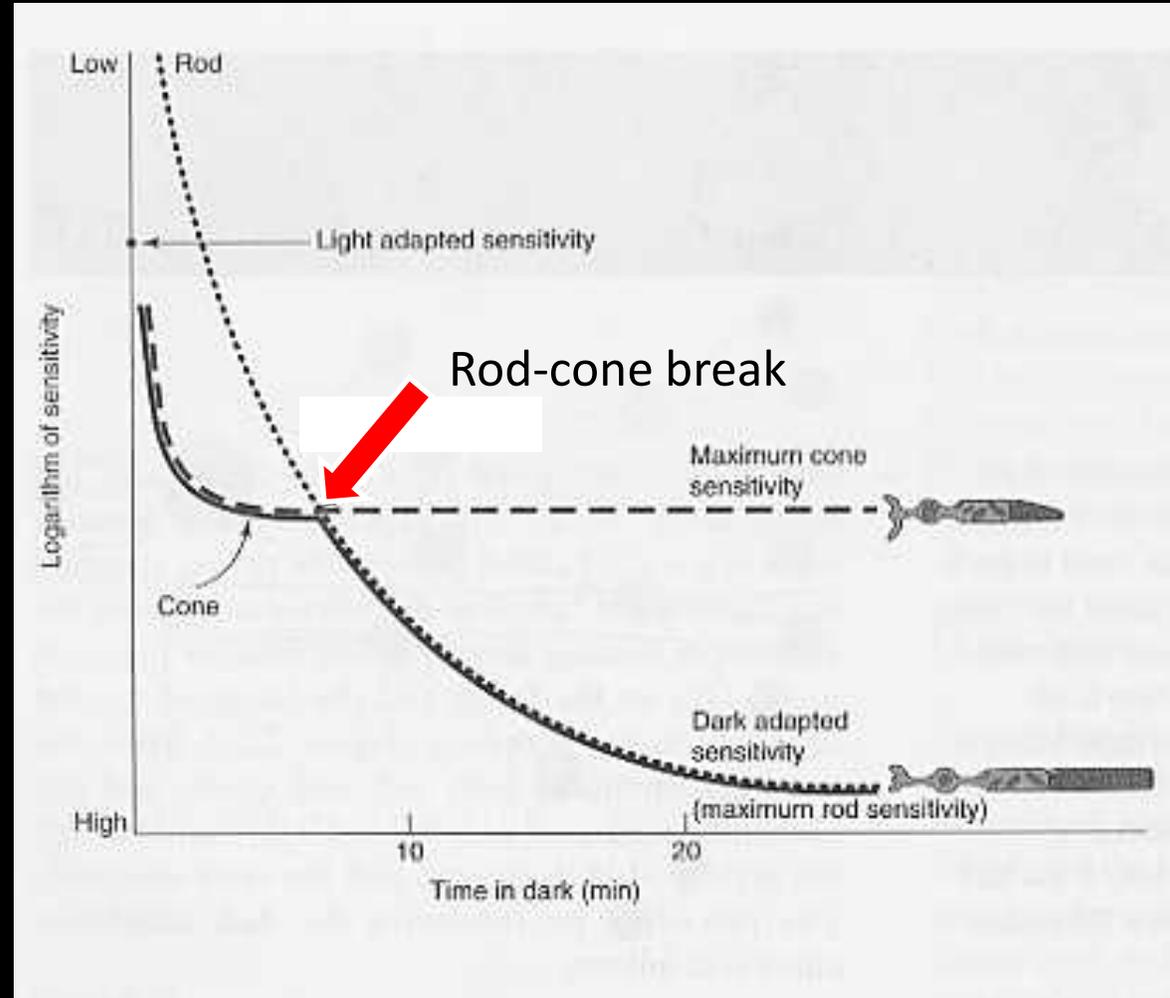


The Purkinje Shift

A change in the relative brightness of colours as the light level changes because of the difference in spectral sensitivity between rod and cone vision (*e.g.*, reds and oranges become darker as rods take over)

Simulated: Dick Lyon & Lewis Collard at Wikimedia

Rod-cone dark adaptation curves



Rod-cone dark adaptation curves

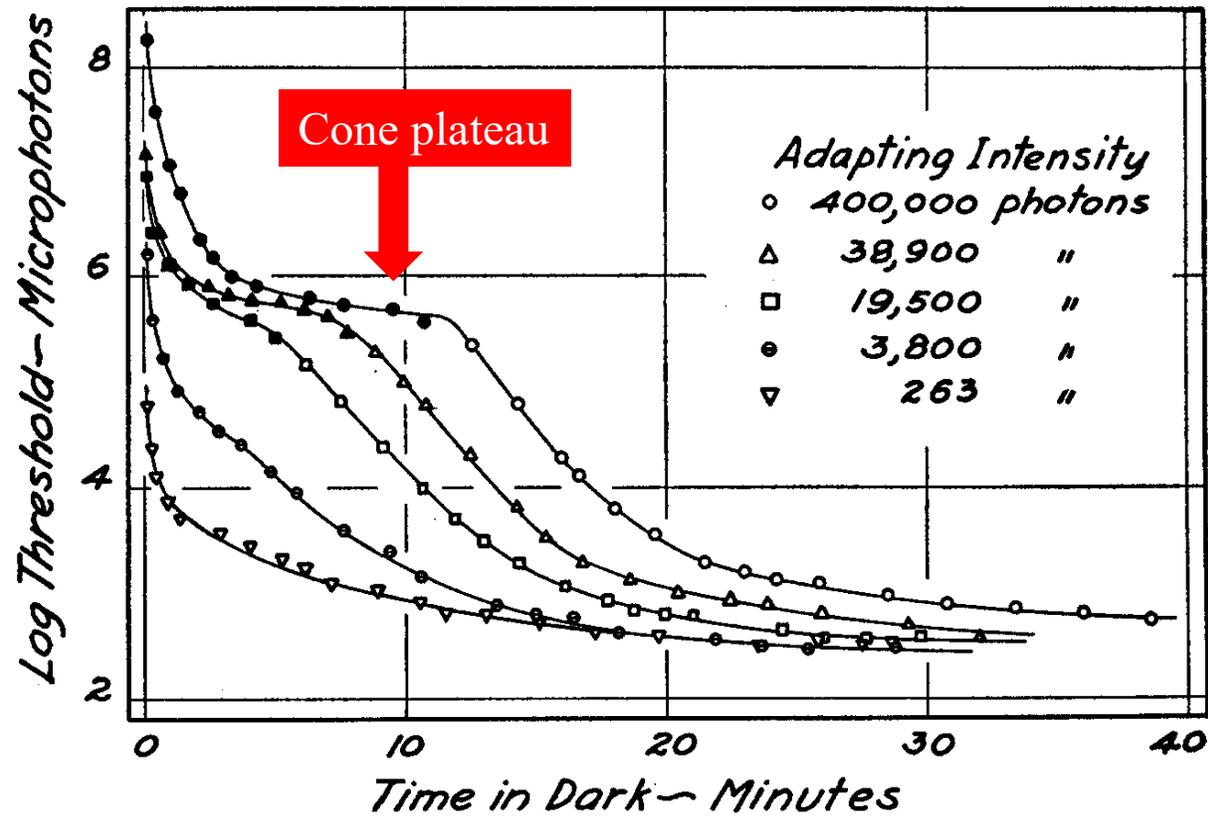


FIG. 2. The course of dark adaptation as measured with violet light following different degrees of light adaptation. The filled-in symbols indicate that a violet color was apparent at the threshold, while the empty symbols indicate that the threshold was colorless.

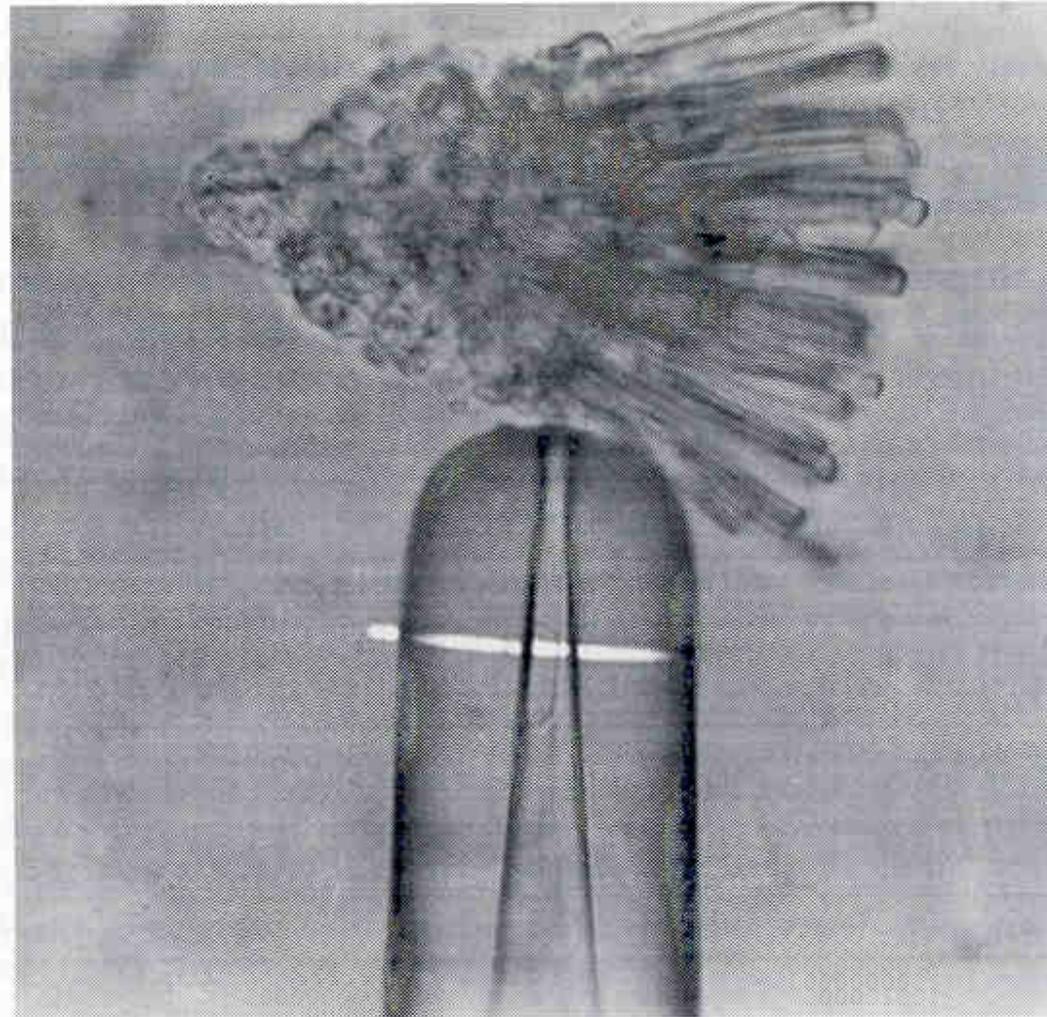
Rods take much longer to recover after a bleach than cones

From Hecht, Haig & Chase (1937)

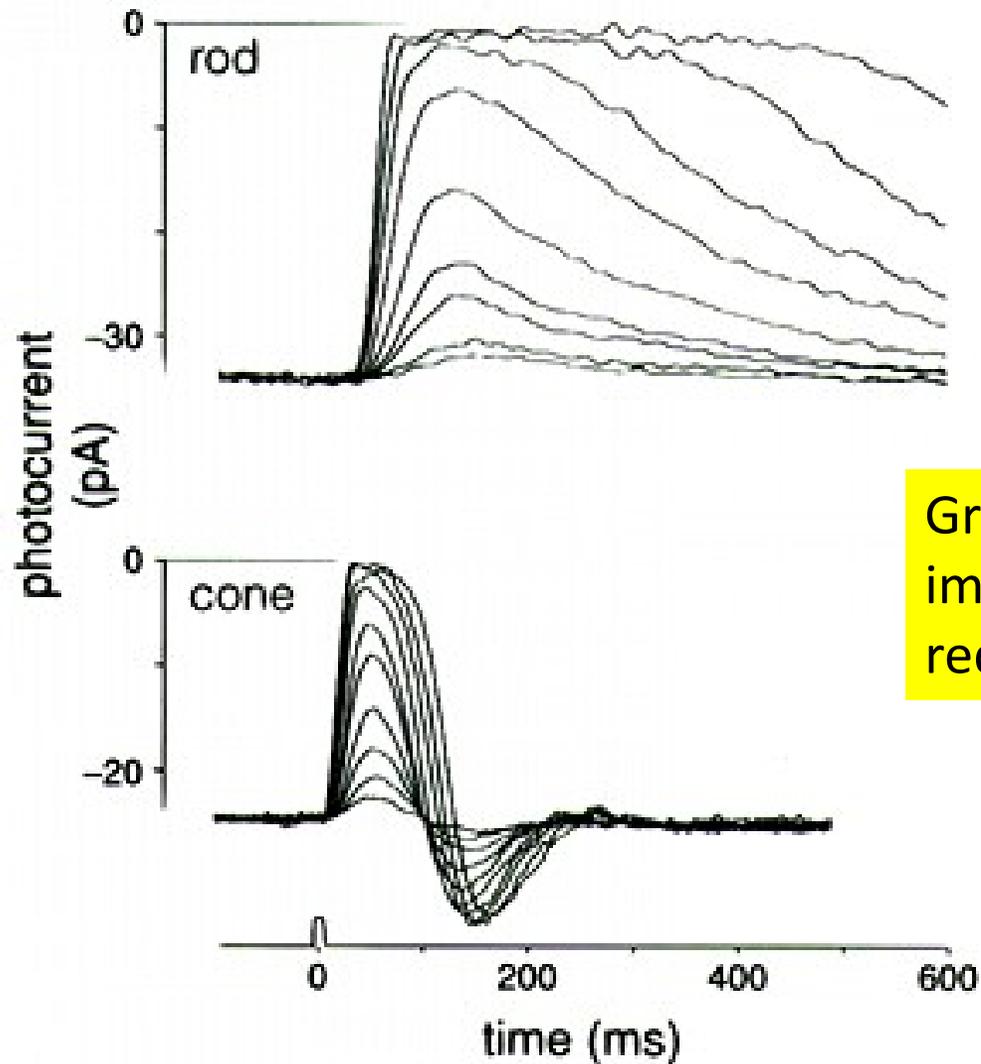
Temporal differences

Suction electrode recording

4.15 MEASURING CONE PHOTOCURRENTS. The image shows a portion of macaque retina suspended in solution. A single photoreceptor from this retinal section has been drawn into a micropipette and is being stimulated by a beam of light passing transversely through the photoreceptor and micropipette. Courtesy of Denis Baylor.



Photocurrent responses



Greater temporal integration improves rod sensitivity (but reduces temporal acuity)

Highest flicker rates that can just be seen (c.f.f.)...

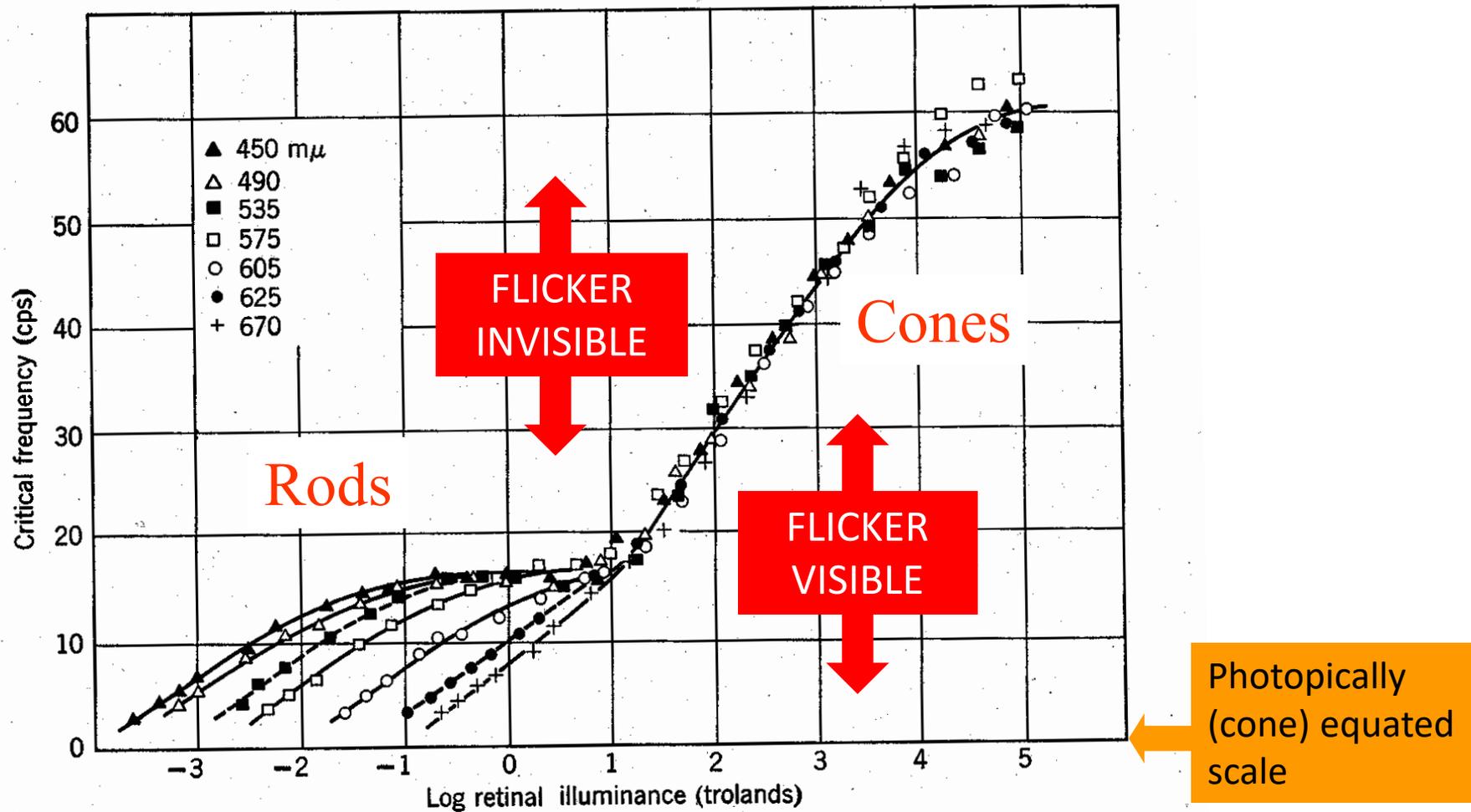


FIG. 10.6 Relation of CFF to log retinal illuminance for seven spectral regions. (Hecht and Shlaer, 1936. Reprinted by permission of The Rockefeller Institute Press from *The Journal of General Physiology*, 1936, 19, 956-979; Fig. 3.)

Spatial differences
(visual acuity)

Rod and cone visual acuities

Visual acuity



The acuity here is defined as the reciprocal value of the size of the gap (measured in arc minutes) that can be reliably identified.

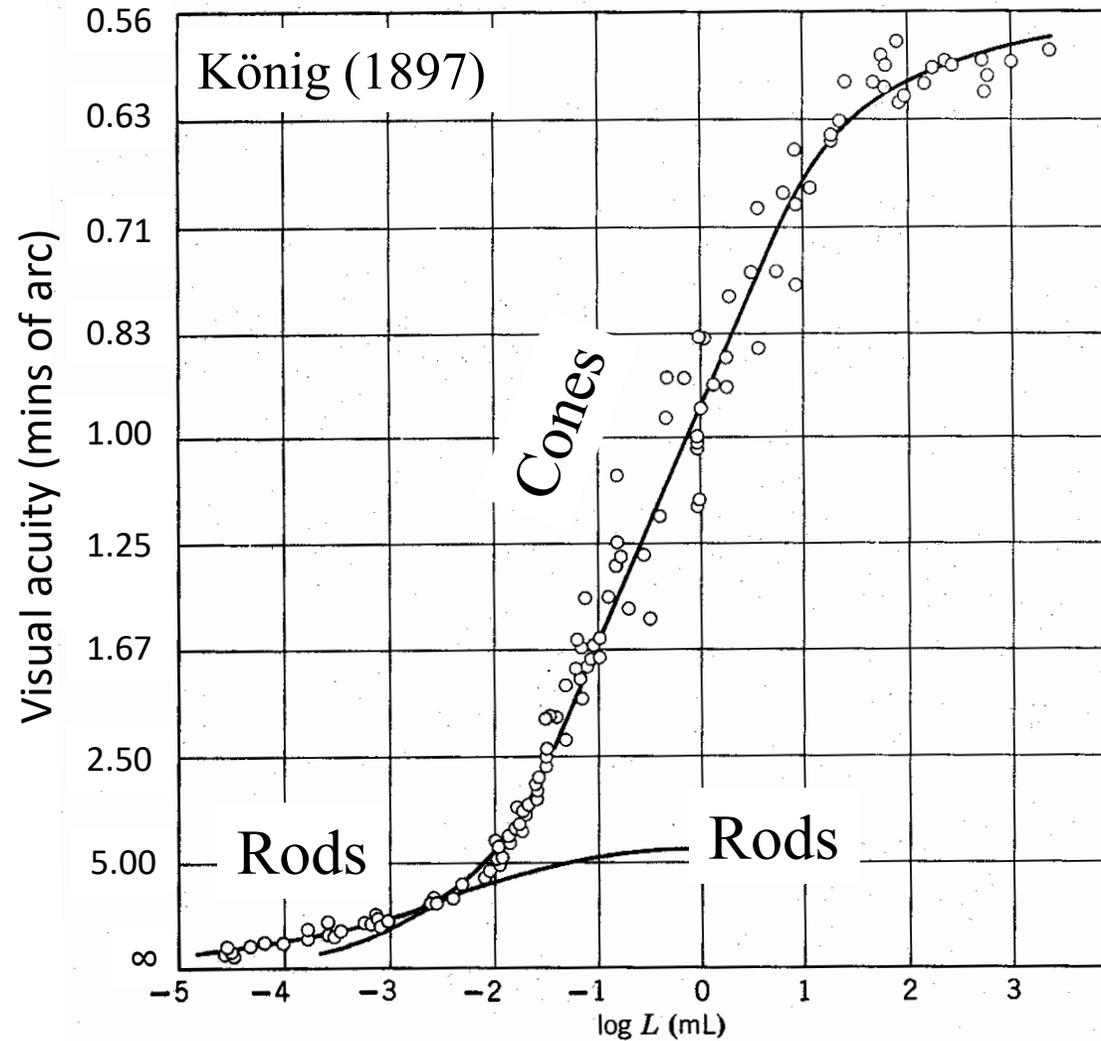


FIG. 11.14 König's data for the relation between visual acuity and illumination, as replotted by Hecht (1934). The shallow curve for the lower limb of the data is an equation for rods, whereas the upper curve is for cones. The task is one of recognizing the orientation of a hook form of test object.

Rod and cone visual acuities

Greater spatial integration improves rod sensitivity but reduces acuity

The loss must be postreceptoral because the rods are smaller than cones in the periphery)

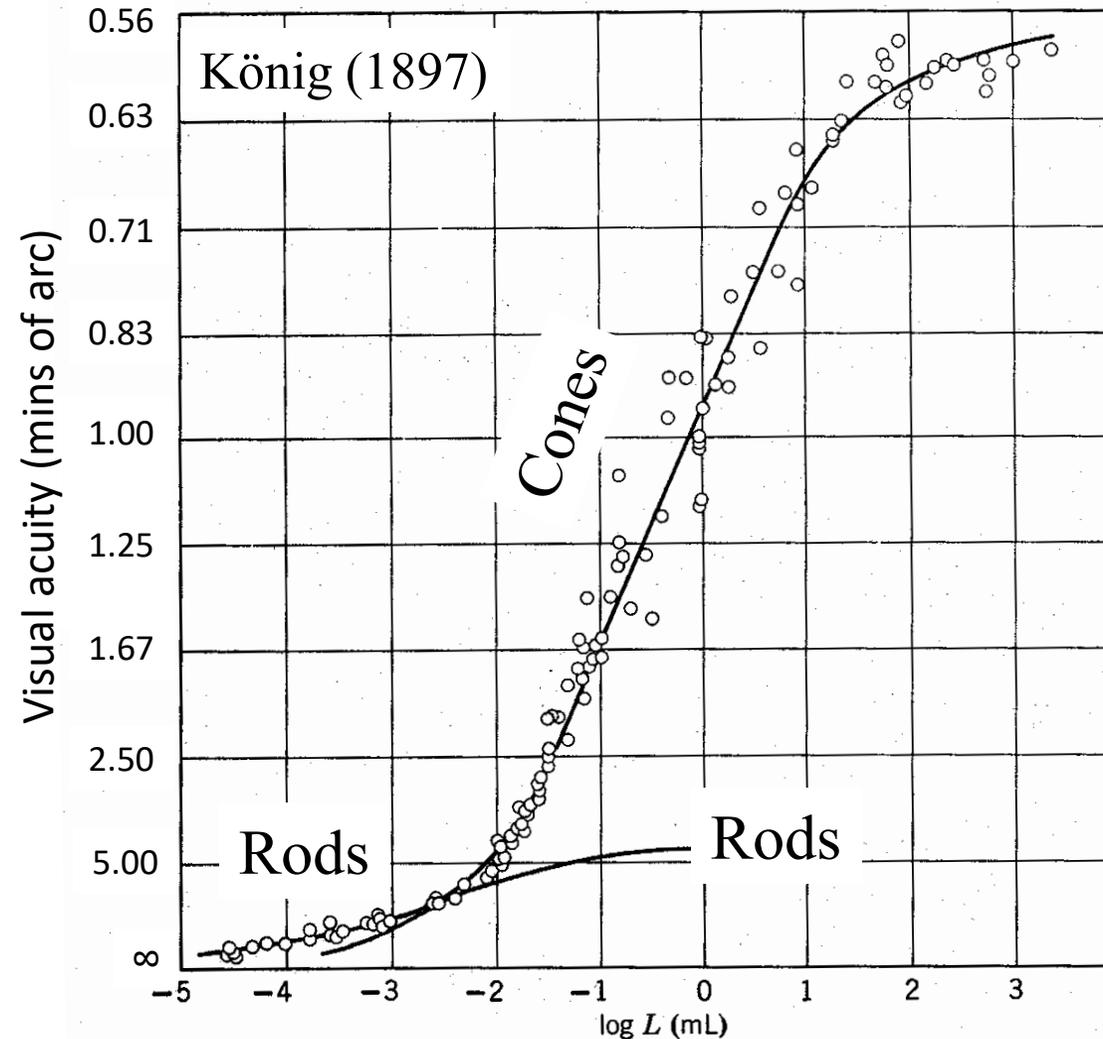


FIG. 11.14 König's data for the relation between visual acuity and illumination, as replotted by Hecht (1934). The shallow curve for the lower limb of the data is an equation for rods, whereas the upper curve is for cones. The task is one of recognizing the orientation of a hook form of test object.

Rod vision is achromatic

Why?

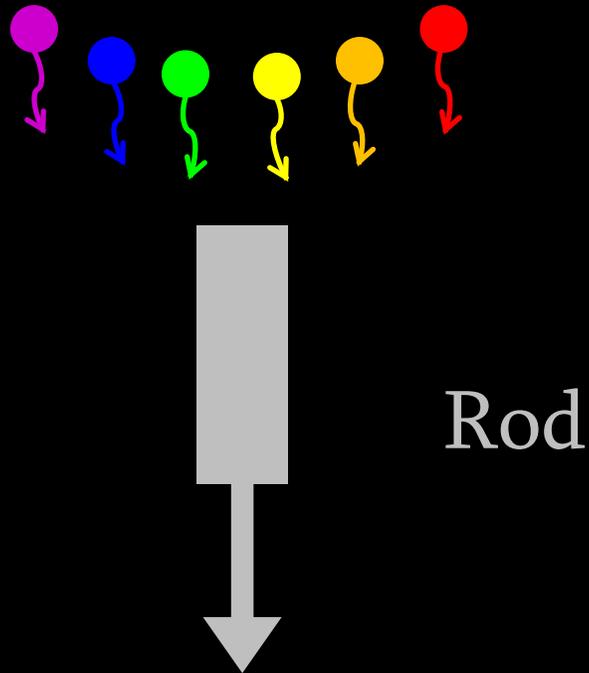
Vision at the photoreceptor stage is relatively simple
because the output of each photoreceptor is:

UNIVARIANT

What does univariant mean?

UNIVARIANCE

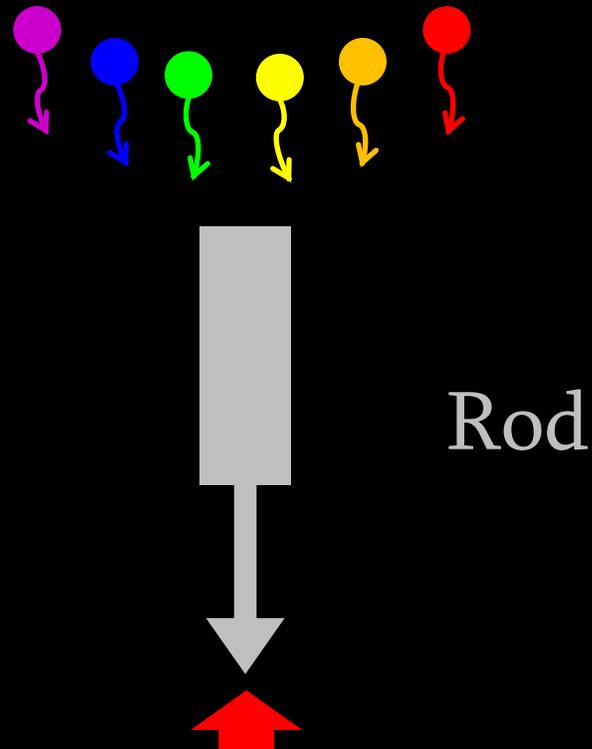
Crucially, the effect of any absorbed photon is *independent* of its wavelength.



Once absorbed a photon produces the *same* change in photoreceptor output whatever its wavelength.

UNIVARIANCE

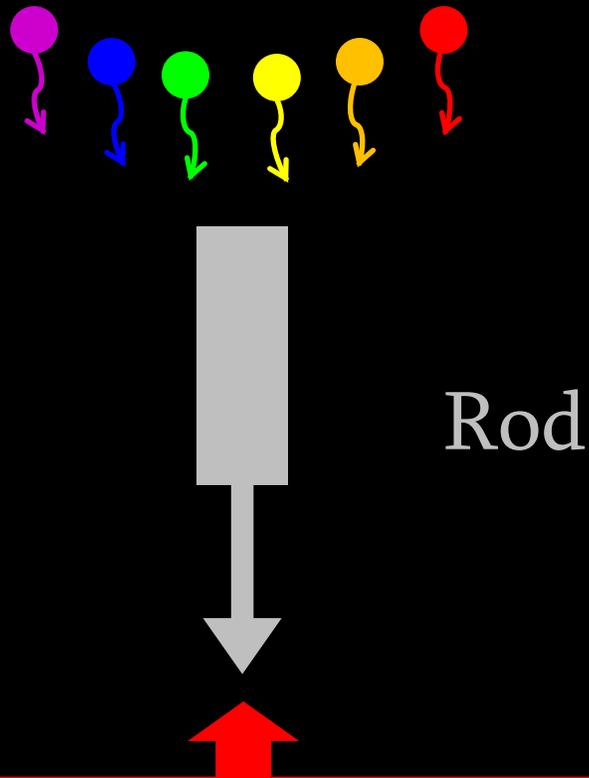
Crucially, the effect of any absorbed photon is *independent* of its wavelength.



So, if you monitor the rod output, you can't tell which "colour" of photon has been absorbed.

UNIVARIANCE

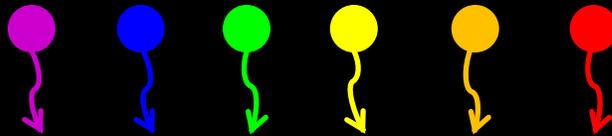
Crucially, the effect of any absorbed photon is *independent* of its wavelength.



All the photoreceptor effectively does is to count photons.

UNIVARIANCE

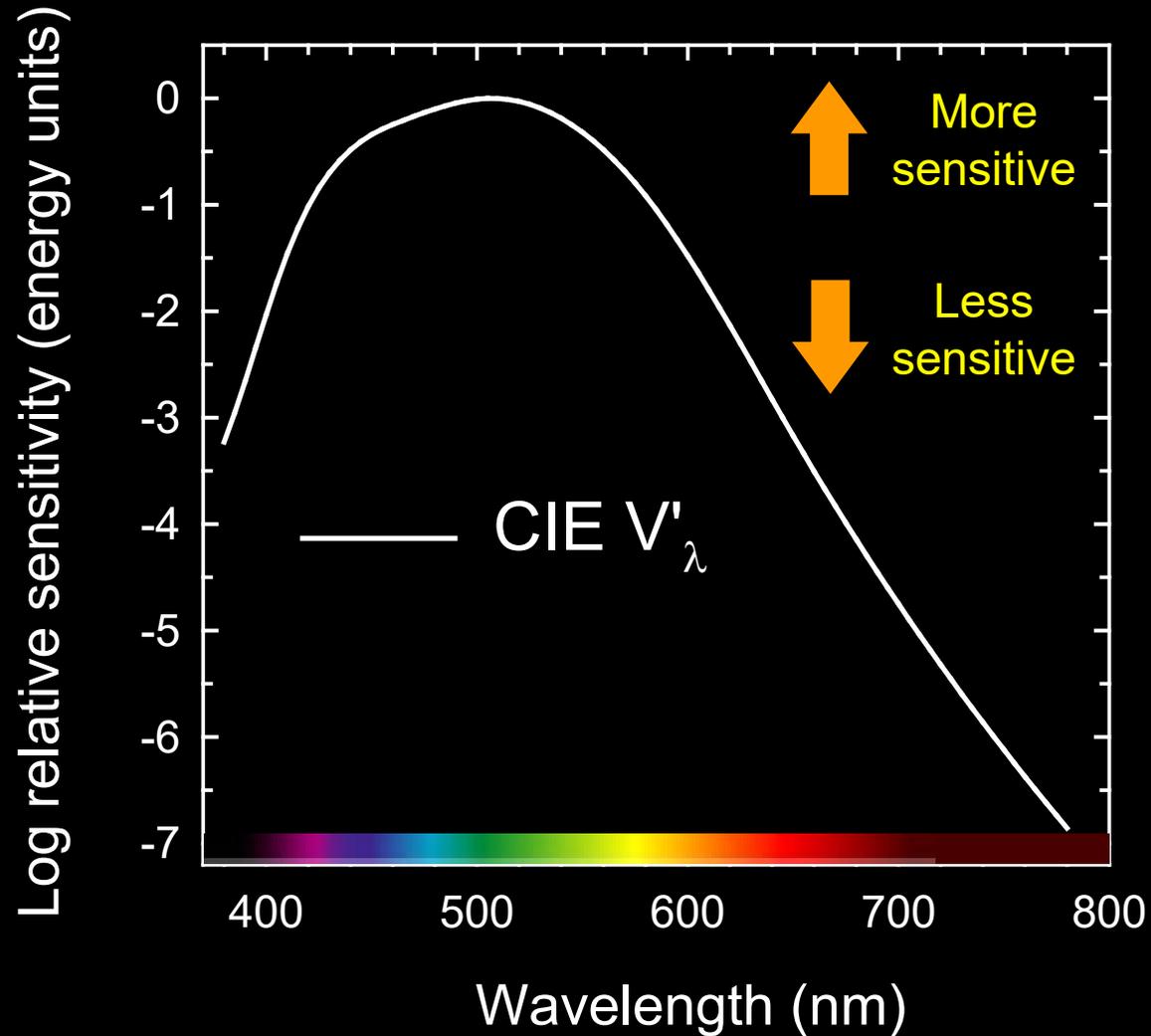
What does vary with wavelength is the **probability** that a photon will be absorbed.



This is reflected in what is called a “spectral sensitivity function”.

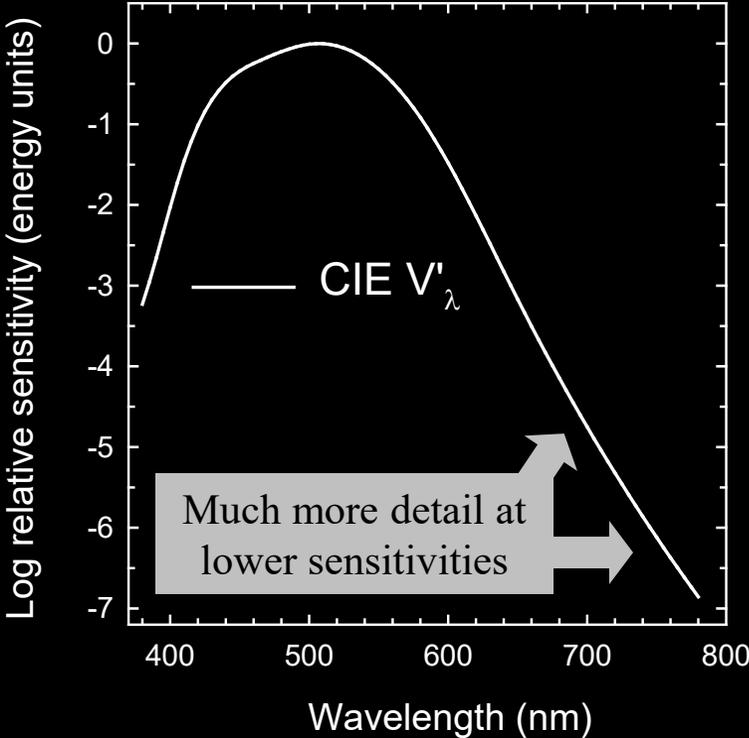
Rod spectral sensitivity function

(also known as the scotopic luminosity curve, CIE V'_λ)



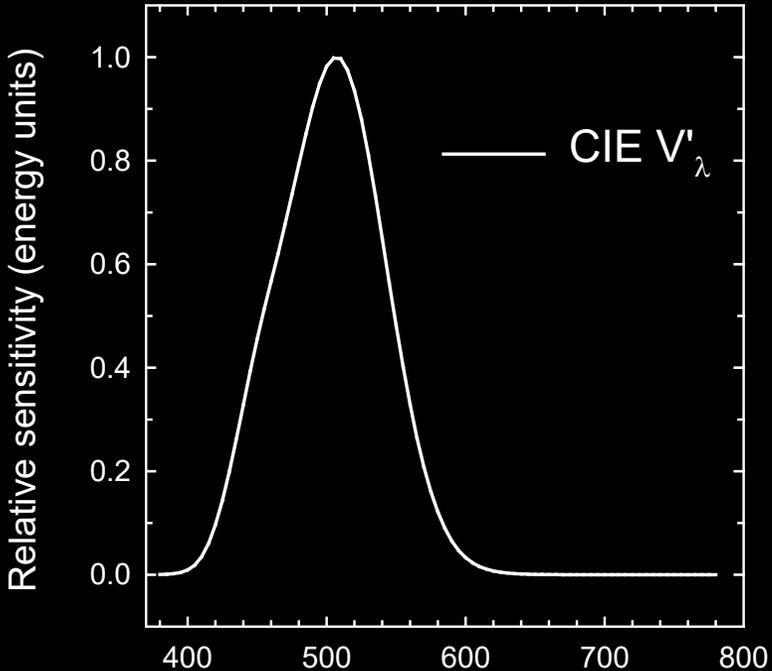
Rod spectral sensitivity function (V'_λ)

Logarithmic sensitivity plot

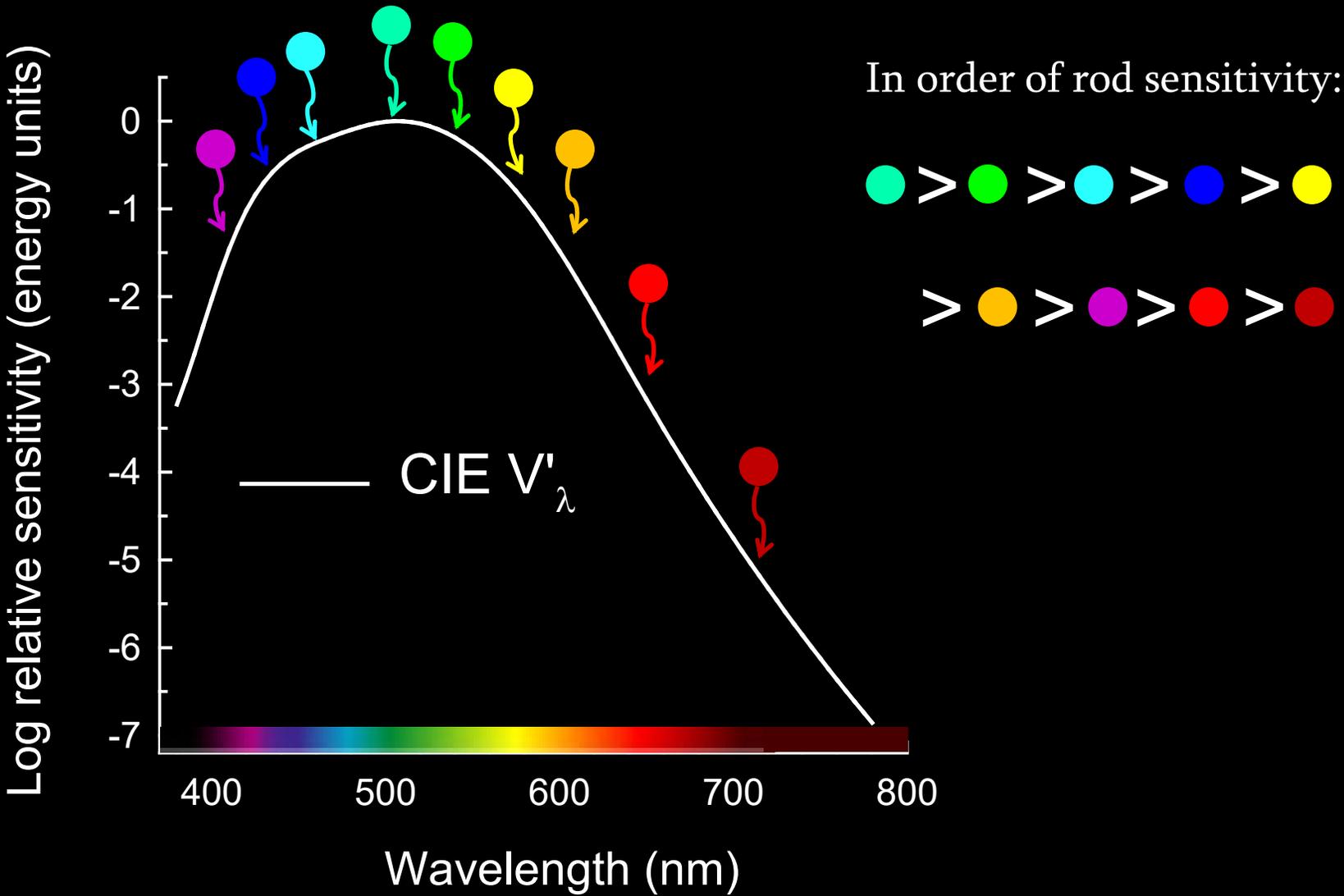


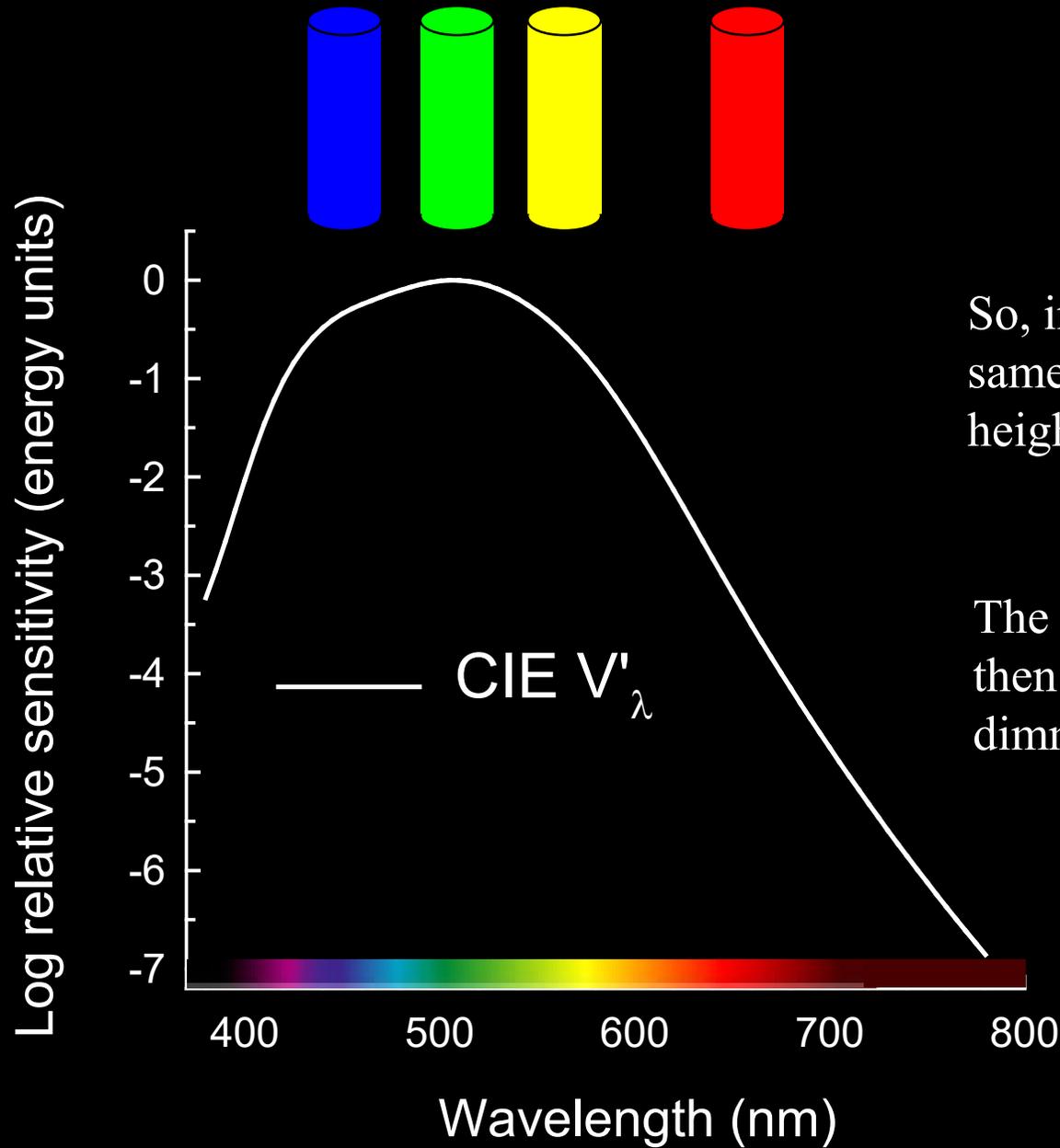
$10^{V'}$
→
 $\log(V')$
←

Linear sensitivity plot



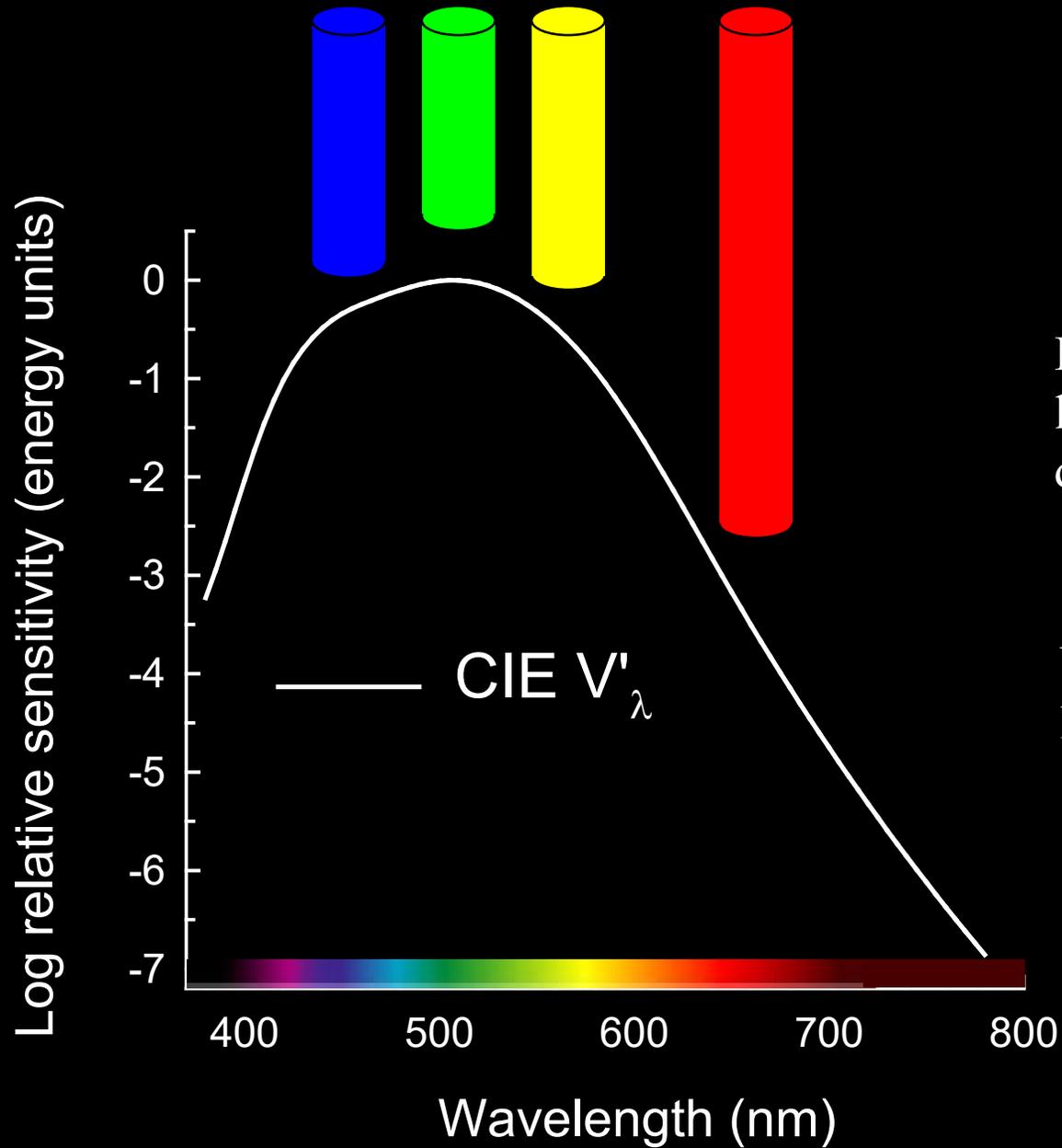
Rod spectral sensitivity function (V'_λ)





So, imagine you have four lights of the same intensity (indicated here by their height). How will they appear?

The green will look brightest, then blue, then yellow and lastly the red will be the dimmest



If instead we adjust the intensities of the light to compensate for the sensitivity differences, how will they appear?

When this has been done, the four lights will look completely identical.



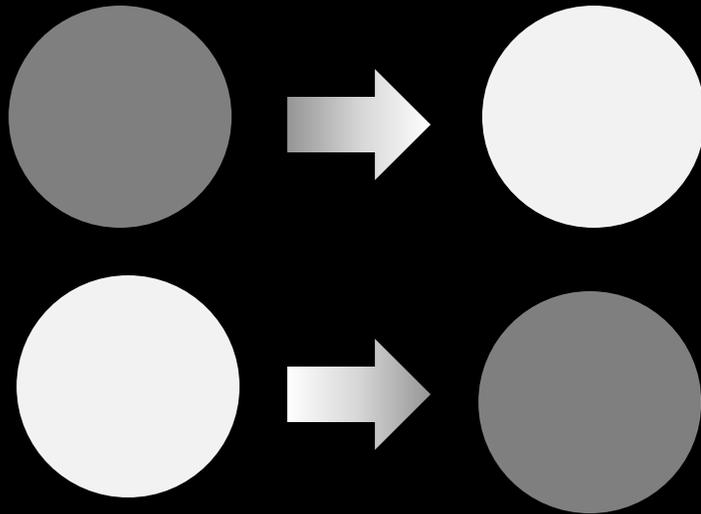
Rod

Changes in light intensity are confounded
with changes in colour (wavelength)



UNIVARIANCE

A change in photoreceptor output can be caused by a change in intensity or by a change in colour. There is no way of telling which.



Colour or intensity
change??

Each photoreceptor is therefore 'colour blind', and is unable to distinguish between changes in colour and changes in intensity.

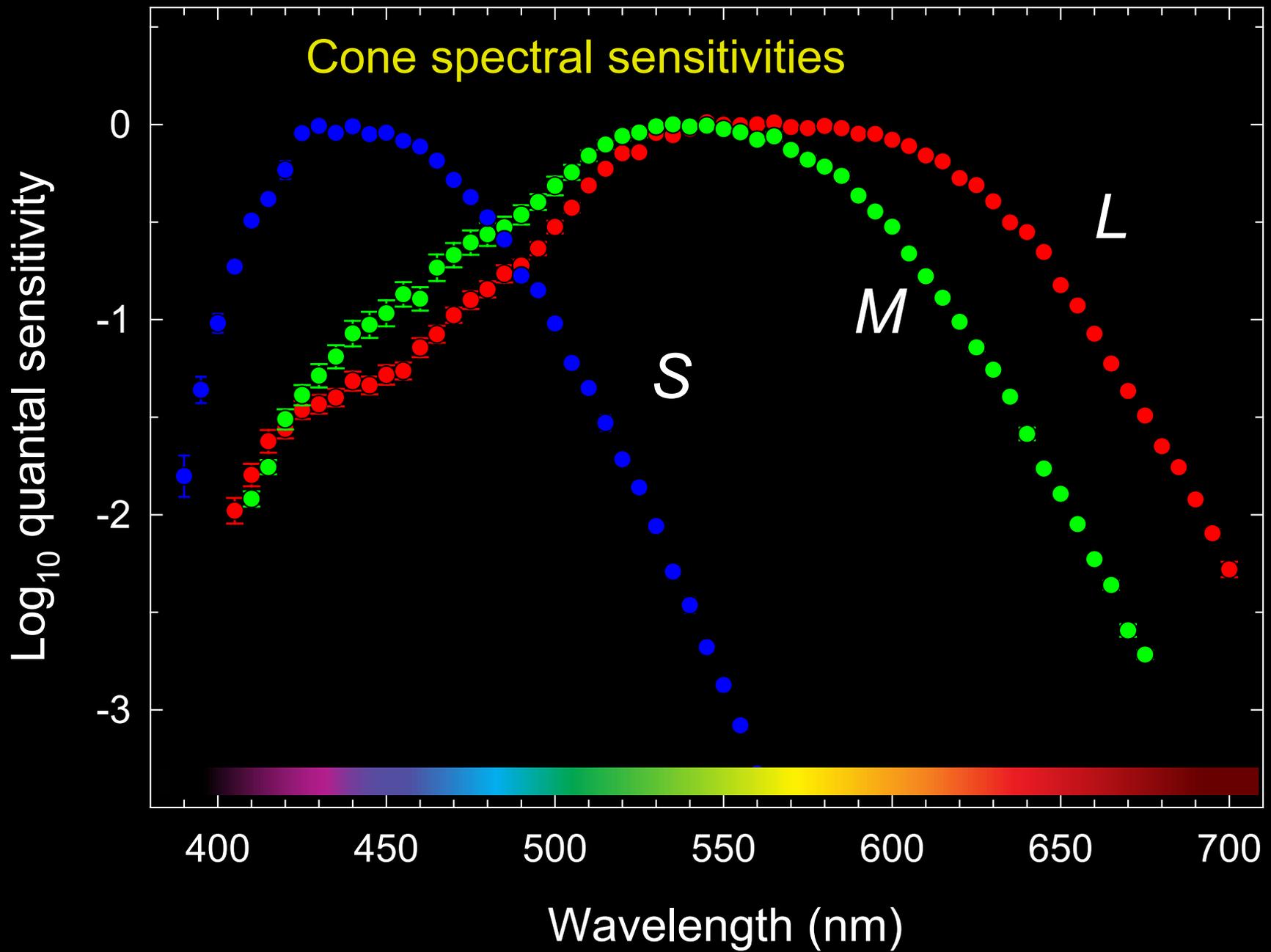
A consequence of univariance is that we are colour-blind when only one photoreceptor operates...



Examples: SCOTOPIC VISION, cone monochromacy

With three cone photoreceptors, our colour vision is trichromatic...

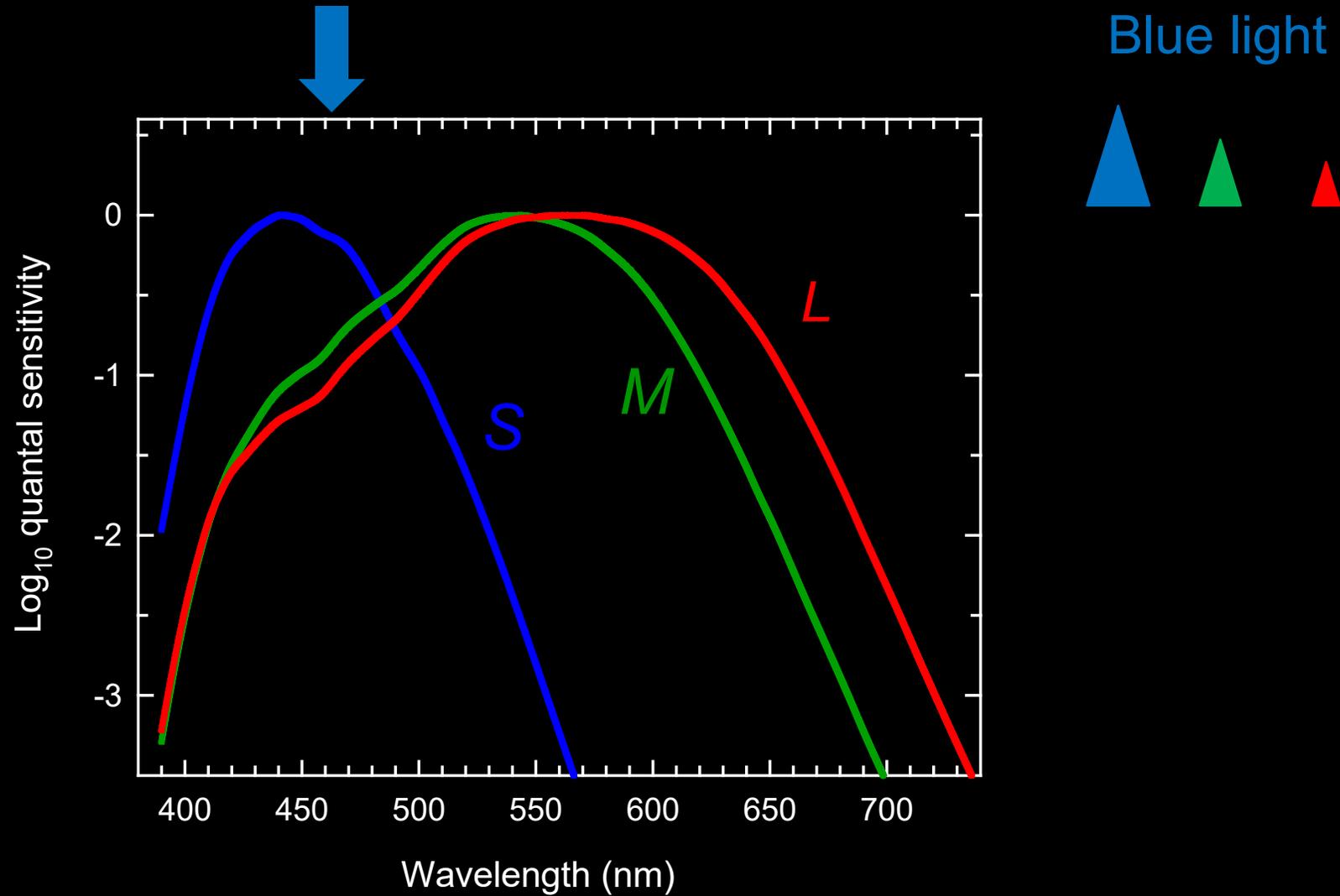




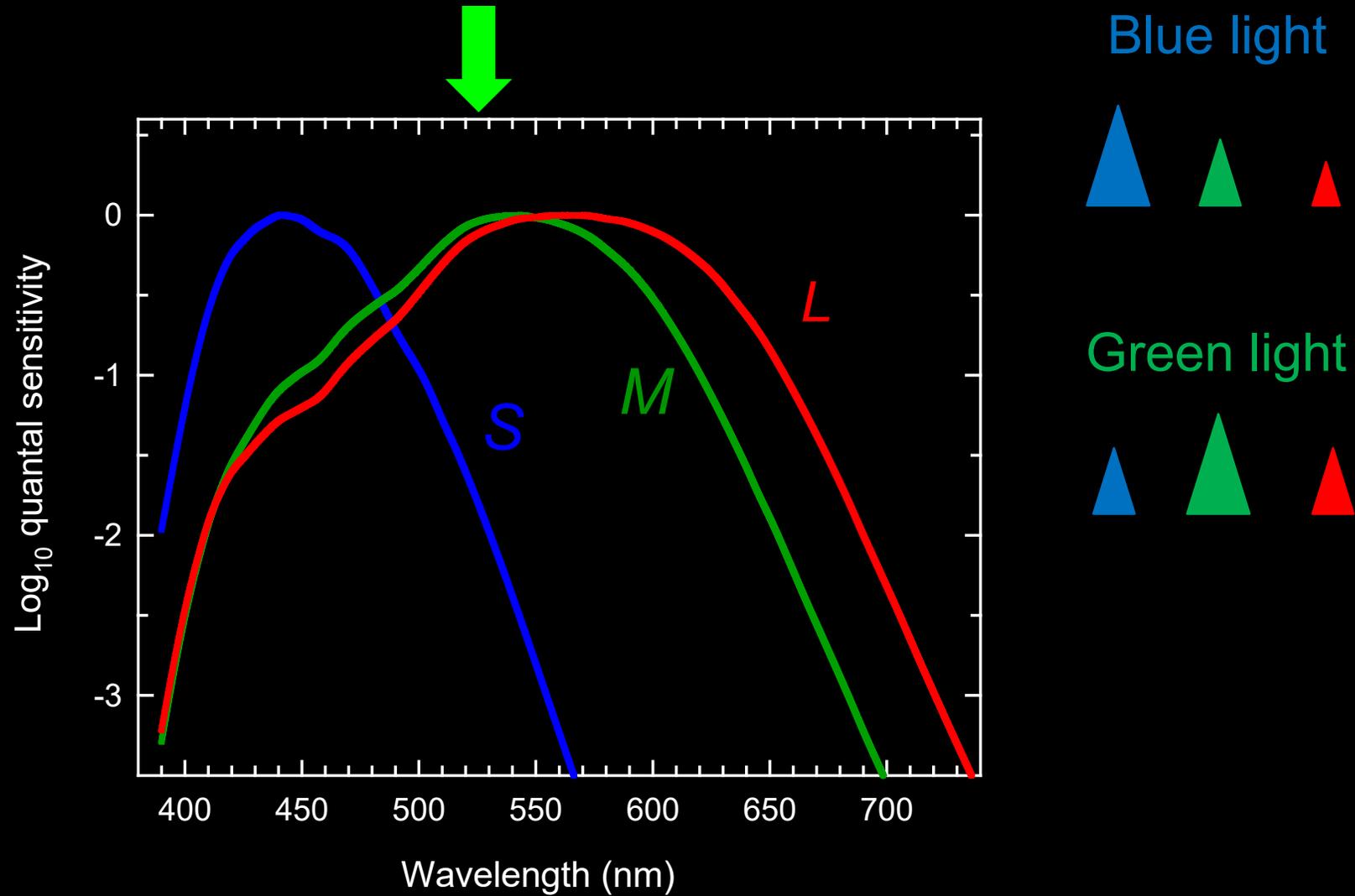
So, if each photoreceptor is colour-blind, how do we see colour?

Or to put it another way: How is colour encoded?

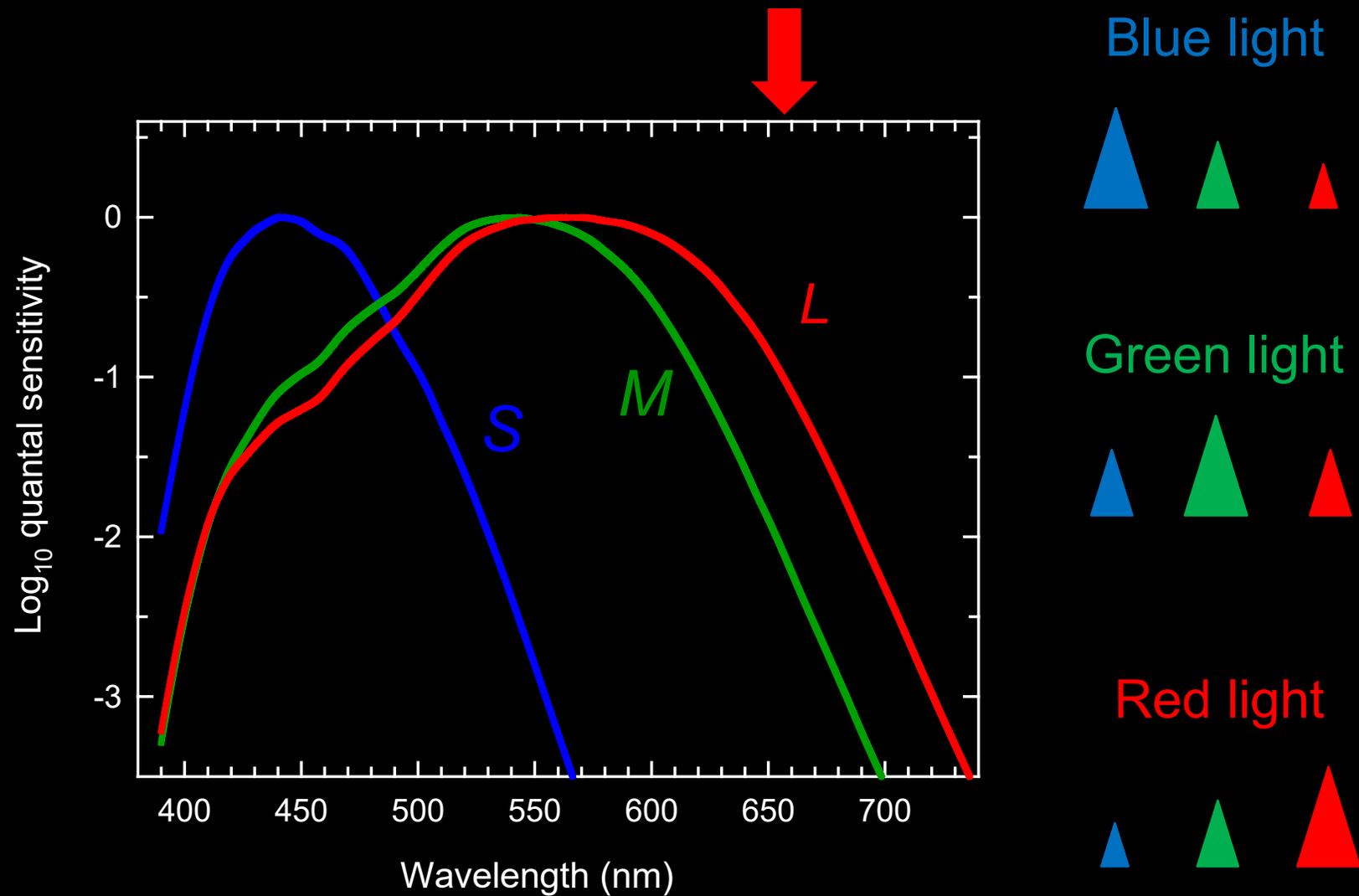
At the photoreceptors, colour is encoded by the relative cone outputs



Colour is encoded by the relative cone outputs



Colour is encoded by the relative cone outputs



Colour is encoded by the relative cone outputs

Blue light



Red light



Green light



Purple light



Yellow light



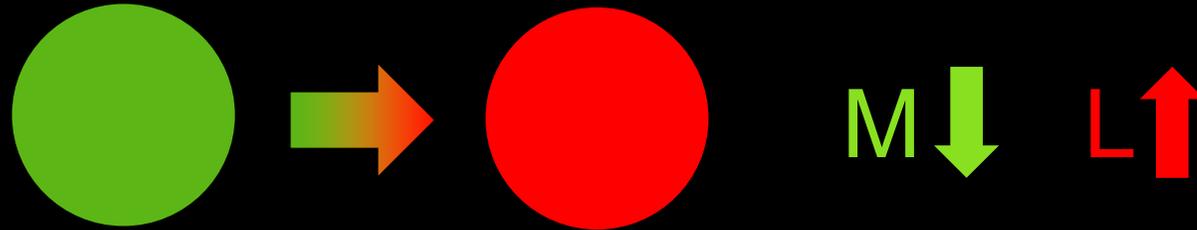
White light



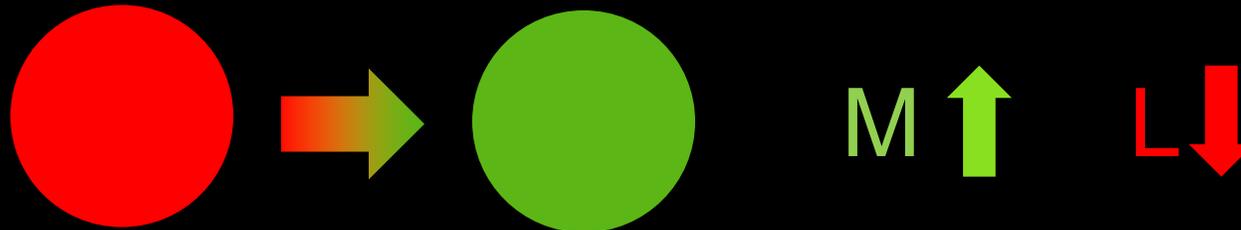
Because there are three univariant cones in the eye, human colour vision is a three-variable “trichromatic” system that depends on the relative outputs of the three cones.

TRICHROMACY

A change in colour from green to red causes a relative increase in the L-cone output but causes a decrease in the M-cone output.



A change in colour from red to green causes a relative increase in the M-cone output but causes a decrease in the L-cone output.



Thus, colour can be encoded by *comparing* the outputs of different cone types...

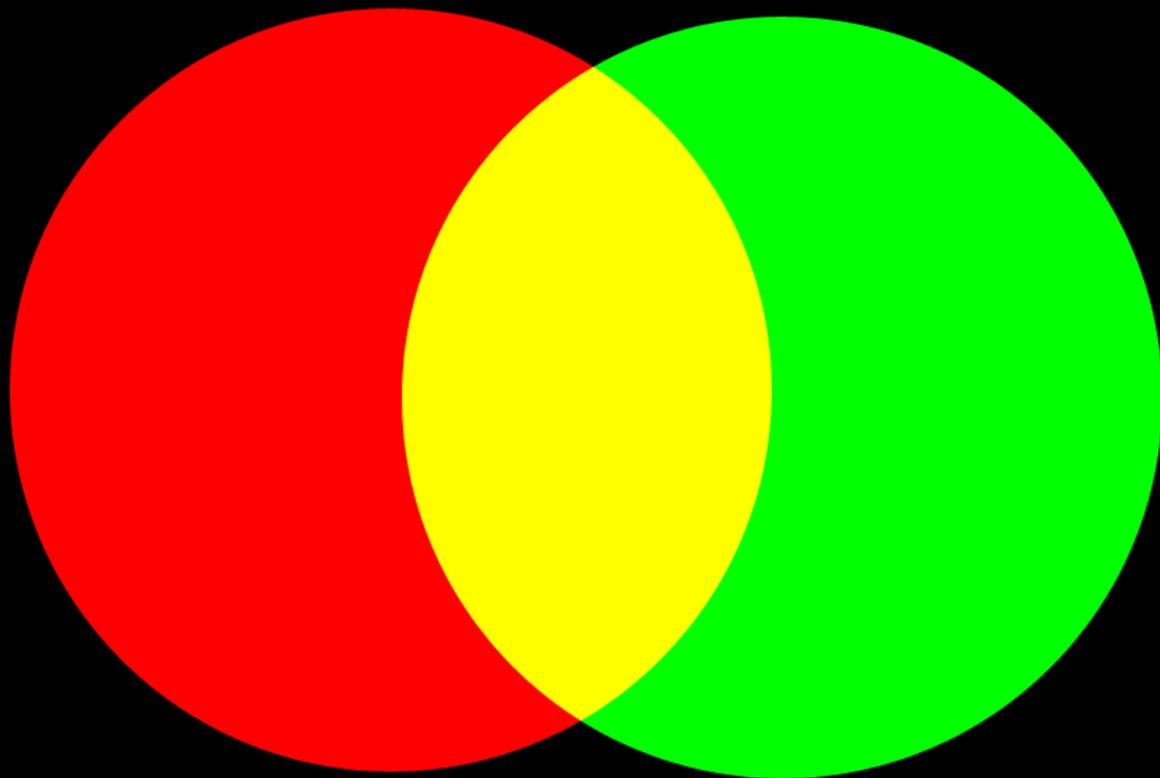
TRICHROMACY

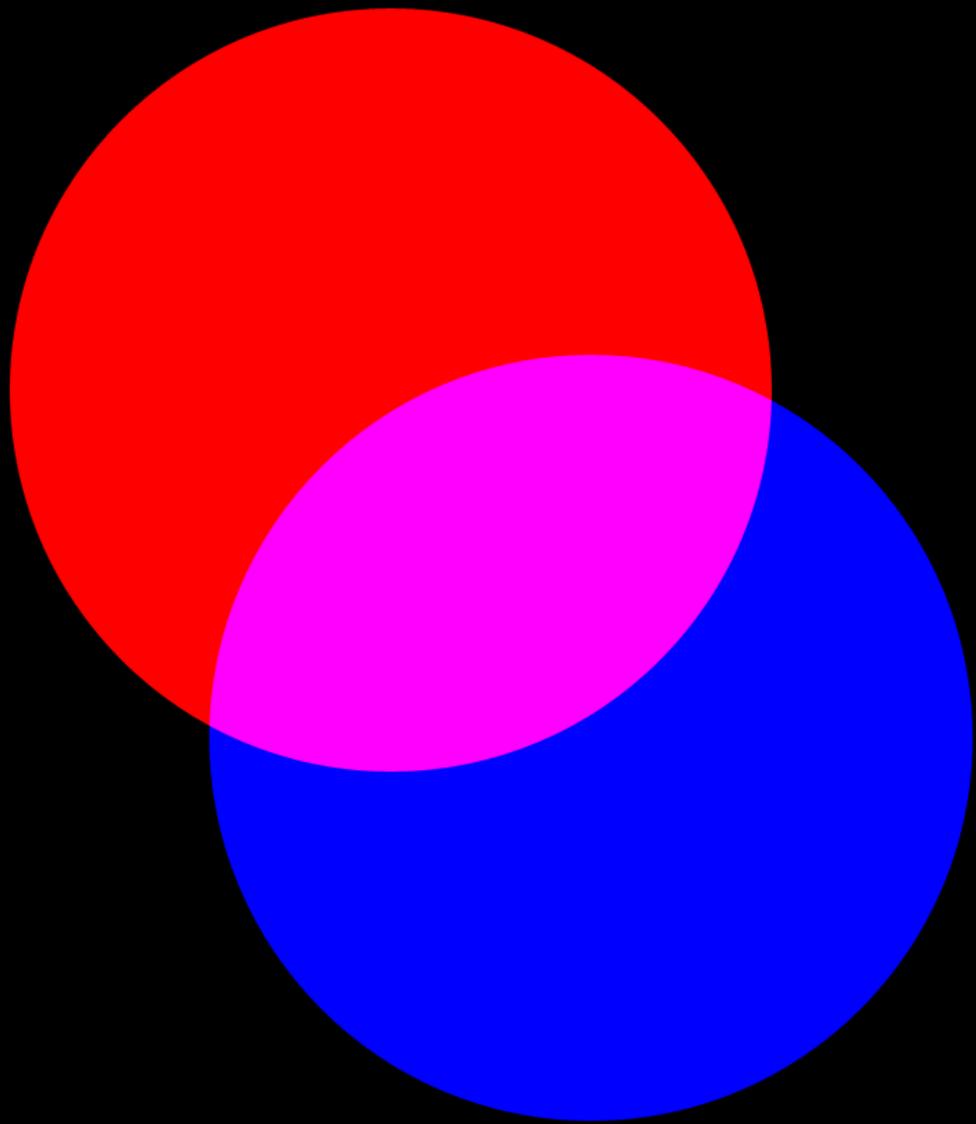
Because we have just three univariant cones, coloured lights are entirely defined by the three cone excitations they produce.

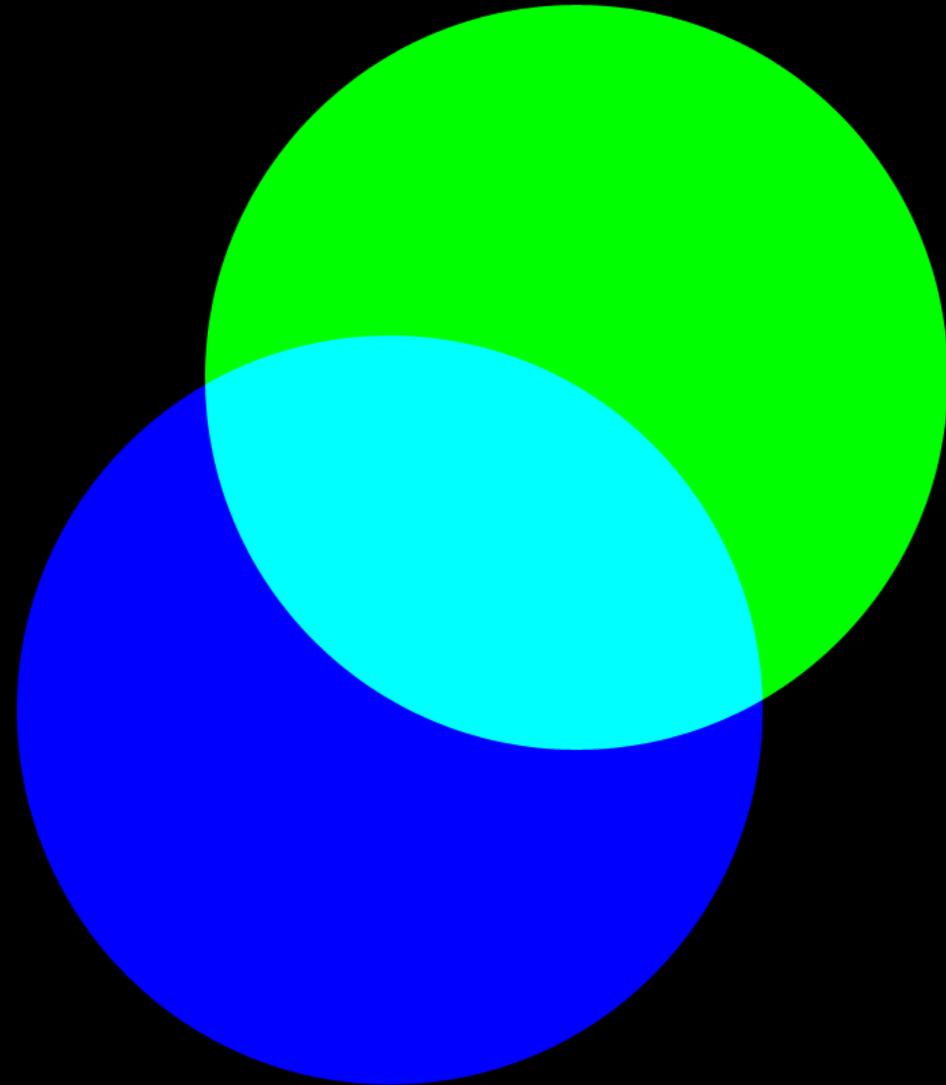
Any pairs of lights that produce the *same* triplet of excitations must be indistinguishable.

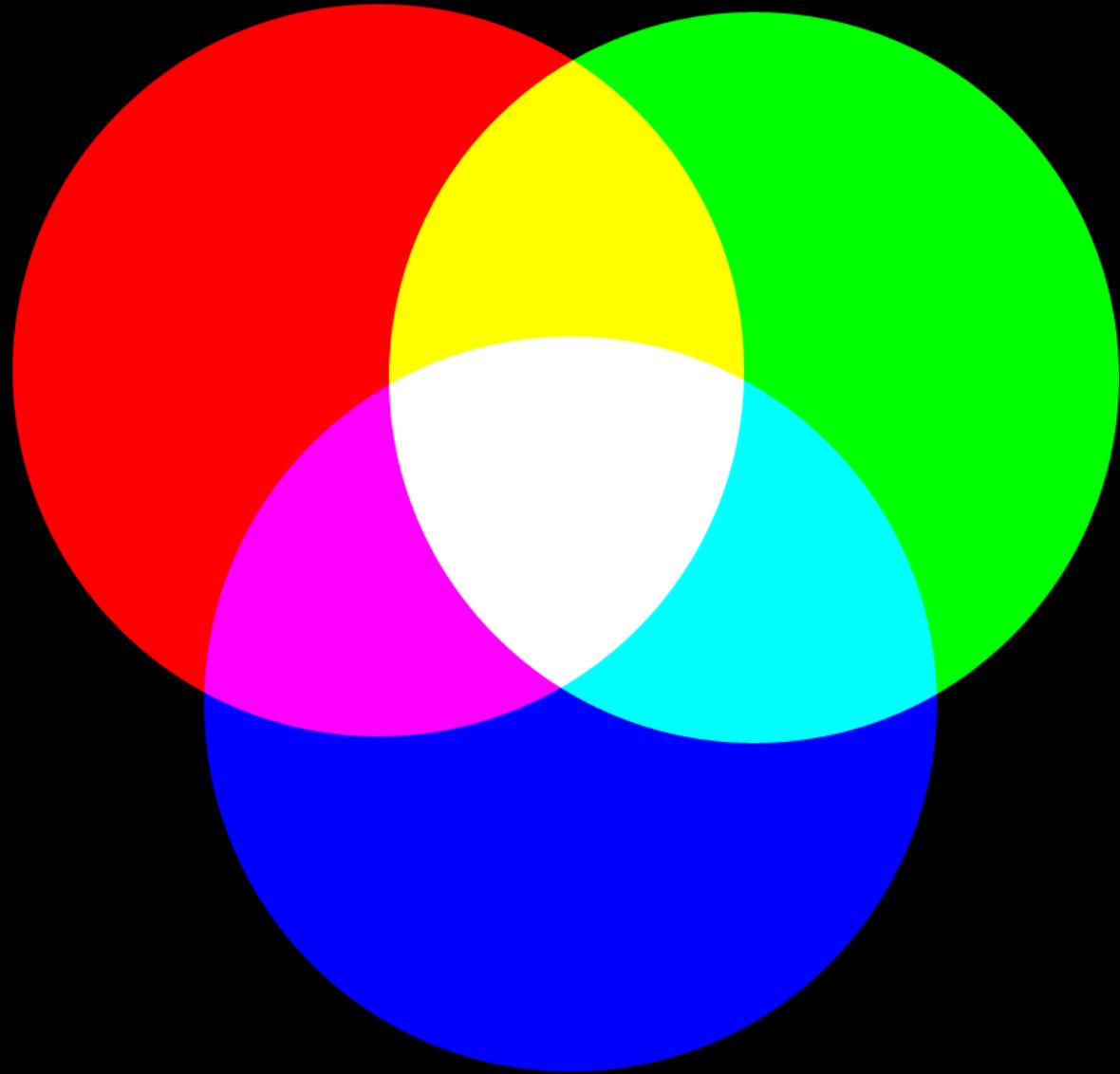
Pairs of lights that are physically different but indistinguishable are known as “metamers”.

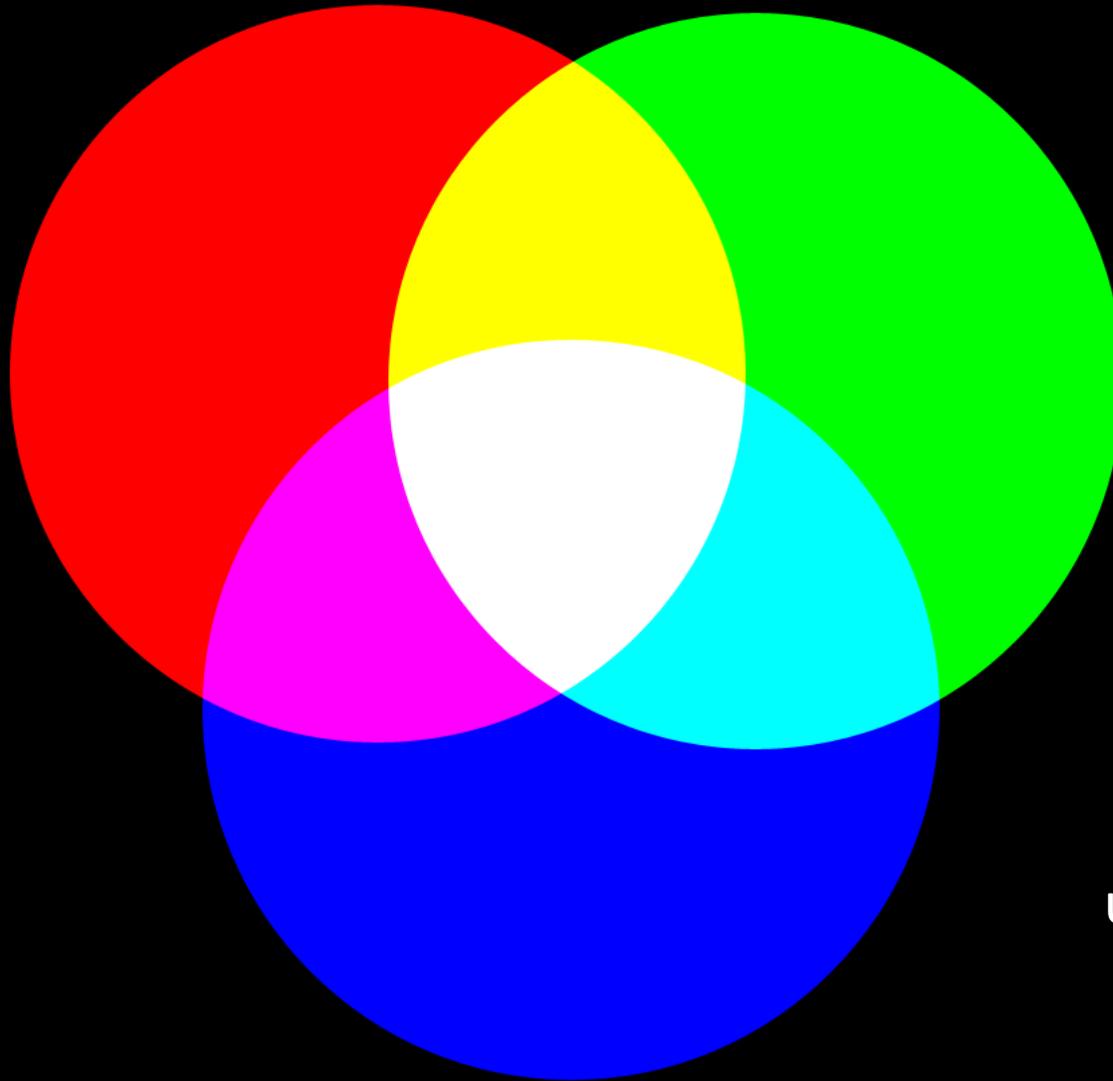
There are many metamers...



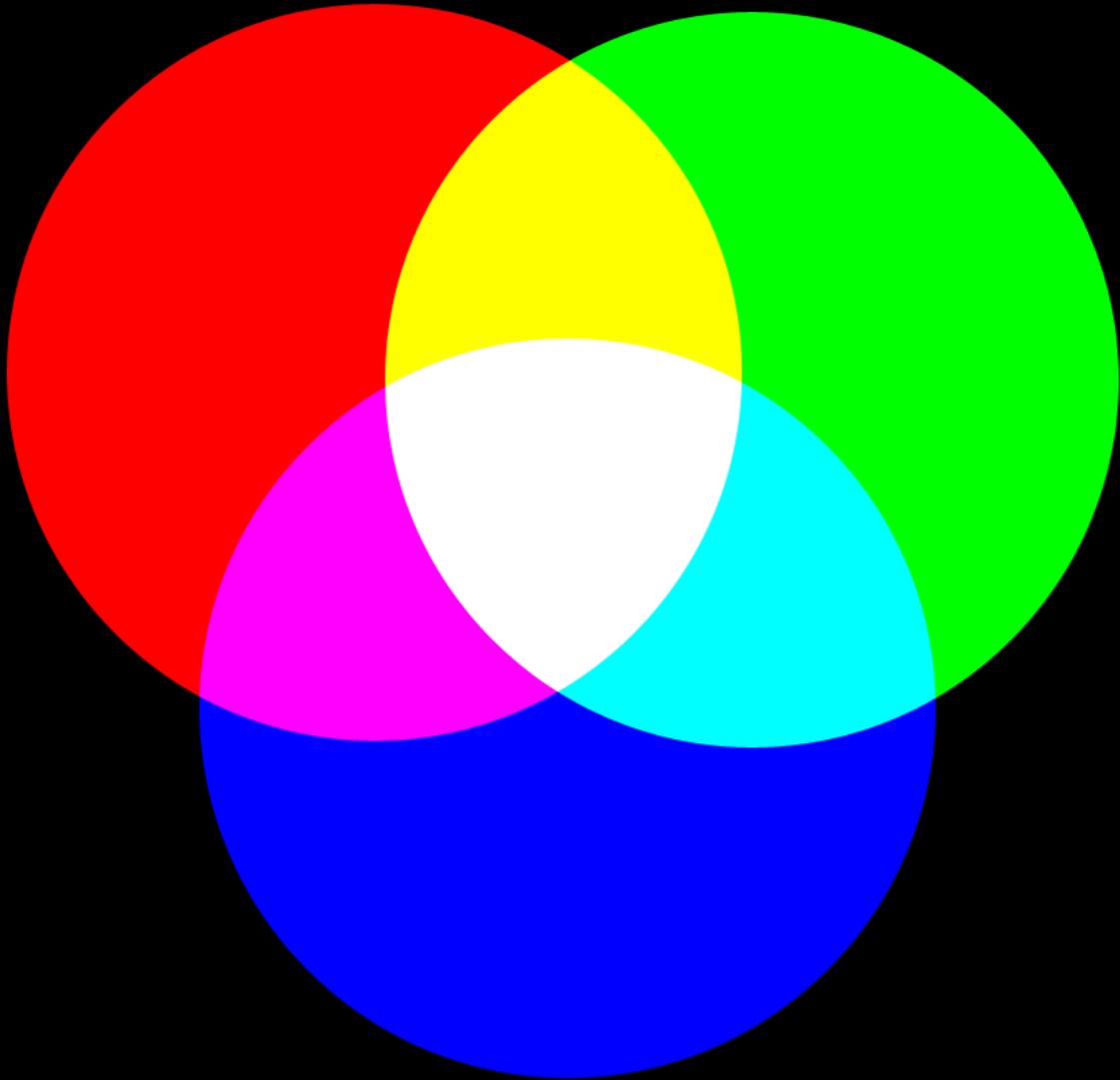






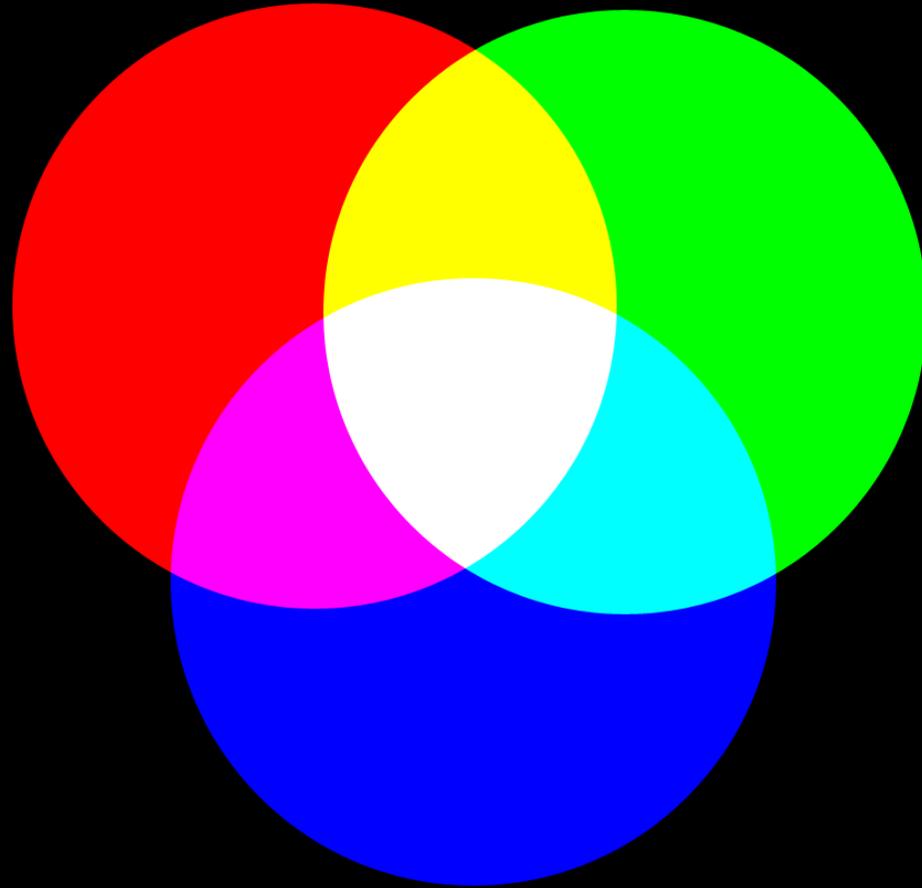


Before we knew about the underlying biology, additive colour mixing done in the 19th century revealed that colour vision was...



TRICHROMATIC

Trichromacy means that colour vision at the input to the visual system is relatively simple.

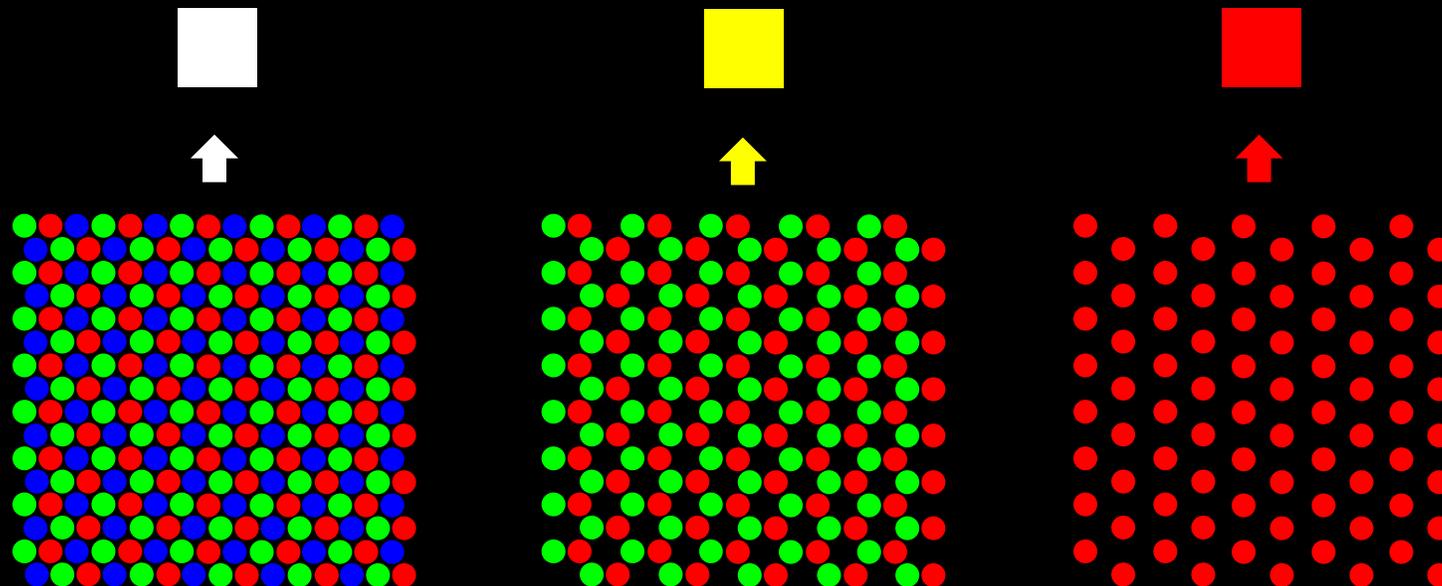


It is a 3 variable system...

Colour TV

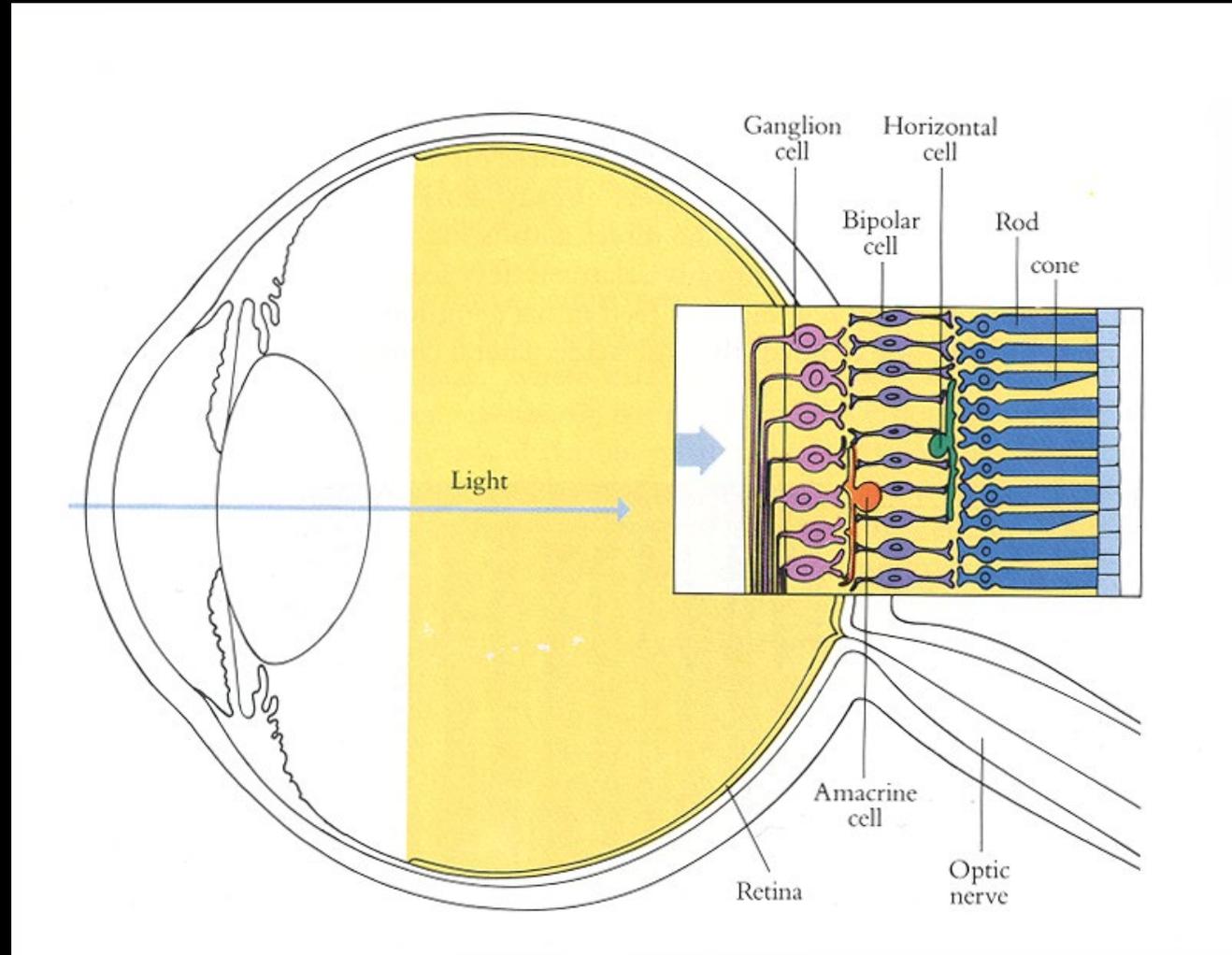
Trichromacy is exploited in colour reproduction, since the myriad of colours perceived can be produced by mixing together small dots of three colours.

The dots produced by a TV or projector are so small that they are mixed together by the eye and thus appear as uniform patches of colour.



POSTRECEPTORAL
COLOUR VISION

But what happens next (i.e., how is colour encoded after the photoreceptors)?

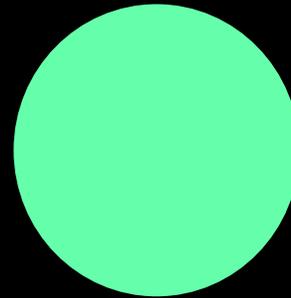


Colour phenomenology

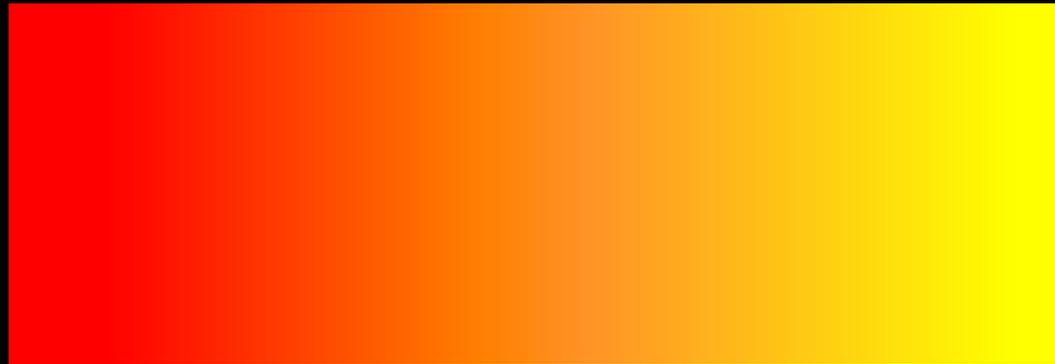
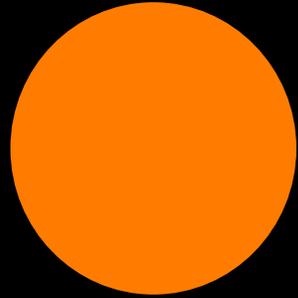


Can provide clues about how colours are encoded after the photoreceptors...

Imagine a single patch of colour inside a dark surround

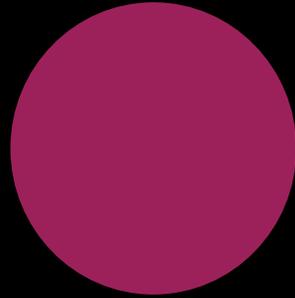


- ▶ Which pairs of colours can coexist in a single, uniform patch of colour?
- ▶ Which pairs can never coexist?



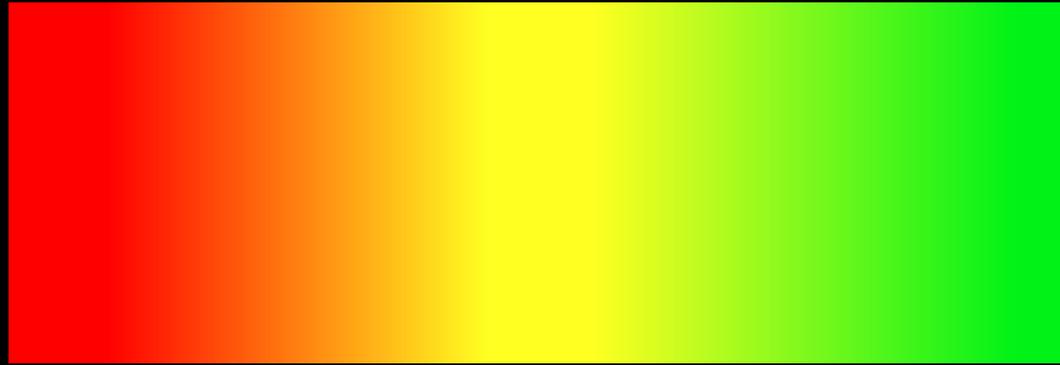
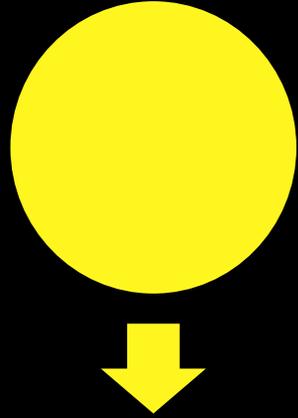
Can a single patch be reddish-yellow?



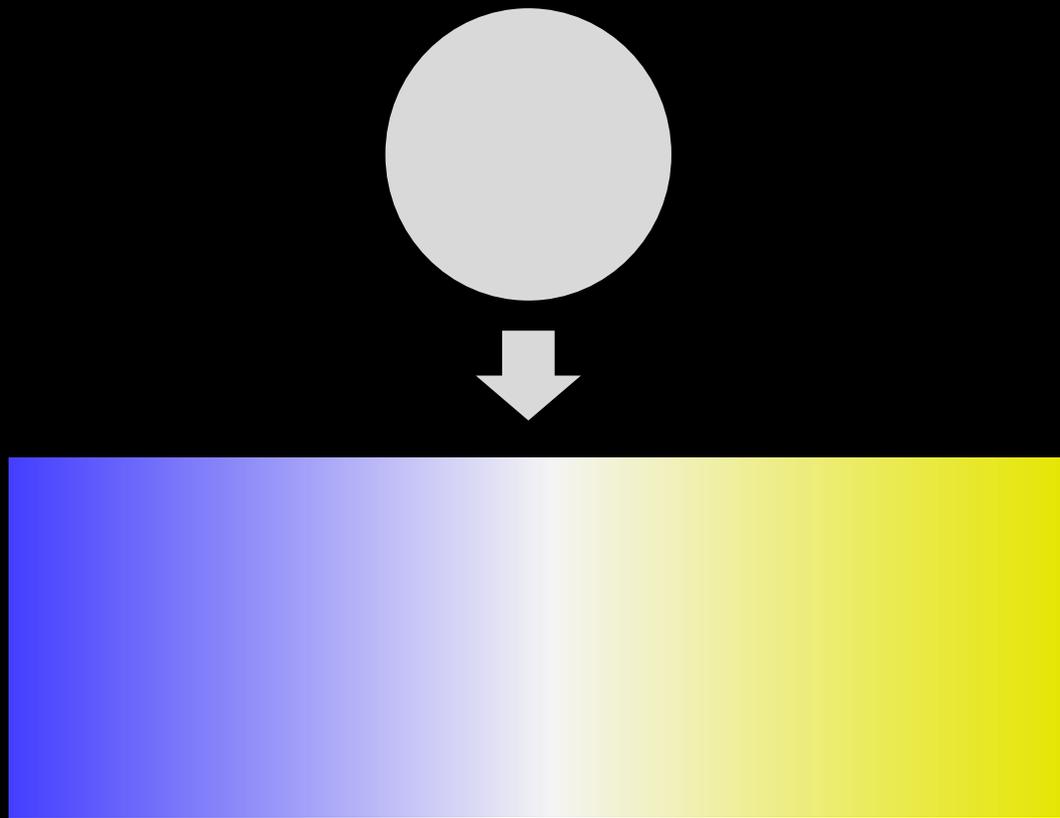


Can it be reddish-blue?



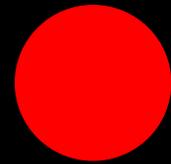


Can it be reddish-green? 

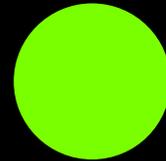


Can it be bluish-yellow? **X**

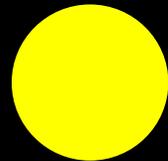
The colour opponent theory of Hering



is opposed to



R-G

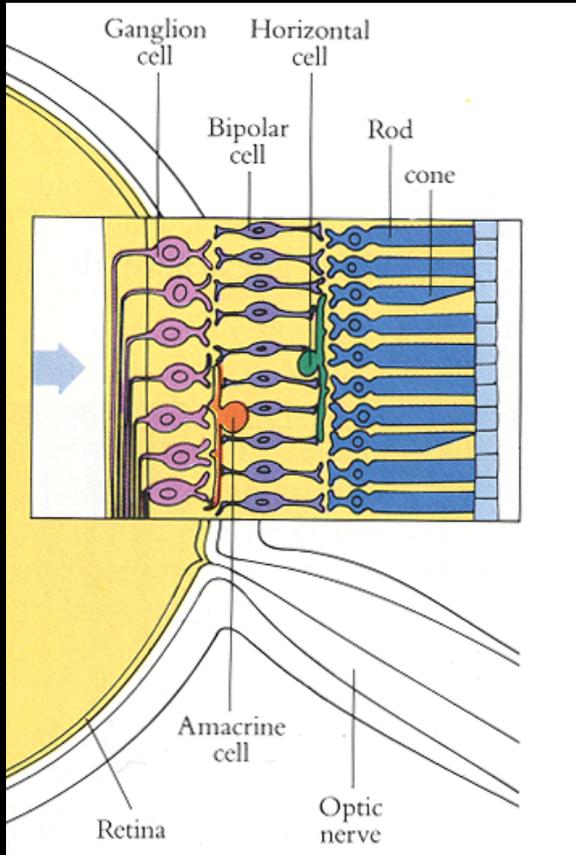


is opposed to

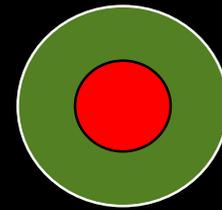


Y-B

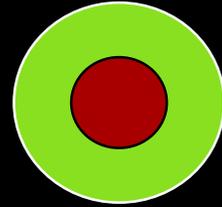
And indeed cells in the early visual pathway oppose the signals from different cone classes and can be loosely classified as “red-green” or “blue-yellow” opponent.



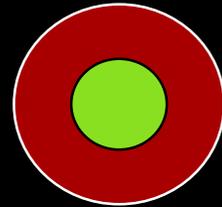
“R-G”



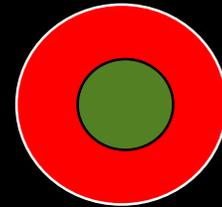
+L-M



-L+M

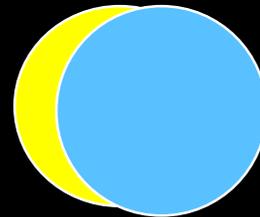


+M-L



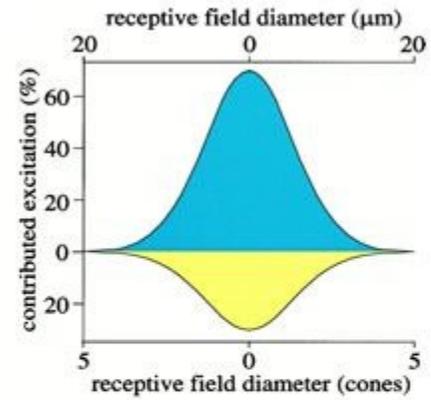
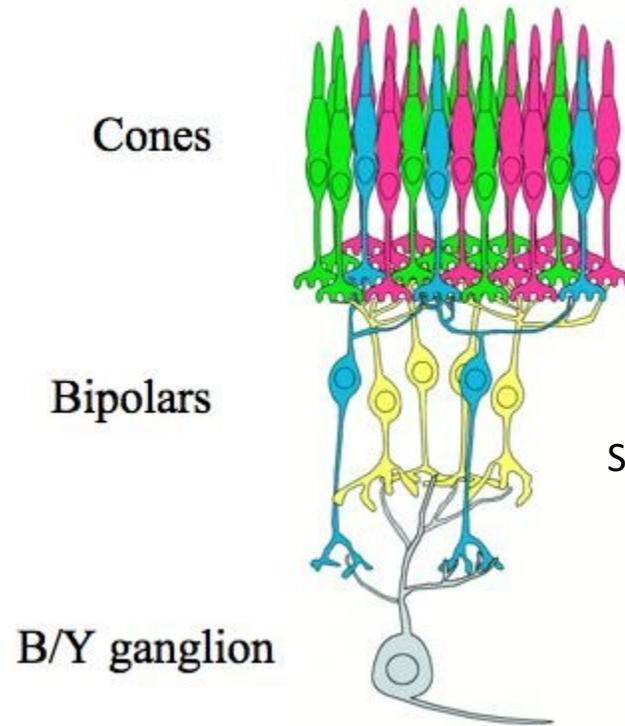
-M+L

“B-Y”

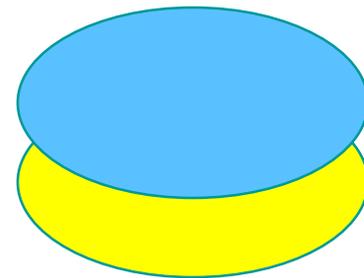


+S-[L+M]

Blue/yellow pathway



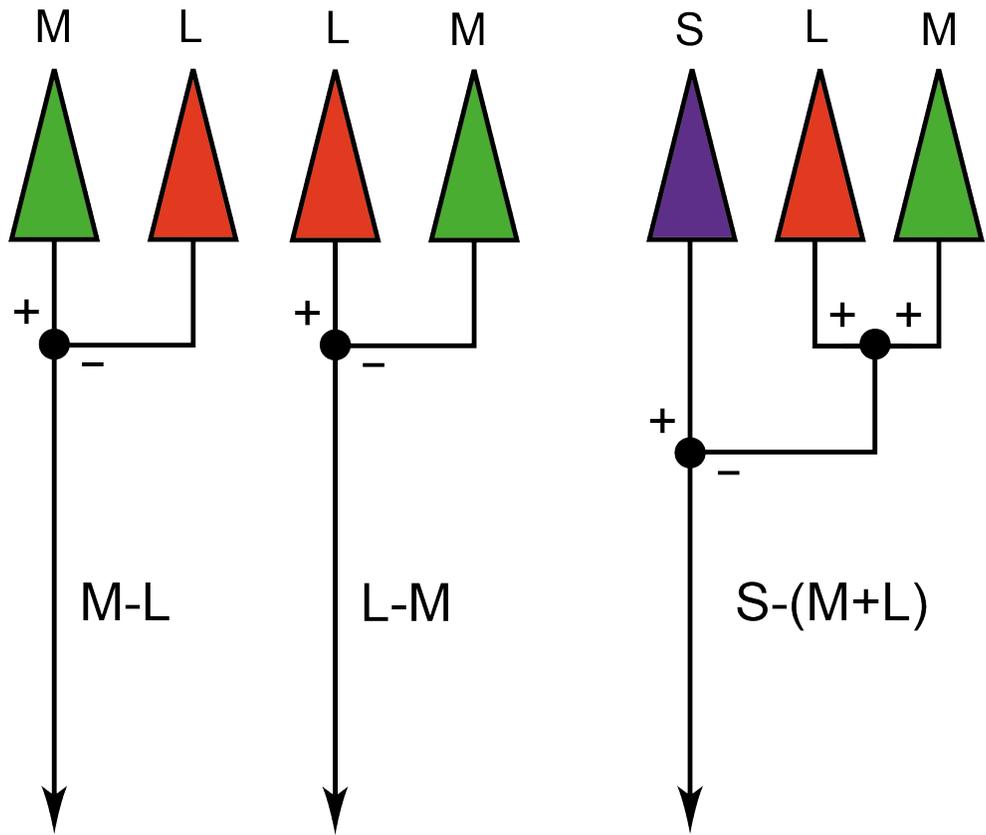
Source: David Heeger



“Koniocellular”
pathway

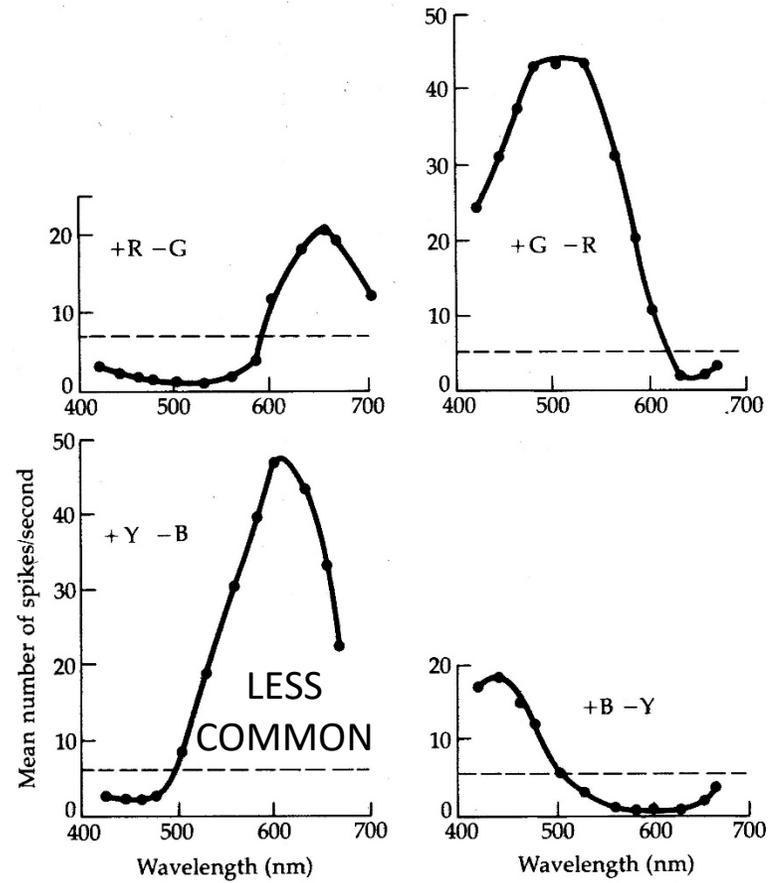
Parvocellular pathways

Koniocellular pathway



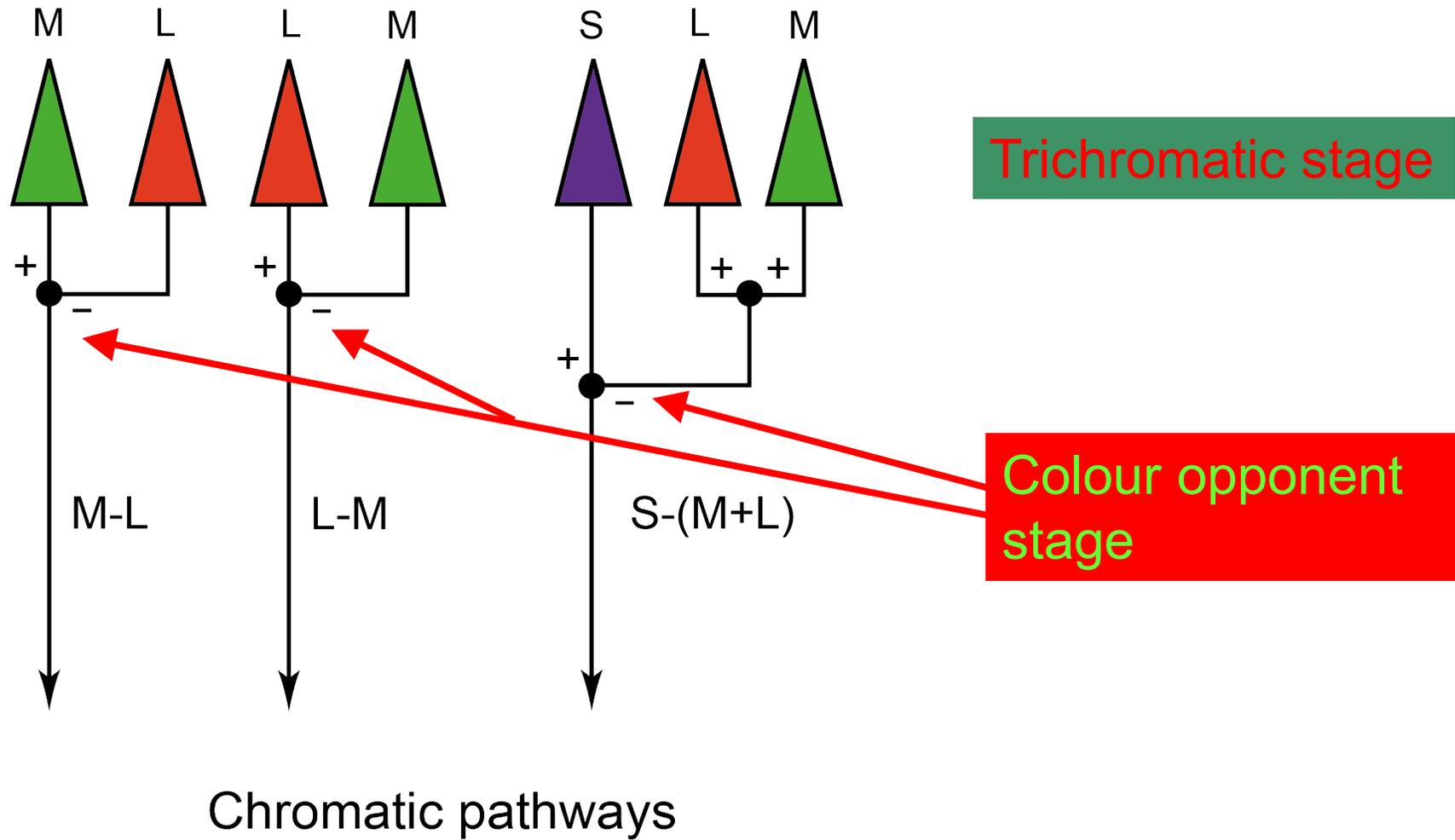
Chromatic pathways

LGN cell responses



8 AVERAGE FIRING RATES of large sample of cells of each of six LGN cell types as a function of wavelength. Top four cells are spectrally opponent ones and bottom two are spectrally nonopponent cells. The cells on the left are, in principle, "mirror images" of those on the right.

Summary

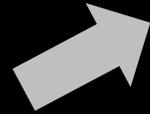


So we've talked about colour
(chromatic) vision, but what about
"luminance" (achromatic) vision?

Colour...



Split the image into...



ACHROMATIC COMPONENTS



CHROMATIC COMPONENTS



CHROMATIC COMPONENTS



By itself chromatic information provides relatively limited information...

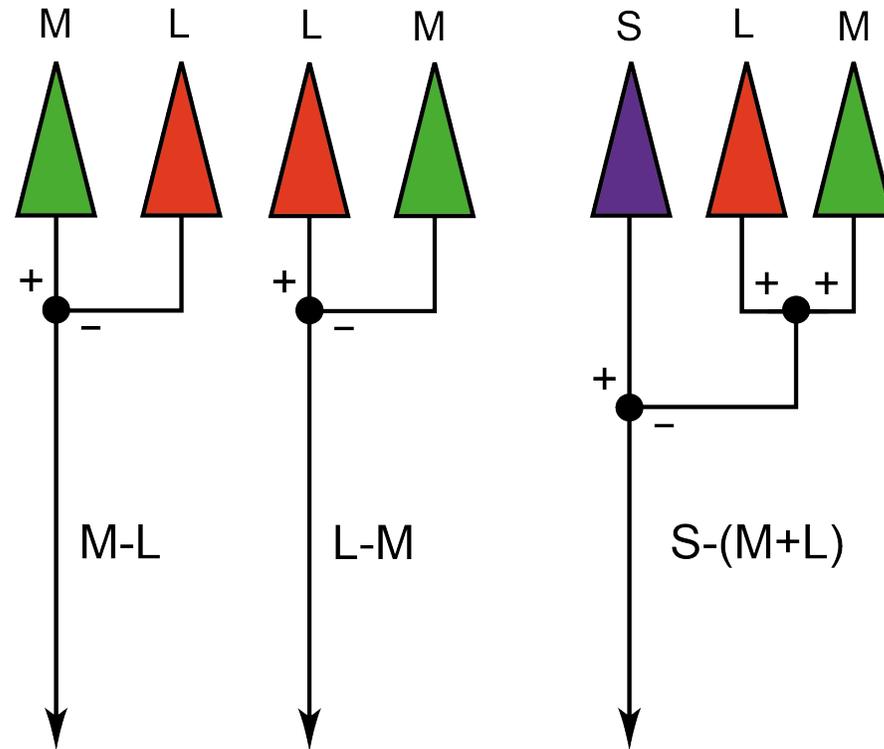
ACHROMATIC COMPONENTS



Achromatic information important for fine detail ...

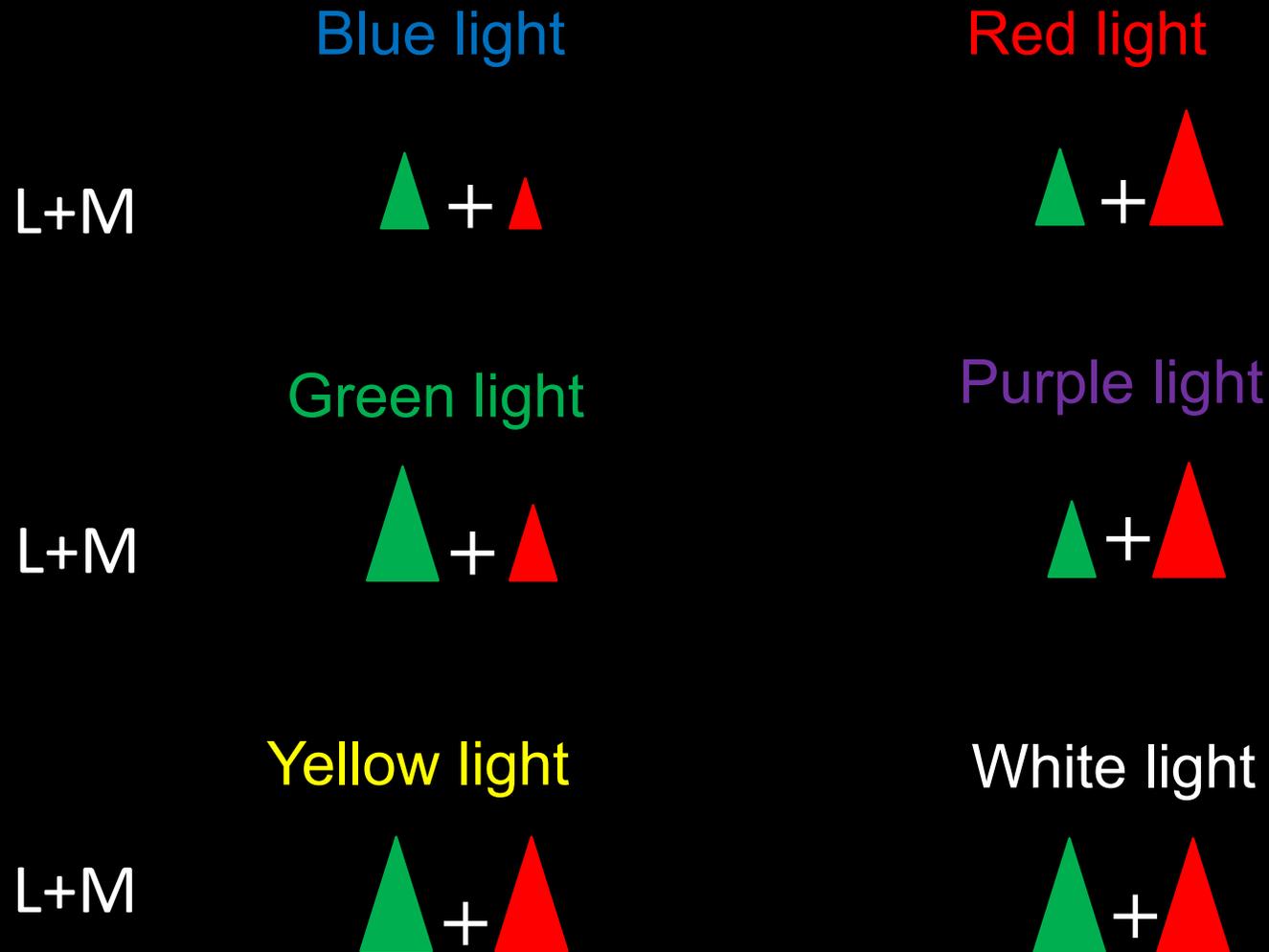
Achromatic and chromatic cone
vision (colour and luminance)

In addition to neural pathways that signal colour there are also pathways that signal intensity or luminance:



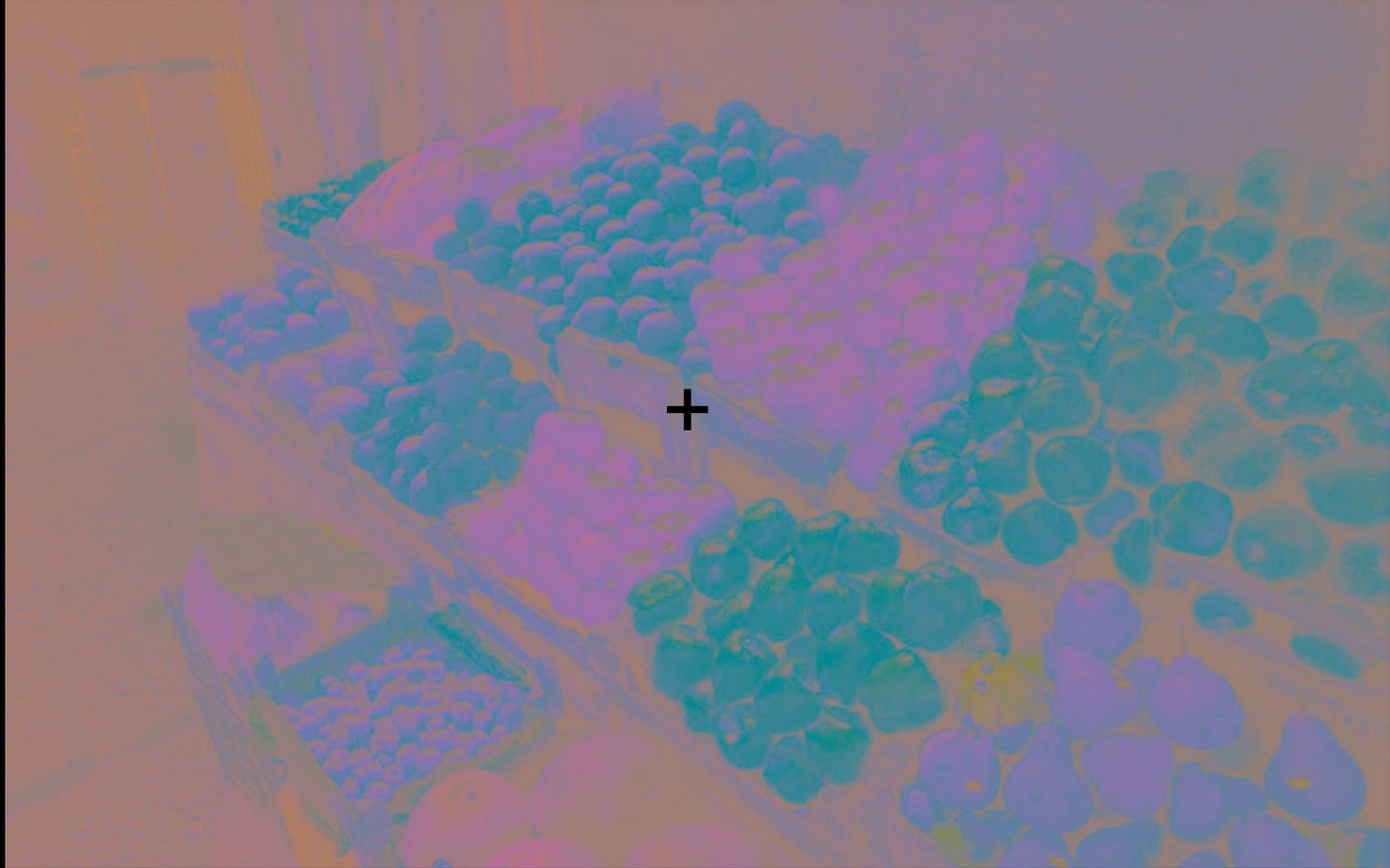
Chromatic pathways

Luminance is encoded by **summing** the L- and M-cone signals:

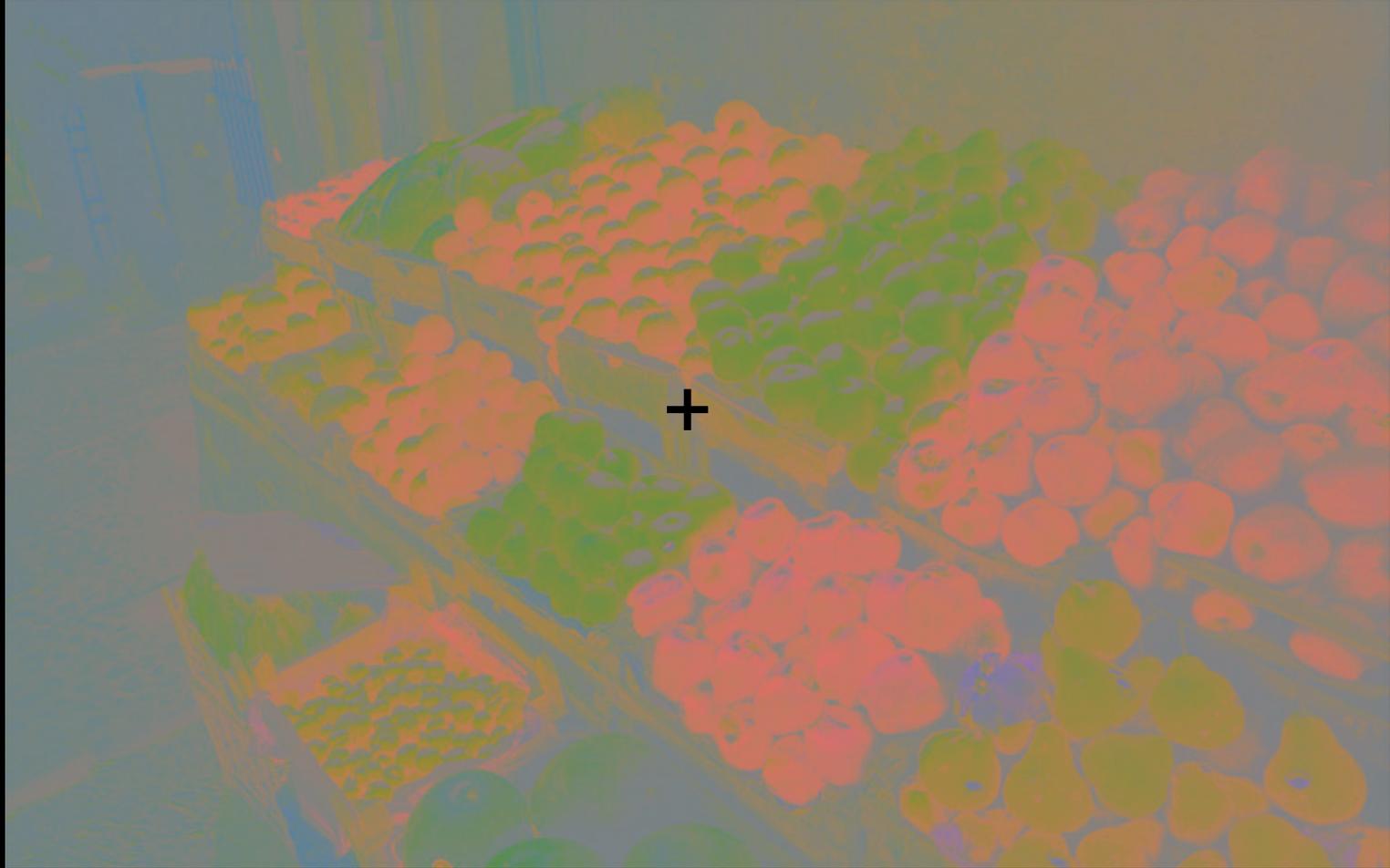


Colour is in many ways secondary to
luminance



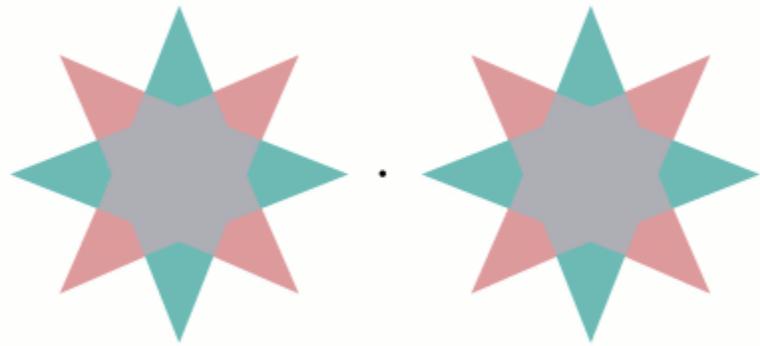






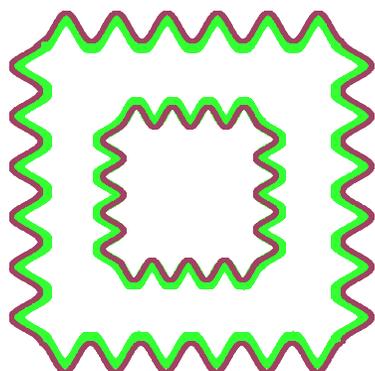
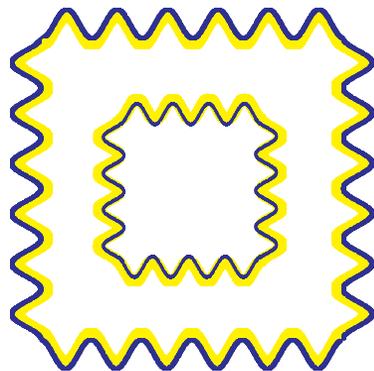
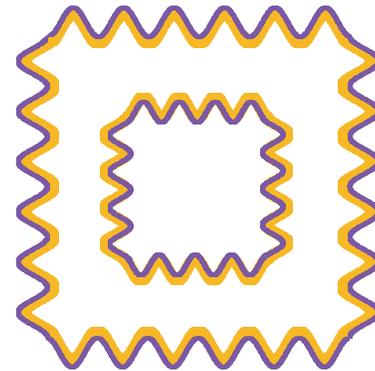
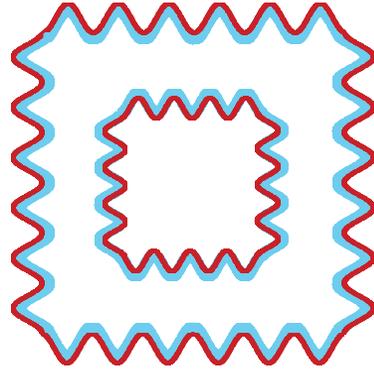




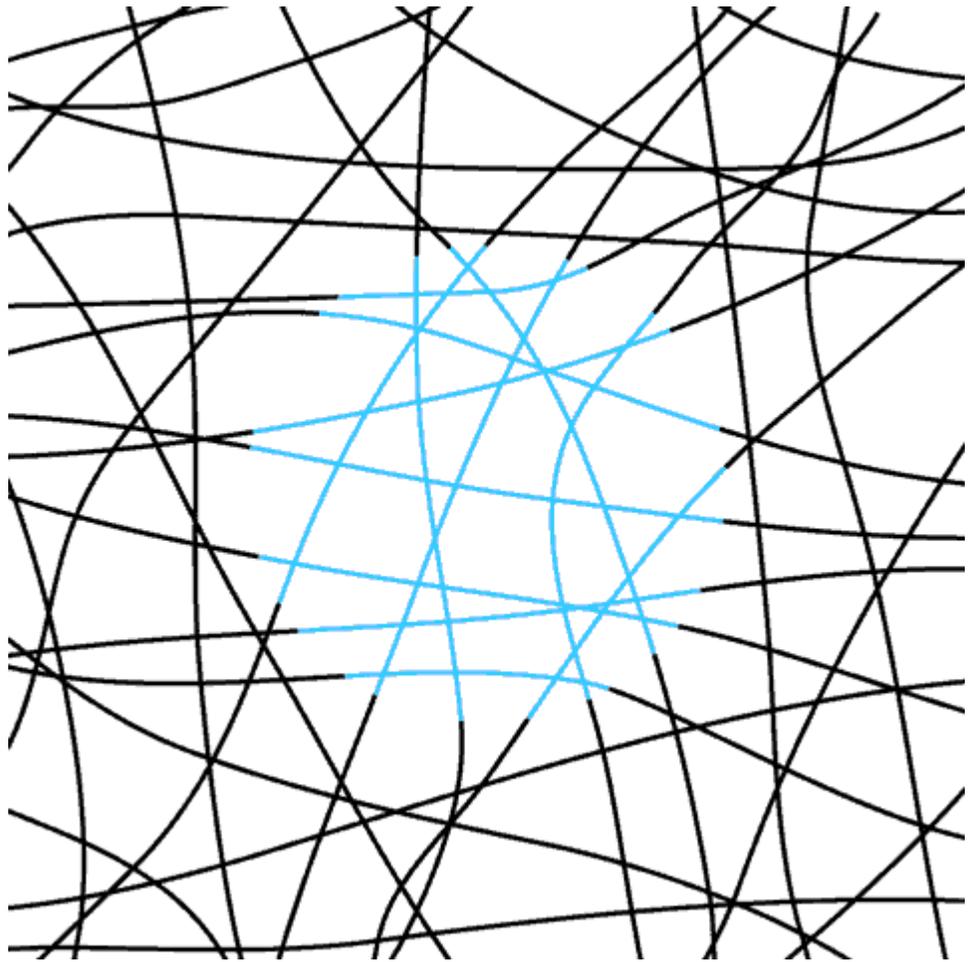


Rob van Lier, Mark Vergeer & Stuart Anstis

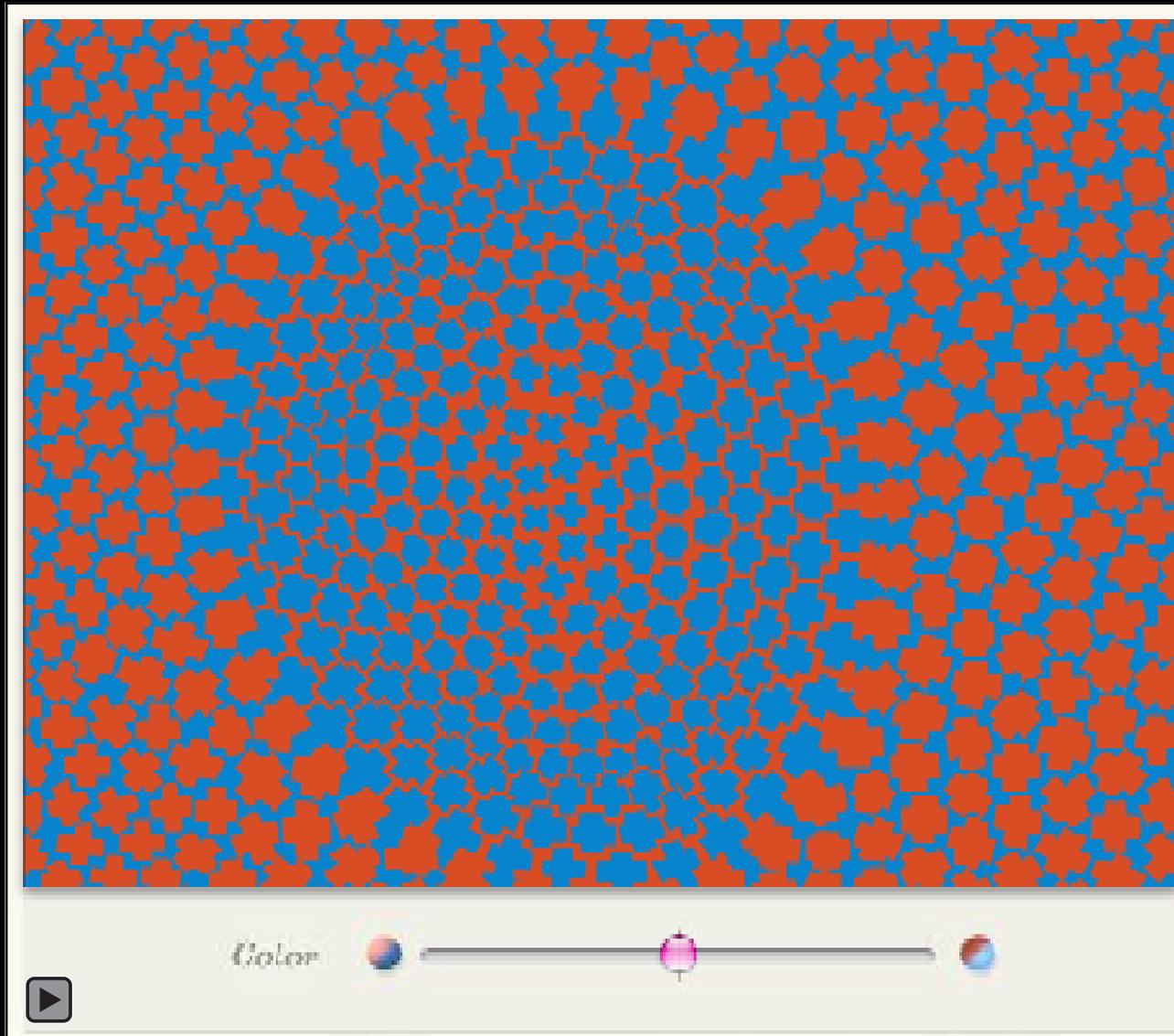
Watercolour
effect



Neon Spreading

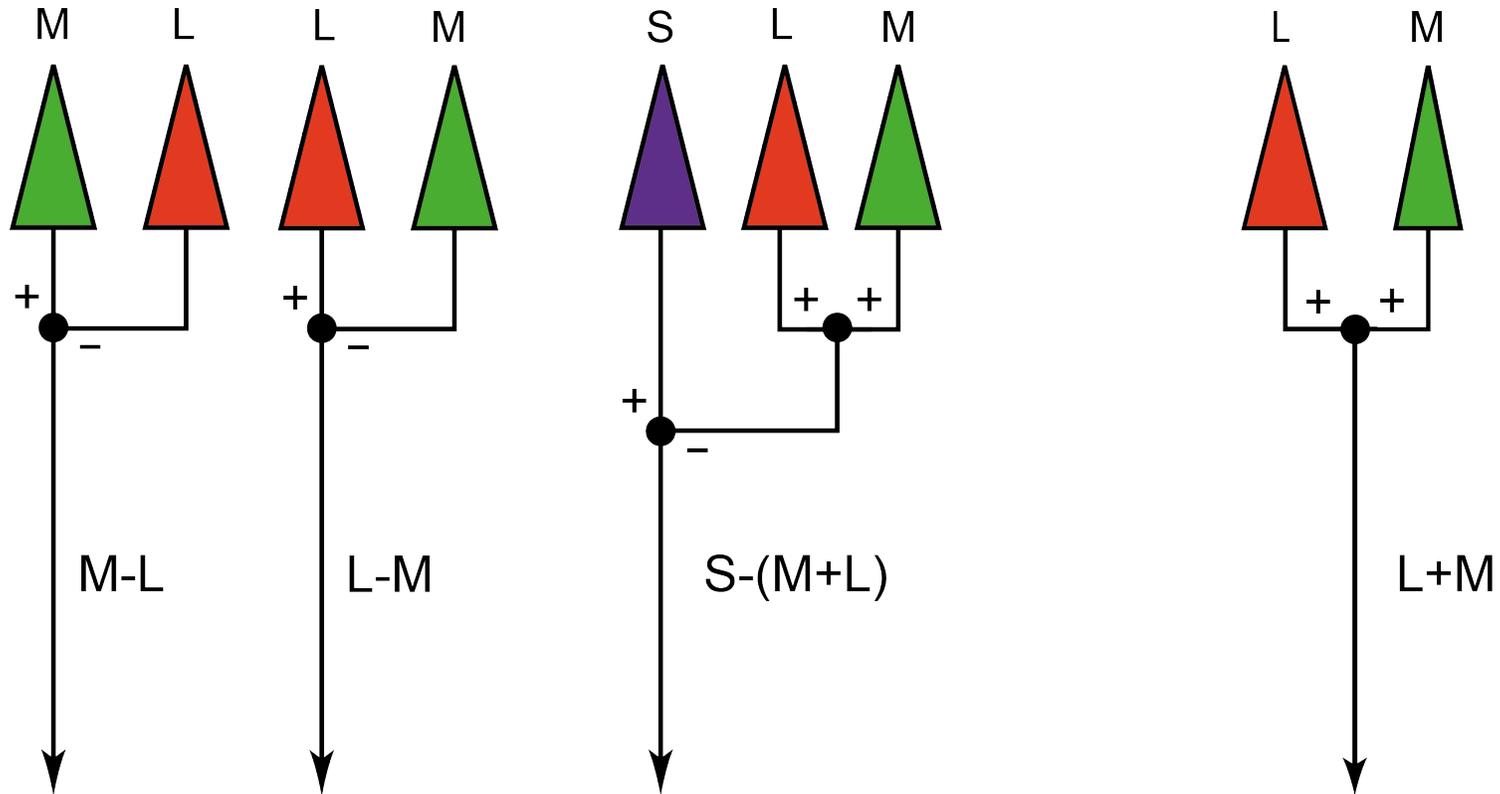


Interesting artistic effects occur when vision depends only on colour (and not on luminance)



'Plus Reversed', Richard Anuszkiewicz, 1960

What are the postreceptoral neural substrates of the chromatic and luminance pathways?

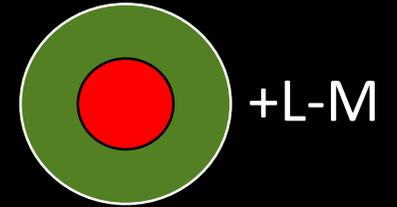
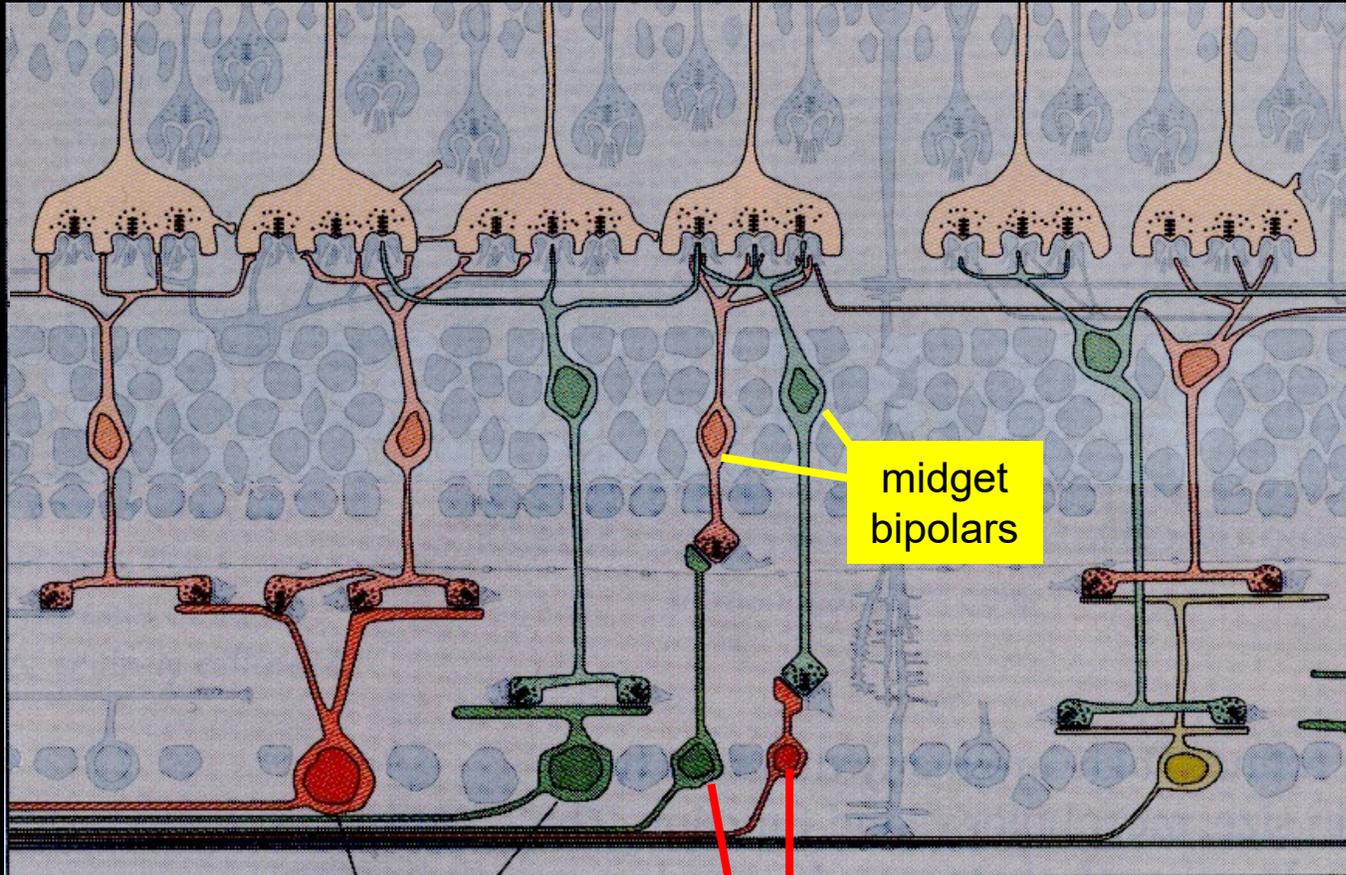


Chromatic pathways

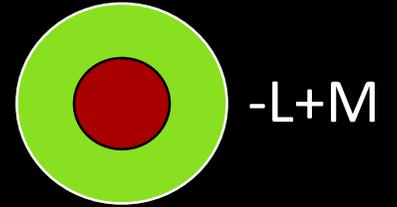
Luminance pathway

Red-green chromatic pathways have been linked to the parvocellular retinal stream for L-M.

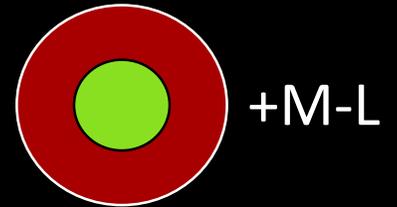
Parvocellular



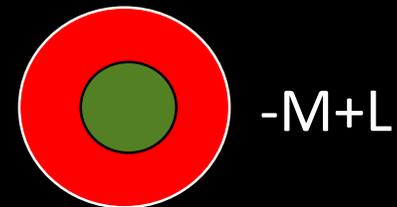
+L-M



-L+M



+M-L



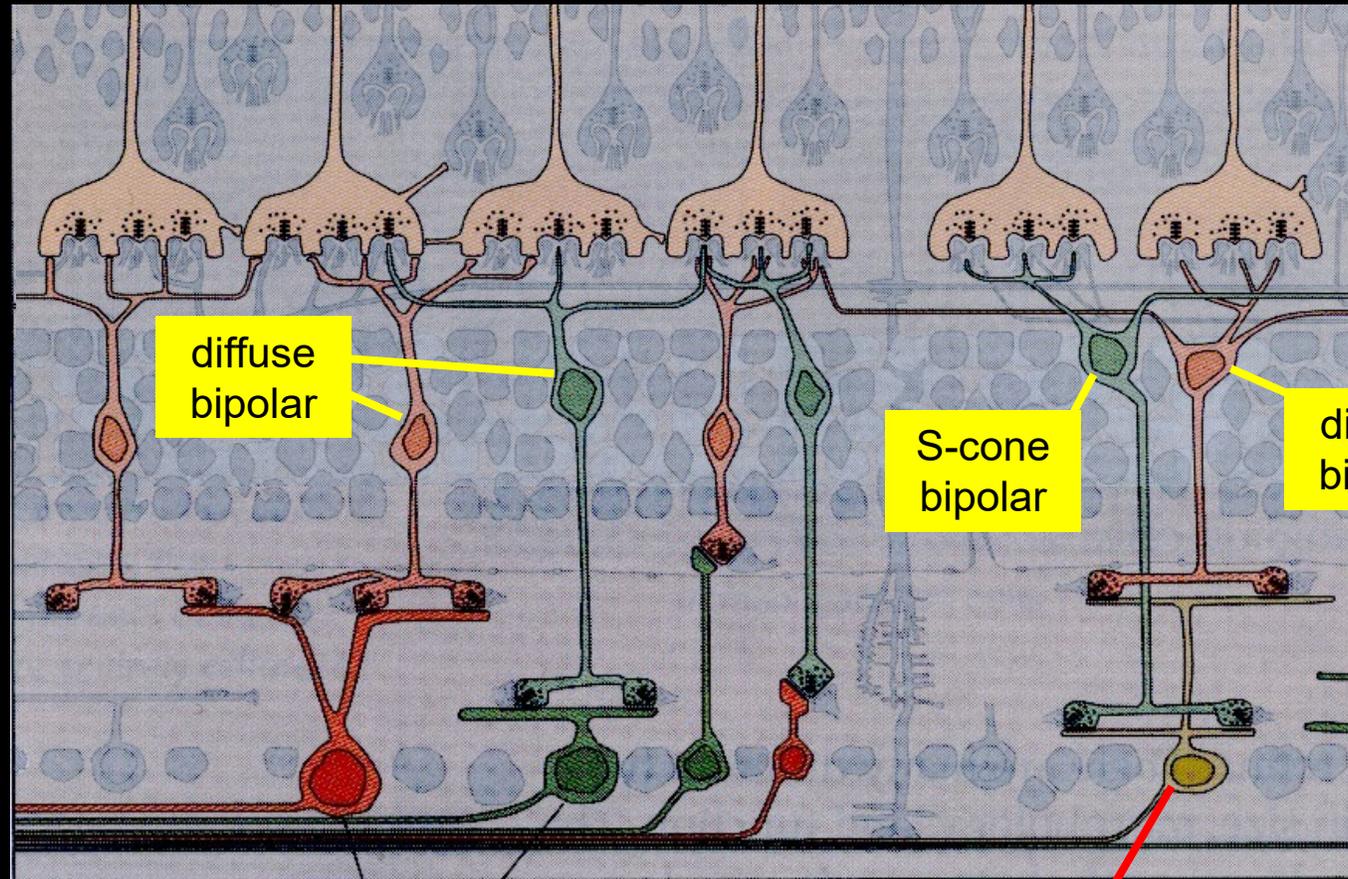
-M+L

midget
ganglion cell

From Rodieck (1998)

Blue-yellow chromatic pathways have been linked to the koniocellular stream...

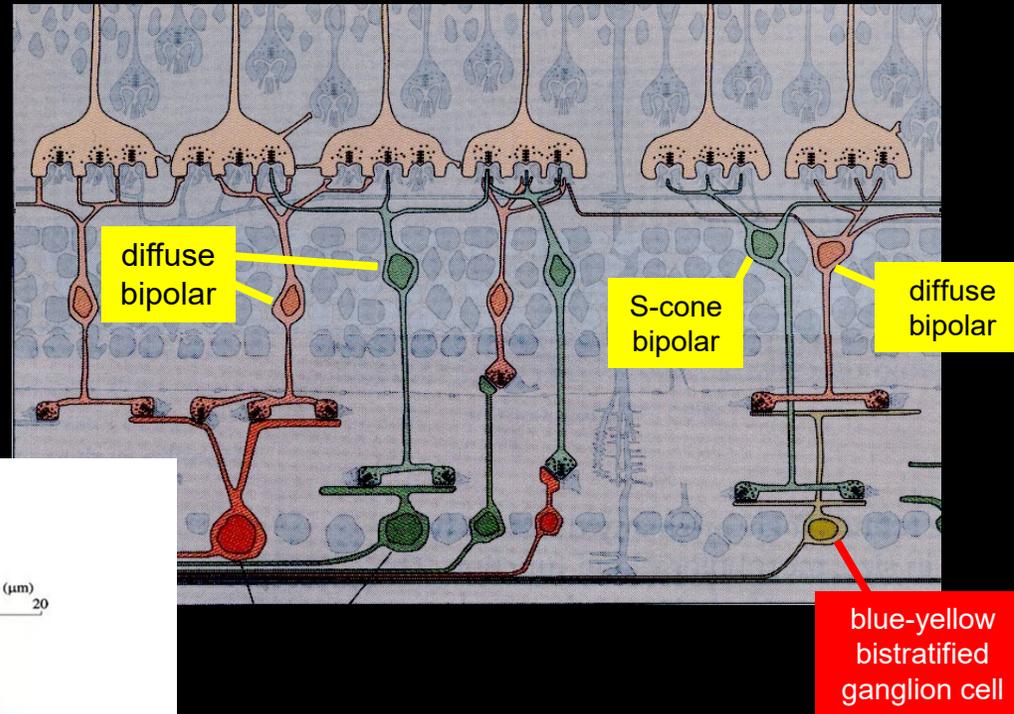
Koniocellular



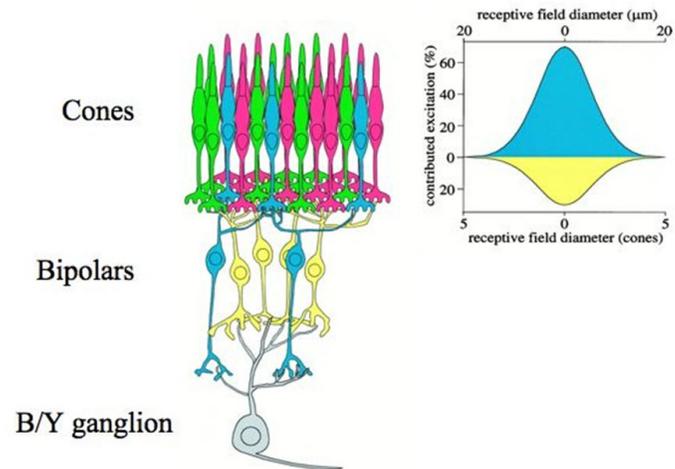
blue-yellow
bistratified
ganglion cell

From Rodieck (1998)

Koniocellular

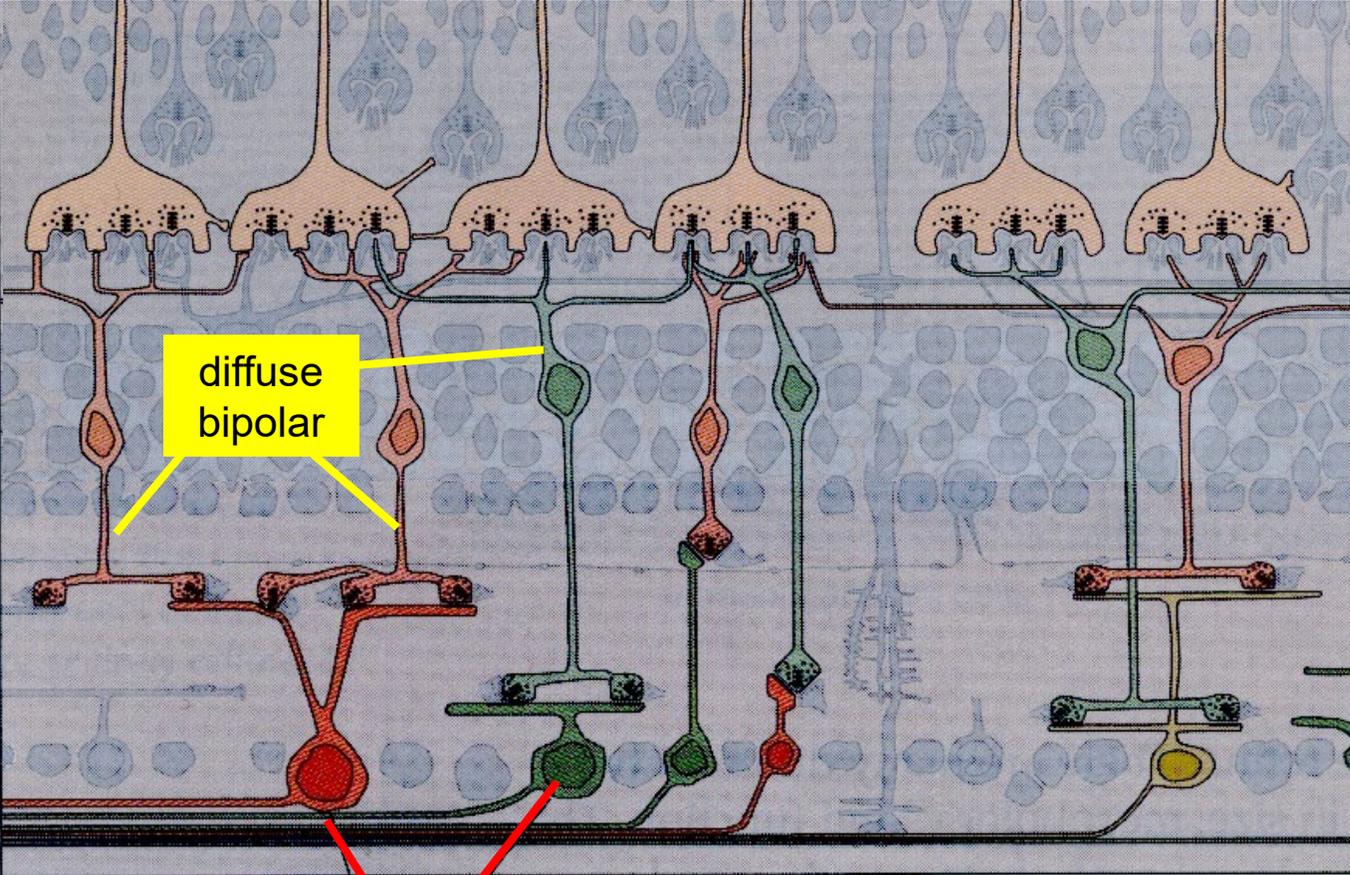


Blue/yellow pathway



Luminance pathways, which produce achromatic percepts, have been linked to the magnocellular stream.

Magnocellular

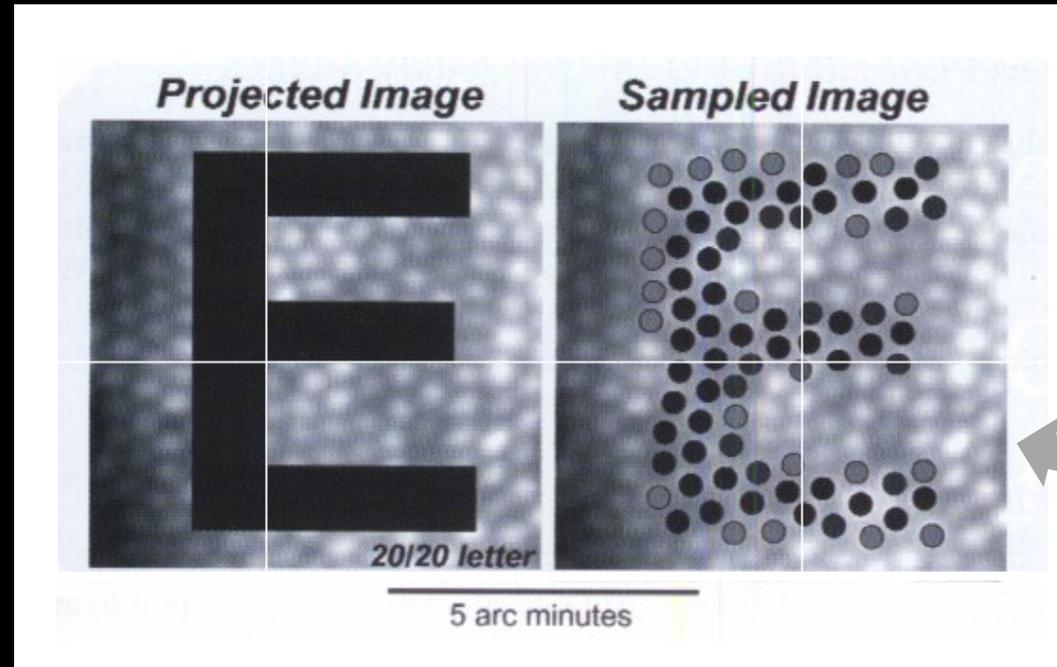


parasol
ganglion cells

From Rodieck (1998)

But the luminance pathways must be made of more than just the magnocellular stream.

Why? Consider spatial acuity...



Austin Roorda, 2004

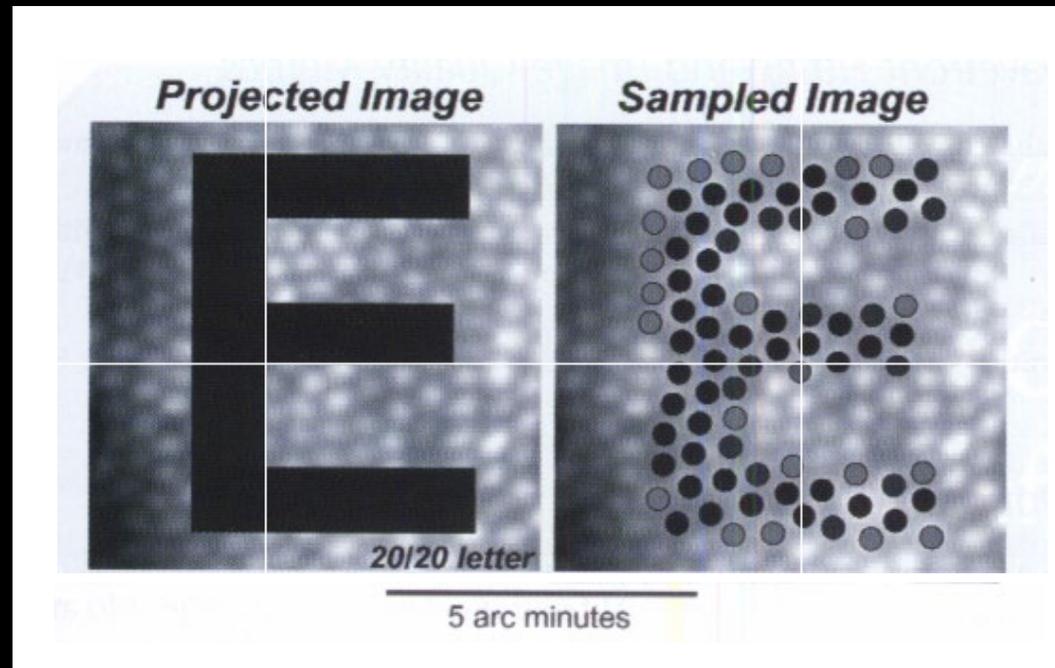
CONE ARRAY
ON RETINA

To be able to resolve this E, the image must be sampled at enough points.

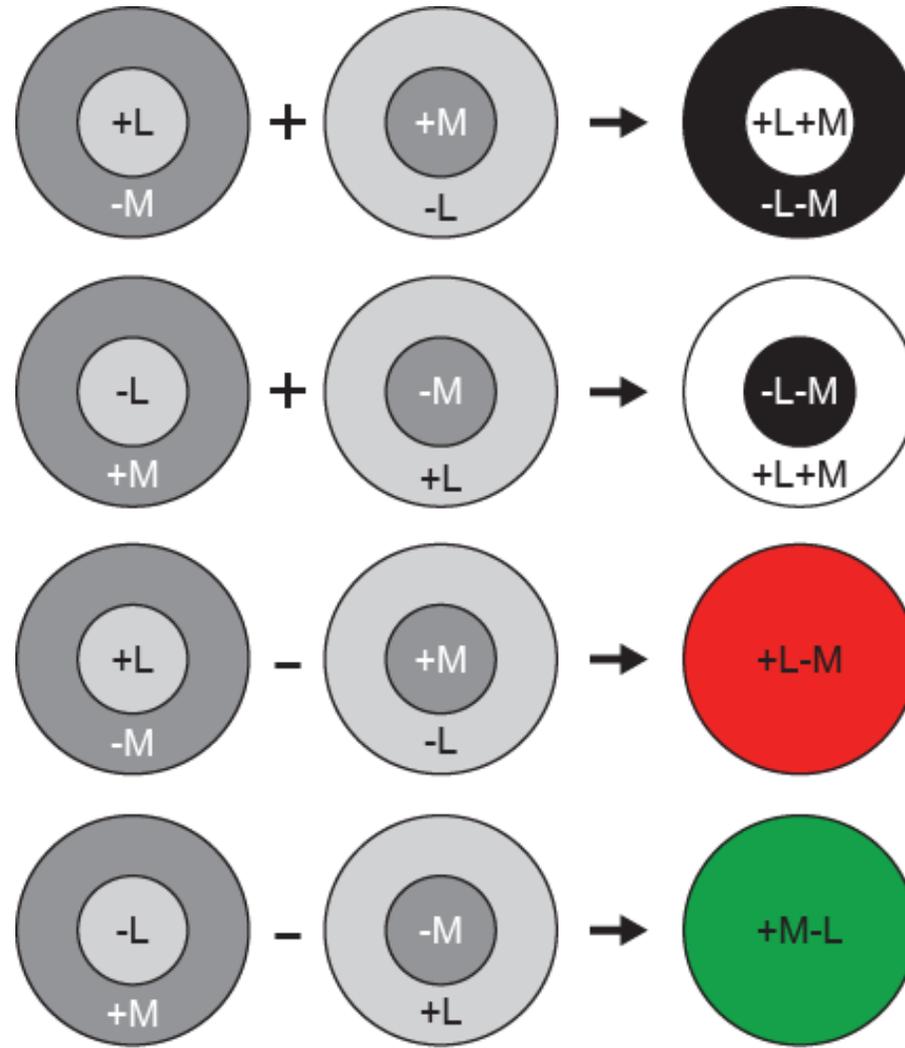
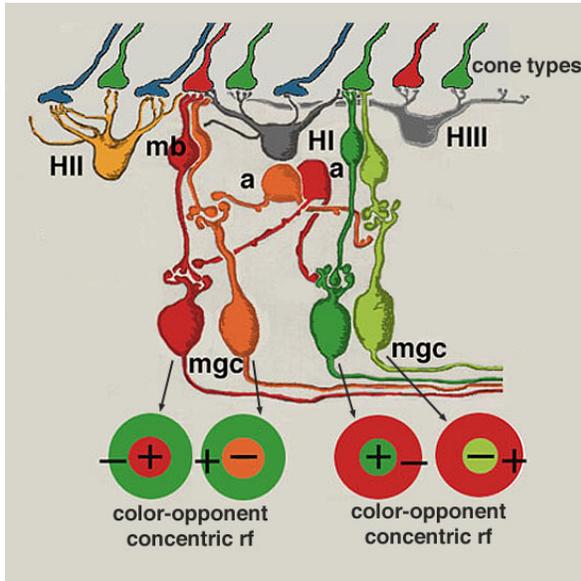
The parvocellular pathway, with its one-to-one cone to bipolar connections, provides enough samples.

The magnocellular pathway, with diffuse bipolar cells and many-to-one cone to bipolar connections, does not.

The parvocellular pathway must be double-duty supporting finely detailed luminance vision as well as more coarsely colour vision.



Colour and luminance information are “multiplexed” in the parvocellular pathway



- Chromatic pathways, which produce chromatic percepts, have been linked to the parvocellular retinal stream.
- Luminance pathways, which produce achromatic percepts, have been linked to the magnocellular stream, but *also* depend on the parvocellular stream.

Parvocellular pathway:

- High spatial frequencies (spatial detail)
- Low temporal frequencies
- Chromatic
- Lower contrast sensitivity

Magnocellular pathway:

- High temporal frequencies (motion/flicker)
- Low spatial frequencies
- Achromatic
- Higher contrast sensitivity

